<table>
<thead>
<tr>
<th>Title</th>
<th>Genome mining for radical SAM protein determinants reveals multiple sactibiotic-like gene clusters</th>
</tr>
</thead>
<tbody>
<tr>
<td>Author(s)</td>
<td>Murphy, Kiera; O'Sullivan, Orla; Rea, Mary C.; Cotter, Paul D.; Ross, R. Paul; Hill, Colin</td>
</tr>
<tr>
<td>Editor(s)</td>
<td>Read, Timothy D.</td>
</tr>
<tr>
<td>Publication date</td>
<td>2011-07-08</td>
</tr>
<tr>
<td>Type of publication</td>
<td>Article (peer-reviewed)</td>
</tr>
</tbody>
</table>
| Link to publisher's version | http://www.plosone.org/article/info%3Adoi%2F10.1371%2Fjournal.pone.0020852  
http://dx.doi.org/10.1371/journal.pone.0020852 |
| Rights | © 2011 Murphy et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. http://creativecommons.org/licenses/by/2.5/ |
| Item downloaded from | http://hdl.handle.net/10468/824 |

Downloaded on 2019-04-12T16:33:26Z
Genome Mining for Radical SAM Protein Determinants Reveals Multiple Sactibiotic-Like Gene Clusters

Kiera Murphy1,2, Orla O’Sullivan2,3, Mary C. Rea2,3, Paul D. Cotter2,3, R. Paul Ross2,3, Colin Hill1,3

1 Microbiology Department, University College Cork, Cork, Ireland, 2 Teagasc Food Research Centre, Moorepark, Fermoy, County Cork, Ireland, 3 Alimentary Pharmabiotic Centre, Cork, Ireland

Abstract

Thuricin CD is a two-component bacteriocin produced by Bacillus thuringiensis that kills a wide range of clinically significant Clostridium difficile. This bacteriocin has recently been characterized and consists of two distinct peptides, Trnβ and Trnα, which both possess 3 intrapeptide sulphur to α-carbon bridges and act synergistically. Indeed,thuricin CD and subtilosin A are the only antimicrobials known to possess these unusual structures and are known as the sactibiotics (sulphur to alpha carbon-containing antibiotics). Analysis of thethuricin CD-associated gene cluster revealed the presence of genes encoding two highly unusual SAM proteins (TrnC and TrnD) which are proposed to be responsible for these unusual post-translational modifications. On the basis of the frequently high conservation among enzymes responsible for the post-translational modification of specific antimicrobials, we performed an in silico screen for novel thuricin CD-like gene clusters using the TrnC and TrnD radical SAM proteins as driver sequences to perform an initial homology search against the complete non-redundant database. Fifteen novel thuricin CD-like gene clusters were identified, based on the presence of TrnC and TrnD homologues in the context of neighbouring genes encoding potential bacteriocin structural peptides. Moreover, metagenomic analysis revealed that TrnC or TrnD homologs are present in a variety of metagenomic environments, suggesting a widespread distribution of thuricin-like operons in a variety of environments. In-silico analysis of radical SAM proteins is sufficient to identify novel putative sactibiotic clusters.

Introduction

The ever-increasing issue of bacterial resistance to conventional antibiotics and consumer demands for safe minimally processed foods have stimulated research interest in natural antimicrobial agents such as bacteriocins [1,2,3]. A variety of approaches can be employed to identify novel bacteriocins with the most traditional being the isolation of strains and a culture-based assessment of their ability to produce novel antimicrobials which have a broad antimicrobial spectra. In contrast, the discovery of thuricin CD was the result of the specific mining of the gut microbiota for potentially therapeutic bacteriocins with a narrow spectrum of activity [4]. This alternative approach stems from the fact that in the clinical setting the use of broad spectrum antimicrobials has lead to the emergence of multi-drug resistant pathogens and the observation that the disruption of the colonic microbiota through encapsulation technologies to ensure the delivery of biologically active peptides to the colon.

Both Trnα and Trnβ are post-translationally modified peptides which contain unusual sulphur to α-carbon linkages, and as a result thuricin CD has been classified as a sactibiotic (sulphur to alpha carbon antibiotic) [9]. The thuricin CD-associated gene cluster has been identified and contains genes, tmC and tmD, predicted to encode two radical S’-adenosylmethionine (SAM) proteins. Radical SAM proteins are characteristically associated with unusual posttranslational modifications and are likely to be involved in the formation of the aforementioned cysteine to α-carbon linkages [10]. Radical SAM-encoding genes are rare in bacteriocin-associated clusters, with subtilosin A and propionicin F being notable exceptions [11,12]. Propionicin F is not a modified peptide and in this instance it is thought that the associated radical SAM enzyme contributes to N-terminal cleavage [12]. However, subtilosin A is a cyclic peptide which undergoes posttranslational modification involving a thiol linkage of sulphur to α-carbon and can therefore be classified as a sactibiotic. The mechanism associated diarrhoea (CDAD) including the hypervirulent strain B1/NAP1/027 [7]. In a distal colon model thuricin CD, while killing C. difficile, had no significant impact on the composition of the microbiota unlike other broad spectrum antimicrobials [8]. However to be used as an oral therapeutic for CDAD thuricin CD, because it is sensitive to proteolytic enzymes, will require the use of
associated with the formation of this unusual posttranslationally modified modification has not been established but is thought to be due to the associated radical SAM enzyme, AlbA.

An alternative approach to the identification of novel posttranslationally modified peptides has involved in silico screening to identify the most highly conserved component of the associated gene clusters i.e. the genes encoding the modification enzymes. This approach has been employed, for example, to identify novel type I and type II lantibiotics [13,14,15,16] and thiazole/oxazole-modified microcin (TOMMS; [17,18,19,20,21]). Here this approach has been adapted to identify novel thuricin CD-like gene clusters. More specifically, an in silico screen of databases was undertaken, using TrnC and TrnD as driver sequences, to identify similar gene clusters. The search revealed 100 TrnC homologs and 53 TrnD homologs, which upon further investigation led to the identification of 15 novel putative thuricin CD-like clusters, i.e. clusters containing genes encoding TrnC and TrnD homologs as well as at least one structural peptide, across three phyla.

