

Title	Structural strength analysis of partially crystalline trehalose
Authors	Maidannyk, Valentyn;Roos, Yrjö H.
Publication date	2017-09-27
Original Citation	Maidannyk, V. A. and Roos, Y. H. (2018) 'Structural strength analysis of partially crystalline trehalose', LWT - Food Science and Technology, 88, pp.9-17. doi:10.1016/j.lwt.2017.09.038
Type of publication	Article (peer-reviewed)
Link to publisher's version	10.1016/j.lwt.2017.09.038
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Download date	2024-04-20 14:27:58
Item downloaded from	https://hdl.handle.net/10468/6009



Accepted Manuscript

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PII: S0023-6438(17)30713-2

DOI: 10.1016/j.lwt.2017.09.038

Reference: YFSTL 6553

To appear in: LWT - Food Science and Technology

Received Date: 7 June 2017

Revised Date: 25 September 2017

Accepted Date: 26 September 2017

Please cite this article as: Maidannyk, V.A., Roos, Y.H., Structural strength analysis of partially crystalline trehalose, *LWT - Food Science and Technology* (2017), doi: 10.1016/j.lwt.2017.09.038.

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Structural strength analysis of partially crystalline trehalose.

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1. Abstract

Strength concept, which is based on the Williams-Landel-Ferry (WLF) model, was developed using mixed structured powders containing amorphous and crystalline components. At the present study, semi-crystalline trehalose powders with various (100:0; 80:20; 60:40; 40:60; 20:80) amorphous to crystalline ratios were analyzed. Amorphous components were prepared from water solution by freeze-drying. Strength analysis, which included water sorption, differential scanning calorimetry, dynamic mechanical analysis and microscopy, was applied. The results indicated that water content significantly decreases glass transition (~100°C) and α-relaxation temperatures (~90°C) as well as structural strength parameter (~10°C), while, the effect of crystalline component is less pronounced. This study can be used in processing and characterization of various partially crystalline food products including nutritional

Keywords: Structural relaxation, WLF, glass transition, semi-crystalline structure.

2. Introduction

formulations and infant formulas.

The improvement of processing and storage techniques requires a deep and fundamental knowledge of thermodynamics and kinetics. Food solids can exist in crystalline, amorphous or partially (e. g. semi) crystalline powders. Crystalline and amorphous materials show significantly different physicochemical properties (Bhandari, Bansal, Zhang & Schuck, 2013), due to differences in microstructure. Crystalline structures have long range molecular order, while amorphous structures are more disordered (short range molecular alignment) (Nurhadi & Roos, 2016). To preserve the taste, flavor and color of food the materials should be maintained in the amorphous form (Roos & Drusch, 2015). Amorphous materials are thermodynamically unstable compared to crystalline structures, however they are fairly stable in the glassy state (e. g. glass) (Slade, Levine & Reid, 1991). At temperatures close to the calorimetric glass transition temperature (Tg), physical properties of solids, such as molecular mobility, viscosity, etc. significantly change and materials are converted to supercooled

- 35 liquids (e. g. rubber) showing time-dependent flow (Angell, Ngai, McKenna, McMillan &
- Martin, 2000). Hence, obtaining the time-dependent characteristics of thermal, electric and
- 37 mechanical changes is practically important. Structural strength concept, proposed by Roos
- and co-workers combine temperature differences (T-T_g) and a practically important time
- 39 factor (critical change in structural relaxation time) (Roos et al., 2015).
- 40 Trehalose is a natural disaccharide of glucose with a high T_g (Green & Angell, 1989).
- 41 Trehalose is widely used in food and biotechnology areas due to specific physicochemical
- 42 properties such as prevention of biomolecules degradation (Crowe, Crowe, Rudolph,
- Womersley & Appel, 1985; Uritani, Takai & Yoshinaga, 1995); preservation of vaccines and
- medical proteins capabilities (Xie & Timasheff, 1997; Miller & de Pablo, 2000).
- 45 Amorphicity of materials can be detected gravimetrically from changing mass during water
- sorption (Buckton & Darcy, 1995; Mackin et al., 2002; Lehto et al., 2006; Nurhadi & Roos,
- 47 2016). Amorphous materials have larger porosity and more hydroscopic properties, hence
- 48 amorphous structures show a higher sorption capacity than crystalline forms (Bhandari,
- 49 Bansal, Zhang & Schuck, 2013; Nurhadi & Roos, 2016).
- 50 Differential scanning calorimetry (DSC) has been used as a method for detection and
- 51 quantification of the amorphous components in carbohydrate systems. For this, sample is
- beated to above the T_g to get dehydration (T_h) (endotherm peak in DSC thermogram) and
- 53 recrystallization (exotherm peak in DSC thermogram). The areas of exothermal and
- 54 endothermal peaks are proportional to the amorphous content in a partially crystalline
- 55 sample. Another approach to detect amorphous component in material by DSC is to
- determine the change of specific heat capacity (C_p) over the glass transition (Saleki-Gerhardt,
- 57 Ahlneck & Zografi, 1994; Sebhatu, Angberg & Ahlneck, 1994; Lehto et al., 2006). Optical
- 58 (light) microscopy usually shows that the partially crystalline structure tends to increase in
- transparency while approaching the transition temperature of dehydration without losing their
- 60 external morphology (Sussich, Urbani, Princivalle & Cesaro, 1998). A dynamic mechanical
- analysis (DMA) in multi-frequency mode is a useful tool to characterize mechanical
- properties including α , β , γ -relaxations, which happen due to variation in molecular mobility
- 63 below and around T_g (Moates, Noel, Parker & Ring, 2001). Knowing the frequency of
- measurements allows obtaining the value of structural relaxation time of α -relaxation process
- 65 (Noel, Parker & Ring, 2000), which may be related to particle structure, collapse and viscous
- 66 flow. As the determination of viscous flow characteristics at temperatures close to the T_g is
- 67 extremely difficult, strength concept provides an estimation of resistance to structural
- changes for amorphous materials above the calorimetry onset temperature during heating.