Results

In silico screen for TrnC and TrnD Protein

The sequences of TrnC and TrnD, radical SAM proteins encoded within the thuricin CD gene cluster [4], were used as driver sequences to perform a homology search against the complete non-redundant database (November 2010). The search revealed 100 TrnC homologs and 54 TrnD homologs (homology being defined as BLASTP E-value < 10^-5), in 112 unique genomes. Tables S1 and S2 provide a full list of TrnC and TrnD homologs, which were found in the genomes of sequenced microorganisms. In each case the corresponding bacterial genomes were viewed and the regions containing the genes of interest were located. None of the strains identified have previously been associated with a bacteriocin producing phenotype.

A more detailed examination of these gene clusters involved an analysis of the regions flanking the trnC and trnD determinants for other open reading frames (ORFs) potentially involved in the biosynthesis of, or immunity to, bacteriocins. The presence of a second nearby Radical SAM gene (including those encoding another TrnC or TrnD homolog) was screened for in each case. Putative Radical SAM proteins were examined for the presence of signature motifs. Enzymes of the Radical SAM superfamily generate radicals by combining a 4Fe-4S cluster and S-adenosylmethionine (SAM) in close proximity. The Radical SAM superfamily motifs. Enzymes of the Radical SAM superfamily are grouped according to the phylum and genus of the associated gene cluster in Firmicutes.

Following this screen, 15 gene clusters from 15 different bacterial strains were deemed worthy of closer analysis. Of these, 10 were gene clusters that contained both TrnC and TrnD encoding genes with putative prepeptide-encoding genes. The remaining 5 contained either a TrnC or a TrnD encoding gene but were deemed noteworthy as the majority of the other thuricin CD operon-associated features were present. Thirteen of these 15 total gene clusters were in the genomes of strains within the phylum Firmicutes. The other 2 strains were Bacteroideidae sp. 3_1_19 of the phylum Bacteroidetes and Petrotoga mobilis SJ95 of the phylum Thermotogae. These 15 clusters are described below and are grouped according to the phylum and genus of the associated strain. Arrow diagrams representing each of these clusters are depicted in Figure 1.

Gene clusters in Firmicutes

Identification of novel Bacillus-associated bacteriocin gene clusters. Bacillus is a large and diverse genus which contains both free living and pathogenic species. Members are Gram-positive, rod-shaped, sporulating and are known to produce a wide range of antimicrobial substances including bacteriocins. Many of the Bacillus bacteriocins are lanthionines, a category of post-translationally modified peptides which are grouped as a consequence of the presence of (B-methyl) lantionine structures. Examples include subtilin [25], ericin S and ericin A [26], haloduracin [14,16] and lichenicidin [13]. Our in silico screen highlighted the presence of thuricin CD-like gene clusters in the genomes of three Bacillus strains, i.e. B. cereus 95/8201, B. thuringiensis serovar pulsatilis BGSC 4CC1 and B. thuringiensis serovar hauzhuangensis BGSC 4BD1.

B. cereus 95/8201. Strains of B. cereus are frequently isolated from soil but also found in foods of plant and animal origin. B. cereus is an opportunistic pathogen and has frequently been associated with human food poisoning [27].

B. cereus strain 95/8201 (GenBank accession ACMF00000000) was isolated from a human patient with endocarditis (Bacillus cereus group Tourasse-Helgason MLST Database) and sequenced by the Naval Medical Research Centre in 2009. A gene cluster was identified in this strain which is very similar to the thuricin CD gene cluster in B. thuringiensis DPC 6431. Bcere0016_53380 (499 residues) resembles TrnC [506 amino acids]. The C-X2-C_X2-C motif occurs as CNLRCY at Cys107 in Bcere0016_53380 and TAAAGGAGGAATTATC). Also screened for was the presence of conserved cysteine residues such as those that occur at positions +5, +9 and +13 in TrnC and TrnB, forming a CX3CX3C formation. Cys +5 is flanked by small hydrophobic amino acids Ala and Val, and those at positions +9 and +13 are coupled with variant small hydrophobic residues. A conserved Ser residue is present at position +13. Additionally, Gly residues occur at positions +17 and +19, which are followed by variant hydrophilic and hydrophobic residues, respectively [4].

Sequencing had revealed that TrnC and TrnB are posttranslationally modified at their respective 21st, 25th and 29th residues [4]. It is believed that each peptide has three sulphur to α-carbon thioether linkages between the cysteine side chains and the α-carbons of the modified residues. These unusual sulphur to α-carbon linkages have also been reported in the antimicrobial subtilosin A [24]. Notably, a SAM-containing enzyme is thought to be responsible for forming the thioether linkages in subtilosin A [24]. This protein bears some resemblance to the SAM-containing enzymes encoded within the thuricin CD operon [4]. Other features that were specifically screened for were the presence of other genes predicted to encode bacteriocin associated proteins, including those involved in transport and processing.

Following this screen, 15 gene clusters from 15 different bacterial strains were deemed worthy of closer analysis. Of these, 10 were gene clusters that contained both TrnC and TrnD encoding genes with putative prepeptide-encoding genes. The remaining 5 contained either a TrnC or a TrnD encoding gene but were deemed noteworthy as the majority of the other thuricin CD operon-associated features were present. Thirteen of these 15 total gene clusters were in the genomes of strains within the phylum Firmicutes. The other 2 strains were Bacteroideidae sp. 3_1_19 of the phylum Bacteroidetes and Petrotoga mobilis SJ95 of the phylum Thermotogae. These 15 clusters are described below and are grouped according to the phylum and genus of the associated strain. Arrow diagrams representing each of these clusters are depicted in Figure 1.

Gene clusters in Firmicutes

Identification of novel Bacillus-associated bacteriocin gene clusters. Bacillus is a large and diverse genus which contains both free living and pathogenic species. Members are Gram-positive, rod-shaped, sporulating and are known to produce a wide range of antimicrobial substances including bacteriocins. Many of the Bacillus bacteriocins are lantibiotics, a category of post-translationally modified peptides which are grouped as a consequence of the presence of (B-methyl) lantionine structures. Examples include subtilin [25], ericin S and ericin A [26], haloduracin [14,16] and lichenicidin [13]. Our in silico screen highlighted the presence of thuricin CD-like gene clusters in the genomes of three Bacillus strains, i.e. B. cereus 95/8201, B. thuringiensis serovar pulsatilis BGSC 4CC1 and B. thuringiensis serovar hauzhuangensis BGSC 4BD1.

B. cereus 95/8201. Strains of B. cereus are frequently isolated from soil but also found in foods of plant and animal origin. B. cereus is an opportunistic pathogen and has frequently been associated with human food poisoning [27].

B. cereus strain 95/8201 (GenBank accession ACMF00000000) was isolated from a human patient with endocarditis (Bacillus cereus group Tourasse-Helgason MLST Database) and sequenced by the Naval Medical Research Centre in 2009. A gene cluster was identified in this strain which is very similar to the thuricin CD gene cluster in B. thuringiensis DPC 6431. Bcere0016_53380 (499 residues) resembles TrnC [506 amino acids]. The C-X2-C_X2-C motif occurs as CNLRCY at Cys107 in Bcere0016_53380 and TAAAGGAGGAATTATC). Also screened for was the presence of conserved cysteine residues such as those that occur at positions +5, +9 and +13 in TrnC and TrnB, forming a CX3CX3C formation. Cys +5 is flanked by small hydrophobic amino acids Ala and Val, and those at positions +9 and +13 are coupled with variant small hydrophobic residues. A conserved Ser residue is present at position +13. Additionally, Gly residues occur at positions +17 and +19, which are followed by variant hydrophilic and hydrophobic residues, respectively [4].

Sequencing had revealed that TrnC and TrnB are posttranslationally modified at their respective 21st, 25th and 29th residues [4]. It is believed that each peptide has three sulphur to α-carbon thioether linkages between the cysteine side chains and the α-carbons of the modified residues. These unusual sulphur to α-carbon linkages have also been reported in the antimicrobial subtilosin A [24]. Notably, a SAM-containing enzyme is thought to be responsible for forming the thioether linkages in subtilosin A [24]. This protein bears some resemblance to the SAM-containing enzymes encoded within the thuricin CD operon [4]. Other features that were specifically screened for were the presence of other genes predicted to encode bacteriocin associated proteins, including those involved in transport and processing.

Following this screen, 15 gene clusters from 15 different bacterial strains were deemed worthy of closer analysis. Of these, 10 were gene clusters that contained both TrnC and TrnD encoding genes with putative prepeptide-encoding genes. The remaining 5 contained either a TrnC or a TrnD encoding gene but were deemed noteworthy as the majority of the other thuricin CD operon-associated features were present. Thirteen of these 15 total gene clusters were in the genomes of strains within the phylum Firmicutes. The other 2 strains were Bacteroideidae sp. 3_1_19 of the phylum Bacteroidetes and Petrotoga mobilis SJ95 of the phylum Thermotogae. These 15 clusters are described below and are grouped according to the phylum and genus of the associated strain. Arrow diagrams representing each of these clusters are depicted in Figure 1.
is identical to the corresponding region in TrnC. Putative SAM binding sequence motifs, GGEPLL (identifies at Gly163 and TNG (identified at Thr195) are also conserved between Bcere0016_53390 and TrnC. Bcere0016_53390 (409 amino acids) resembles TrnD (414 residues). The signature motif occurs as CMNMCCKH (at Cys103 in Bcere0016_53390) in both. The GGEPLL (Gly150) and TNG (T187) motifs in Bcere0016_53390 are identical to those in TrnD. Bcere0016_53360 and Bcere0016_53370 are the predicted structural genes and are 100% identical to B. thuringiensis DPC 6431 structural genes, trnB and trnX respectively. Other genes of note in this cluster are those encoding a putative ABC transporter (Bcere0016_53350) and a C-terminal protease (Bcere0016_53400). The ABC transporter may be responsible for the export of the bacteriocin from the cytoplasm and the peptide may have a role in immunity or leader cleavage.

B. thuringiensis serovar puusilusius BGSC 4CC1. B. thuringiensis are noted for their ability to produce a parasporal crystal upon sporulation. Due to its insecticidal properties it has been used commercially as a biological insecticide [28]. B. thuringiensis serovar puusilusius BGSC 4CC1 (Accession NZ_ACN000000000) was sequenced by the Naval Medical Research Centre in 2009. Within this genome, putative TrnC (Bthur0012_58010) and TrnD (Bthur0012_58000) encoding genes were identified which are surrounded by other genes associated with bacteriocin synthesis. The TrnC homolog (e-value 3e-30) encoded by Bthur0012_58010 contains the GGEPLL motif at Gly35 while the TrnD homolog (e-value 8e-23) encoded by Bthur0012_58000 has a GGDPLL at Gly167 and a C-X3-C-X2-C motif (CNKNCIHC) at Cys116. The TrnC homolog (e-value 5e-30) encoded by Bthur0012_58010 has a GGEPLL motif at Gly35 while the TrnD homolog (e-value 2e-23), encoded by Bthur0012_58000 are identical to those in TrnC. Bthur0012_58010 and Bthur0012_58000 are the predicted structural genes and are 100% identical to B. thuringiensis DPC 6431 structural genes, trnB and trnX respectively. Other genes of note in this cluster are those encoding a putative ABC transporter (Bthur0016_53350) and a C-terminal protease (Bthur0016_53400). The ABC transporter may be responsible for the export of the bacteriocin from the cytoplasm and the peptide may have a role in immunity or leader cleavage.

B. thuringiensis serovar haushongenius BGSC 4BD1. B. thuringiensis serovar haushongenius BGSC 4BD1 (Accession NZ_ACN000000000) was also sequenced by the Naval Medical Research Centre in 2009. The thuricin CD-like gene cluster identified here is very similar to those found in the other Bacillus species listed above. The BGSC 4BD1-associated cluster contains two putative Radical SAM encoding genes, Bthur0011_60810 and Bthur0011_60830. Bthur0011_60820 encodes a TrnC homolog (e-value 6e-30), contains 359 amino acids and a GGEPLL motif at Gly35. The TrnD homolog (e-value 2e-23), encoded by Bthur0011_60830, is 409 amino acids, has a GGDPLL motif at Gly167 and a TNG motif at Thr195. Interestingly, the two predicted structural genes, Bthur0011_60810 and Bthur0011_60830 are identical to those found in B. thuringiensis serovar puusilusius BGSC 4CC1 described above. Adjacent genes potentially of relevance are predicted to encode an ABC-type transporter (Bthur0011_60840) and a permease (Bthur0011_60850).

Identification of novel Clostridium- and Clostridia-associated bacteriocin gene clusters

Clostridium (family Clostridaceae) is a genus of gram positive, anaerobic, endospore-forming, Firmicutes, of which there are approximately 100 species. These include important pathogens such as C. difficile, C. perfringens and C. tetani. Several Clostridium-associated bacteriocins have been characterised from this genus, including boticin B [29] and bacteriocin 28 [30]. Bioinformatic analysis revealed the presence of thuricin CD-like clusters in genomes of the genus Clostridium, specifically within the genomes of, C. cellulolyticum H10, C. papyrosolvens DSM 2782, C. thermocellum ATCC 27405 C. difficile QCD-634e2, C. difficile 630 and C. difficile QCD-326g50. Four further clusters were identified within the broader class Clostridia. They also included two clusters identified in the genus Anaerococcus (family Carboxydotheraceae); Anaerococcus tetradius ATCC 35098 and Anaerococcus proteus DSM 20548. The thuricin CD-like clusters were also identified in Thermoaerobacter methanomethylicum A3 (family Thermoaerobacteraceae) and Caldicellulosiruptor kristjanssonii 177R1B (family Thermoaerobacteraceae).

C. cellulolyticum H10. C. cellulolyticum H10 (Accession WP_002504697) is a mesophilic cellulolytic bacterium that was originally isolated from decayed grass compost [31]. The ability of this species to degrade cellulose is an active area of interest as the products of this degradation, ethanol and hydrogen, may be used as an alternative form of energy [32]. The C. cellulolyticum H10 sequence was published in 2009 by US DOE Joint Genome Institute. Within the thuricin CD-like cluster of this strain, Ccel_0328 encodes an apparent TrnC homolog (e-value 7e-30). This 486 amino acid protein contains a GGEPLL motif at Gly145, a TNG motif at Thr177 and a C-X3-C-X2-C motif (CNKNCIHC) at Cys94. Ccel_0328 was also predicted to encode a TrnD homolog (e-value 2e-6). An adjacent gene also expressed low level homology with TrnD (e-value 0.81). Two possible structural genes, Ccel_0331 and Ccel_0332, were predicted. These are predicted to encode peptides 32 amino acids and 63 amino acids long, respectively. Both contain GG motifs and cysteine residues, albeit not in the C-X3-C-X2-C motif formation. Other adjacent genes are predicted to encode a glycosyl transferase, Ccel_0333 and an A-type ATPase (ATPas associated with diverse cellular activities), Ccel_0327.

C. papyrosolvens DSM 2782. C. papyrosolvens DSM 2782 (Accession NZ_ACX000000000) was isolated from intertidal mud from the Don River in Scotland [33] and sequenced in 2009 by the US DOE Joint Genome Institute. The gene cluster of note examined here contains a putative TrnC (482 amino acid) encoding gene, CcapDRAFT_0066 (e-value 4e-28). CcapDRAFT_0066 has a GGEPLL motif at Gly154 and a C-X3-C-X2-C (CNLRCKY) motif at Cys103. The protein (375 amino acids) encoded by the adjacent gene bears some slight resemblance to TrnD (e-value 2.8). CcapDRAFT_0064 may encode a structural peptide with a GG motif at Gly78. Other bacteriocin-associated genes include those encoding a dipetidase (CcapDRAFT_0068) and two branched chain amino acid transporters (CcapDRAFT_0062 and CcapDRAFT_0063).

C. thermocellum ATCC 27405. C. thermocellum is a thermophilic, cellulolytic bacterium capable of directly converting cellulosic substrate into ethanol. The ATCC 27405 strain (Accession NC_009012) was sequenced in 2007 by the US DOE Joint Genome Institute. While this particular gene cluster only contains one Radical SAM protein and one putative structural peptide encoding gene, the high similarity of their sequences to those found in the other clusters is noteworthy. As a consequence the cluster was the focus of closer inspection. The Radical SAM-encoding gene identified, Cthc_0906, resembles both TrnC (e-value 3e-16) and TrnD (e-value 3e-5). This protein contains 450 amino acids, a GGEPPM motif at Gly153 and a TNG motif at Thr187. A
C-X₃-C-X₂-C motif, namely CNLRCKYC, is identified at Cys104. Interestingly, the CNLRCKYC sequence was also observed in Calkr_0222, a TrnC homolog in *C. kristjanssonii* 177R18. Cthe_0907 is believed to encode a structural gene. It is 46 amino acids long, contains a GG motif at Gly19 and a C-X₃-C-X₃-C motif (CQTSCQyAC) at Cys24. Within this cluster other genes potentially of relevance include *secD*, a gene involved in protein export (Cthe_0904) and also a signal peptidase-encoding gene (Cthe_0909).

**C. difficile QCD-63q42.** *C. difficile* is the leading cause of nosocomial infections associated with antibiotic use and is responsible for substantial morbidity and mortality worldwide [34]. *C. difficile* QCD-63q42 is one of three *C. difficile* strains in which thuricin CD-like gene clusters were identified in this study. *C. difficile* QCD-63q42 (Accession NZ_ABHD00000000) was isolated from a 67 year old male patient with severe *C. difficile* Associated Disease (CDAD) and was sequenced in 2007 by McGill University. Like the aforementioned *C. thermocellum* ATCC 27405 gene cluster, the QCD-63q42 gene cluster is lacking a TrnD homolog and only has one structural gene but was included for the same reasons. The TrnC homolog encoded by CdiQCD-6_020200015426 (e-value 4e-21) contains 475 amino acids, a GGEPLL motif at Gly148 and the C-X₃-C-X₂-C motif (CNLHCDYC) at Cys97. The proposed structural peptide, encoded by CdiQCD-6_020200015431, is larger than those previously observed (79 amino acids) but does contain two GG motifs, at Gly 21 and Gly 39, and also a C-X₃-C-X₃-C motif, CGALCANLC, at Cys25. Nearby genes include those encoding a hydrolase (CdiQCD-6_020200015426) and an ABC transporter (CdiQCD-6_020200015451).

**C. difficile 630.** The complete genome sequence of *C. difficile* strain 630 (Accession NC_009009) was sequenced in 2006 by the Sanger Institute [34]. The strain was isolated from a patient with severe pseudomembranous colitis in Switzerland and is a virulent, highly transmissible, drug resistant strain. CD0162 is a homolog of both TrnC and TrnD (e-values of 5e-30 and 2e-0, respectively). While the adjacent CD0163 was annotated as hypothetical, alignment of its sequence with TrnD confirmed it is a TrnD homolog (e-value 1e-12). Both CD0162 (473 amino acids) and CD0163 (356 amino acids) lack the signature C-X₃-C-X₂-C motif but both possess a GG motif.

CD0163B is thought to encode a potential structural gene of 67 amino acids containing a GG motif at Gly47 and a C-X₃-C-X₃-C motif (CTIMCPYTC) at Cys20. The gene cluster also contains genes that code for an ABC transporter (CD0161) and a two-component response regulator (CD0139 and CD0160).

**C. difficile QCD-32g58.** *C. difficile* QCD-32g58 (Accession NZ_AAML00000000) was sequenced in 2007 by Washington...
University. This hyper-virulent strain was responsible for a multi-institutional outbreak of CDAD in Quebec, Canada [35]. Analysis of this genome revealed the presence of a TrnC homolog-encoding gene, CdfIQ_0405000000917 (e-value 6e-30). The corresponding protein contains a GGEPILL motif at Gly153 and a C-X3-C-X2-C motif, CNLRCDCYC. As is the case with some of the other Clostridia-associated clusters strains examined, an adjacent gene, CdfIQ_0405000000922, was found to encode a protein (356 amino acids) that does not significantly resemble TrnD (e-value 4.5). A screen for potential structural genes revealed the presence of three adjacent genes of relevant size. Upon further analysis, the 67 amino acid CdfIQ_0405000000932 was determined to be most likely to be the structural peptide-encoding gene as its product is predicted to contain a GG motif at Gly47 and a C-X3-C-X3-C motif, CTIMCPYTC at Cys20. Surrounding genes of note include those encoding an ABC transporter (CdfIQ_0405000000912) and a response regulator (CdfIQ_040500000907).

**Anaerococcus tetradius ATCC 35098.** *A. tetradius* is a strictly anaerobic bacterium, usually found in fresh water and hot springs. Lantibiotics have previously been associated with related microbes, such as ruminicocin A produced by *Ruminococcus gnavus* [36]. *A. tetradius* ATCC 35098 (Accession ACGC000000000) was sequenced in 2009 by Baylor College of Medicine. The gene cluster of interest within this genome may contain as many as 11 genes. BLAST searches revealed that the proteins encoded by HMPREF0077_1416 (e-value 3e-17) and HMPREF0077_1424 (e-value 3e-23) are TrnC homologs. HMPREF0077_1416 (e-value 3e-17) and HMPREF0077_1424 (e-value 3e-23) are TrnC homologs mentioned previously. Tmath_1976 is 453 amino acids long and contains a putative bacteriocin-associated genes. BLAST searches revealed that the proteins encoded by HMPREF0077_1416 (e-value 3e-17) and HMPREF0077_1424 (e-value 3e-23) are TrnC homologs. HMPREF0077_1416 contains 323 amino acids was examined. It contained a C-X3-C-X3-C motif, CNLKCDYC. As is the case with some of the other Tmath_1976-associated clusters strains examined, an adjacent gene, Tmath_1977, was also identified. The corresponding peptide is predicted to contain 32 residues, a GG motif at Gly14 and several cysteine residues. The product of Tmath_1977 contains 461 residues, a C-X3-C-X3-C motif, namely CNLRCDCYC, at Cys104, which has been observed in other TrnC homologs mentioned previously. Tmath_1976 is 453 amino acids long and contains a GTEPILL motif at Gly153, a TNG at Thr117 and a C-X3-C-X3-C motif, CNLRCDCYC, at Cys104 and has 457 amino acids. Another protein annotated as a Radical SAM, HMPREF0077_1421, and containing 323 amino acids was examined. It contained a C-X3-C-X3-C motif, CNFSCSLHC, at Cys18 but lacked GGEPILL and TNG motifs. A sequence alignment with TrnD showed a good agreement and gave an e value of 4e-4. While this is outside the threshold set at e-value <10^-6, its presence remains noteworthy. In addition to the predicted radical SAM proteins, a hypothetical protein associated with this cluster shares homology with a methylase (HMPREF0077_1423). Methylases contribute to the production of a number of non-ribosomal antibiotics [37]. HMPREF0077_1413 and HMPREF0077_1414 encode homologues of the SaliRK two component signal transduction system associated with the production of the lantibiotic salivaricin A2, produced by *Streptococcus salivarius* K12 [38]. In close proximity are genes encoding a response regulator (HMPREF0077_1414) and two ABC transporters (HMPREF0077_1418 and HMPREF0077_1426).

**A. prevoti DSM 20548.** *A. prevoti* is an anaerobic, mesophilic, non-motile, non-sporulating bacterium. It is an opportunistic pathogen found in the microflora of the mouth, skin or vagina of the host [39]. Strain DSM 20548 (Accession CP001709) was isolated from a human plasma sample and was sequenced in 2009 by the US DOE Joint Genome Institute. It contains a thuricin CD-like gene-cluster of interest on its plasmid pAPRE01. A putative TrnC (e-value 8e-37)-encoding gene, Aprc_1838, was identified in this cluster. The corresponding 518 amino acid protein is predicted to contain a GGEPILL motif at Gly184 and a C-X3-C-X2-C motif, CNLRCDCYC at Cys132. A CNLRCDCYC motif was also observed in TrnC homologs encoded by gene clusters in *C. kristjanssonii* 177R1B, *C. papyrosolvens* DSM 2782 and *C. thermocellum* ATCC 27405. A hypothetical gene, Aprc_1836, adjacent to the putative TrnC, of 426 amino acids was aligned with TrnD but no significant similarity was revealed. There are two potential structural genes, Aprc_1838 and Aprc_1839. Aprc_1838 encodes a 65 AA protein, with a GA site at Gly14 and several cysteine residues. The product of Aprc_1839 has a GG motif at Gly182 and C-terminal cysteine but is much larger than typical bacteriocin pre-peptides, consisting of 409 amino acids. The flanking region also contains genes encoding a histidine kinase (Apre_1834) and two ABC transporters (Apre_1835 and Apre_1840).

**Thermoanaerobacter mathranii A3.** *T. mathranii* A3 (Accession NC_014721) is a rod-shaped, motile, ethanol-producing, extremely thermophilic, anaerobic bacterium isolated from a hot spring in Iceland [40]. This strain was recently sequenced in 2010 by the US DOE Joint Genome Institute and contains a putative bacteriocin gene cluster. The *T. mathranii* A3 genome contains two radical SAM-encoding genes, the TrnC homolog-encoding Tmath_1971 (e value 3e-10) and the TrnD homolog-encoding Tmath_1976 (e value 3e-6). Tmath_1971 contains 461 residues, a GGEPILL at Gly153 and interestingly a C-X3-C-X3-C motif, namely CNLRCDCYC, at Cys104, which has been observed in other TrnC homologs mentioned previously. Tmath_1976 is 453 amino acids long and contains a GTEPILL motif at Gly151, a TNG at Thr185 and a C-X3-C-X3-C motif, CNLRCDCYC, at Cys101. A putative structural gene, Tmath_1977, was also identified. The corresponding peptide is predicted to contain 32 residues, a GG motif at Gly14 and two C-terminal cysteine residues. Other surrounding genes of note include those encoding two ABC transporters (Tmath_1974 and Tmath_1975) and *spaD*, a gene involved in protein export (Tmath_1975).

**Caldicellulosiruptor kristjanssonii 177R1B.** *Caldicellulosiruptor kristjanssonii* 177R1B (Accession NC_014721) was isolated from a hot spring biomat in Iceland and sequenced in 2010 by the

![Table 1. Highly conserved residues shared by trnD and 10 homologs.](https://www.plosone.org/doi/supplementary/10.1371/journal.pone.0020852.s001)
Figure 3. Protein sequence alignment of TrnC and all the TrnC homologs identified in this study.
doi:10.1371/journal.pone.0020852.g003
US DOE Joint Genome Institute. The 177R1Bthuricin CD-like gene cluster contains a number of thuricin CD-associated features. Calkr_0222 encodes a 492 amino acid TrnC homolog (e-value of 5e-77) that contains a GGEPLL motif at Gly172, a TNG motif at Thr193 and a C-X3-C- X2-C motif (CNLQCRYC) at Cys114. The TrnD homolog, encoded by Calkr_0223, contains 421 amino acids (e-value of 7e-61). A GGEPLL motif at Gly154, a TNG motif at Thr193 and a C-X3-C- X2-C motif (CNLQCRYC) at Cys110 were identified. Two possible structural peptides, encoded by Calkr_0221 and Calkr_0220, contain features associated with Trnβ and Trnξ. Calkr_0221 contains 41 residues, a GG motif at Gly14 and a C-X3-C- X2-C motif (CFWICGSFC) at Cys19. Calkr_0220 is 45 amino acids long and contains two possible GG cleavage sites at Gly14 and Gly25 and also a C-X3-C- X2-C motif (CLIALCVCYGC) at Cys19. There are also adjacent peptidase- (Calkr_0224) and ABC transporter- (Calkr_0226) encoding genes.

Gene clusters in non-Firmicutes

The final two thuricin CD-like gene clusters of note were identified in strains that were non Firmicutes. i.e. *Petrotoga mobilis* SJ95 of the phylum Thermotogae phylum and *Bacteroides* sp. 3_1_19 of the phylum Bacteroidetes. These are described below. *Petrotoga mobilis* SJ95 (Accession NC_010003) is a rod-shaped, gram-negative, thermophilic bacterium from the order Thermotogales. It is able to reduce elemental sulphur to hydrogen sulfide and was isolated from the hot oilfield water of a North Sea oil reservoir [42]. There have been no reports to date of bacteriocin production by this species. Strain SJ95 was sequenced in 2007 by the US DOE Joint Genome Institute. A gene cluster of note was identified in this strain, which contains a number of potential bacteriocin-associated genes in close proximity to putative *trnC* and *trnD* determinants, Pmob_0678 (e-value 3e-36) and Pmob_0677 (e-value 1e-18) respectively. Pmob_0678 is a 464 amino acid protein with a GGEPLL motif at Gly136, a TNG motif at Thr168 and a C-X3-C- X2-C motif, CNLQCRYC at Cys114. Pmob_0677 contains 430 residues and the only observable motifs were a GG at Gly 173 and Gly32. A putative ABC transporter, Pmob_0676, was also observed nearby. The probable structural peptide is the 197 aa long hypothetical protein Pmob_0674, with GG motifs at Gly92 and Gly332. A putative ABC transporter, Pmob_0676, was also observed nearby. The probable structural peptide is the 197 aa long hypothetical protein Pmob_0674, with GG motifs at Gly92 and Gly332. A putative ABC transporter, Pmob_0676, was also observed nearby. The probable structural peptide is the 197 aa long hypothetical protein Pmob_0674, with GG motifs at Gly92 and Gly332. A putative ABC transporter, Pmob_0676, was also observed nearby. The probable structural peptide is the 197 aa long hypothetical protein Pmob_0674, with GG motifs at Gly92 and Gly332. A putative ABC transporter, Pmob_0676, was also observed nearby.

**Table 2.** Highly conserved residues shared by trnC and 18 homologs.

<table>
<thead>
<tr>
<th>From Alignment</th>
<th>Alignment position</th>
<th>% Conservation</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Motifs</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CX3CX2C</td>
<td>202–209</td>
<td>89</td>
</tr>
<tr>
<td>GGEPLL</td>
<td>280–287</td>
<td>68</td>
</tr>
<tr>
<td>TTN</td>
<td>315–317</td>
<td>89</td>
</tr>
<tr>
<td>LL</td>
<td>320–321</td>
<td>63</td>
</tr>
<tr>
<td>SXDG</td>
<td>340–343</td>
<td>89</td>
</tr>
<tr>
<td>GXG</td>
<td>358–361</td>
<td>58</td>
</tr>
<tr>
<td>DG</td>
<td>547–548</td>
<td>63</td>
</tr>
<tr>
<td>PC</td>
<td>552–553</td>
<td>63</td>
</tr>
<tr>
<td>CE</td>
<td>553–555</td>
<td>68</td>
</tr>
<tr>
<td>GN</td>
<td>565–566</td>
<td>58</td>
</tr>
<tr>
<td>CX3CX2C</td>
<td>597–610</td>
<td>100</td>
</tr>
<tr>
<td>CWA</td>
<td>600–602</td>
<td>53</td>
</tr>
<tr>
<td><strong>Single Residues</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>D</td>
<td>44;243;349;401</td>
<td>58; 95; 79; 74</td>
</tr>
<tr>
<td>G</td>
<td>137;538;566; 573</td>
<td>58; 84; 100; 84</td>
</tr>
<tr>
<td>T</td>
<td>199</td>
<td>74</td>
</tr>
<tr>
<td>Y</td>
<td>222; 281</td>
<td>63; 68</td>
</tr>
<tr>
<td>M</td>
<td>233</td>
<td>89</td>
</tr>
<tr>
<td>S</td>
<td>254</td>
<td>68</td>
</tr>
<tr>
<td>F</td>
<td>280; 289</td>
<td>84; 74</td>
</tr>
<tr>
<td>K</td>
<td>293; 300</td>
<td>100; 58</td>
</tr>
<tr>
<td>N</td>
<td>333</td>
<td>74</td>
</tr>
<tr>
<td>E</td>
<td>324; 346; 650</td>
<td>74; 74; 63</td>
</tr>
<tr>
<td>R</td>
<td>352</td>
<td>100</td>
</tr>
<tr>
<td>C</td>
<td>535; 553; 629</td>
<td>100; 100</td>
</tr>
</tbody>
</table>

Alignment of TrnC and TrnD- like Radical SAM proteins

The availability of a significant number of TrnC- and TrnD-like radical SAM protein sequences enabled further *in silico* analysis to identify conserved motifs and residues. Alignment of TrnD and 9 homologs (Fig. 2) reveals a number of completely conserved motifs and residues, which are summarized in Table 1. A radical SAM superfamily signature motif CXC3CX2C was identified in 8 sequences. *B. thuringiensis serovar huazhongensis* BGSC 4BD1 Bthuo011_60030 and *C. difficile* 630 CD0163 did not contain this motif. Radical SAM enzymes generate catalytic radicals by
combining a 4Fe-4S cluster and S-adenosylmethionine (SAM) in close proximity. The reaction requires the input of one electron, which is supplied by the 4Fe-4S cluster that is bound to the protein via cysteine ligands that reside in the CX3-CX2-C motif [48]. Kaminska et al [10] demonstrated that mutation of this motif abolishes activity of Cfr, a member of radical SAM superfamily which encodes a methyltransferase. Conserved cysteine residues are also located in a CX2-CX5-C conformation starting at alignment position 456. Another feature common to all radical SAM proteins is a glycine-rich sequence motif proposed to be the SAM-binding site. Both GGE/DP and TNG/A motifs were identified in 60% and 50% of radical SAM proteins respectively. These putative SAM binding sequence motifs have also been observed in AtsB, a Fe-S oxidoreductase arylsulfatase regulatory protein [49]. In addition to these, other motifs of note included a YD/N motif present in 60% of the aligned proteins, a CN motif also found in 60% of the alignments and a PC/S motif present in 50%. A number of conserved single residues were identified including a number of aromatic residues. Alignment of the TrnC homologs (Fig. 3) also revealed several conserved regions summarized in Table 2. As with the TrnD homologs, the heterogeneity of TrnC homologs is most pronounced within the C-terminal regions which are believed to be responsible for binding of substrates and auxiliary cofactors, whereas the sequences of the N-termini are well conserved. The characteristic CX3-CX2-C motif was found in 12 TrnC homologs; B. thuringiensis serovar pulsarius BGSC 4CC1 Bthur0012_58010, Thermoanaerobacter mathranii A3 Tmath_1971 and Clostridium difficile QCD-63q42 CdifQCD-6_020200015426 did not contain the motif. Tmath_1971 and CdifQCD-6_020200015426 were also lacking the GGE/DP and CX2CX5CX4C motif. Bthur0012_58010 did contain the GGEPLL and the CX2CX5CX4C motif. Other motifs of note include a SXDG motif located starting at alignment position 340 and conserved in 89% of the strains. A CEK/R and a MX2D/E motif were conserved in 68.75% of the proteins. Highly conserved single residues included, again, a number of highly conserved aspartic acid residues. One of four conserved aspartic acid residues was found to be adjacent to a glycine in TrnC and 7 other homologs. These are likely to be important for the maintenance of the SAM binding site [50].
The conserved nature of these TrnC and Trn D homologs facilitated a phylogenetic analysis of their relatedness. A neighbour-joining tree generated from aligned conserved regions on the basis of percent identity was constructed for both sets of homologs. The unrooted cladogram of the Trn D homologs showed that *B. thuringiensis* DPC 6431 TrnD is most closely related to its homolog in *B. cereus* 95/8201, clustering together in the same branch (Fig. 4). This supports the findings from the in-silico screen. The cladogram of Trn C homologs (Fig. 5) is quite similar to that of the Trn D homologs. Once again *B. thuringiensis* DPC 6431 is shown to be most closely related to *B. cereus* 95/8201.

**Discussion**

Thuricin CD, a bacteriocin with a narrow spectrum of activity which includes *C. difficile*, was recently identified as providing a potential therapeutic solution to this important pathogen [4]. *C. difficile* infection (CDI) is the leading cause of nosocomial infections associated with antibiotic use and is responsible for substantial morbidity and mortality worldwide [51]. The posttranslational modifications found in thuricin CD are unusual, and had not previously been associated with a two-peptide bacteriocin. In addition to potentially providing a treatment for CDAD, it, together with subtilisin A, may also represent a previously undefined structural class of bacteriocins, the sactibiotics. Bearing this in mind an in-silico approach was used to identify other thuricin CD-like gene clusters. An in-silico approach to identify strains that may be capable of bacteriocin production has recently become a viable alternative to culture based approaches as a consequence of the increasing generation and availability of genomic and metagenomic sequence data. While in silico identification of gene clusters in a strain will not always lead to the detection of an associated bacteriocin, previous studies have shown that there is a high correlation [13,52].

The initial in silico strategy adopted here resulted in the initial identification of 99 TrnC and 53 TrnD homologs. Further analysis

![Figure 5. Rectangular cladogram of TrnC and all the TrnC homologs encountered during the screen. Cladogram was generated using the R package and visualised in DENDROSCOPE. doi:10.1371/journal.pone.0020852.g005](image-url)
led to the identification of 15 novel clusters which prior to this study had not been identified as potential bacteriocin-like clusters. These gene clusters are predominantly within the Firmicutes, with all but two gene clusters from species within this phylum. It was interesting to also find gene clusters within the Bacteroidetes phylum, namely Bacteroides sp_3_1_19, and also within the Thermatogae phylum, namely Petrotoga mobilis SJ95, because bacteriocin production within these phyla is quite rare. The availability in the future of additional species corresponding to these genera will reveal whether these gene clusters are exceptional or not. The analysis of metagenomic sequence data further highlighted the distribution of gene clusters containing TrnC and TrnD-like genes. While at present it is not possible to predict definitively if these metagenomic homologues are encoded with bacteriocin-like gene clusters, the possibility that at least a portion of these encode novel sacitiotics is a fascinating prospect. Indeed, the harnessing of such novel antimicrobials from genomic/metagenomic material through heterologous expression or through isolation and investigation of corresponding natural producers should greatly expand the collection of sacitiotics at our disposal.

The importance of thuricin CD as a potential therapeutic against pathogenic C. difficile has previously been reported [4]; locating thuricin CD-like bacteriocins may have huge benefits to the field. Genomic mining with TrnC and TrnD from the Thuricin CD operon revealed 100 and 54 homologs respectively; resulting in the identification of 15 with novel potential sacitiotic-encoding gene clusters. The identification of additional clusters from diverse metagenomic samples highlights the broad distribution of potential sacitiotic gene clusters.

Materials and Methods

Sequence Analysis

The protein sequences of TrnC and TrnD from Bacillus thuringiensis DPC 6431 were used as driver sequences to mine bacterial genomes using the BLASTP web server on NCBI (www.ncbi.nlm.nih/BLAST) using default parameters [53]. Homology was defined as an e-value greater than 1e-6 and greater than 20% identity. Genes surrounding the TrnC and TrnD homologs were examined using the genome viewer on NCBI and also using the DNA sequence viewer ARTEMIS [54].

Phylogenetic analysis

Protein alignments were generated by MUSCLE [55]. A phylogenetic tree was constructed using the TrnC and TrnD homologs identified in the screen. Protein trees were built using the R statistical package (http://cran.r-project.org) and the resultant phylogenetic trees were visualised using the DENDROSCOPE package [56].

Metagenomic Analysis

The TrnC and TrnD protein sequences were used as driver sequences to search for homologs in metagenomic datasets. The metagenomic portal CAMERA (http://web.cameracalit2.net) [44] was used to BLAST against all publicly available metagenomic datasets. The criterion for homolog detection was a threshold of 1e-6 and greater than 20% identity.

Supporting Information

Table S1 Bacterial genomes in which TrnC homologs were identified. (DOC)

Table S2 Bacterial genomes in which TrnD homologs were identified. (DOC)

Table S3 TrnC homologs in metagenomic databases. (DOC)

Table S4 TrnD homologs in metagenomic databases. (DOC)

Author Contributions

Conceived and designed the experiments: OOS PC. Performed the experiments: KM. Analyzed the data: KM OOS. Wrote the paper: KM OOS PC. Principal investigator and intellectual input: KPR CH.

References