Strength analysis, which included water sorption, DSC and DMA, was successfully applied for various carbohydrate-protein, carbohydrate-carbohydrate and miscible models such as trehalose-whey protein isolate (WPI) (Fan & Roos, 2016a, b; Maidannyk & Roos, 2016), lactose-WPI (Fan & Roos, 2016a, b; Maidannyk & Roos, 2017), lactose-trehalose (Fan & Roos, 2016a) and trehalose-maltodextrin (Maidannyk, Nurhadi & Roos 2017). These studies showed that structural strength linearly depends on concentration of components and significantly decreases with increasing water content in a system. However, structural strength analysis of partially crystalline systems has not been addressed. The main aim of the present study was to develop the strength model using partially crystalline systems of trehalose. For this, effect of crystalline component on structural strength of amorphous trehalose was under investigation and effects of water content on structural strength of the partially crystalline systems were also studied.

3. Materials and Methods

3.1 Materials

D-(+)-Trehalose crystalline dihydrate (Hayashibara Co., Ltd., Okayama, Japan) and deionized water (KB scientific, Cork, Ireland) were used without purification.

3.1.1 Amorphous structure

Amorphous structure was obtained by freeze-drying (Lyovac GT2, Steris[®], Hürth, Germany). For this, trehalose solution (total solid of 20%) in water was prepared. After that, $5x10^{-3}$ L aliquots of solution were frozen in pre-weighted and semi-closed with septum in $1x10^{-2}$ L glass vials (Schott, Mulheim, Germany) at -20°C for 24 h, then at -80°C for 3 h, followed by freeze-drying for 60 h at pressure p < 10 Pa. All vials were hermetically sealed under the vacuum conditions inside the freeze dryer at p < 10 Pa and stored over P_2O_5 in vacuum desiccators (Roos & Karel, 1990) at room temperature (25 ± 1°C) to protect samples from water uptake.

3.1.2 Crystalline structure

102	Crystalline trehalose powder was prepared by grinding commercial trehalose powder and
103	stored in a desiccator over P2O5 at 25±1°C for 3 days. A small amount of amorphous
104	structure can be detected after grinding procedure (Willart, Dujardin, Dudognon, Danede &
105	Descamps, 2010; Nurhadi & Roos, 2016).
106	
107	3.1.3 Partially crystalline structures
108	
109	Partially crystalline trehalose systems were prepared by blending accurately weighted amount
110	of 100% crystalline and 100% amorphous structures at 100:0; 80:20; 60:40; 40:60; 20:80 and
111	0:100 amorphous:crystalline ratios. Mixing was done by spatula and shaking the closed vials
112	(Nurhadi & Roos, 2016).
113	
114	3.2 Determination of the initial water content
115	
116	Samples of trehalose powder with final weight 0.5-1.0 g were dried at 70°C with absolute
117	pressure P _{abs} <10 ³ Pa for 24 hours in a WTB Binder vacuum oven (Mason Technology®,
118	Tuttingen, Germany) to measure the initial water content of the material. The difference in
119	mass of samples before and after drying was defined as initial water content.
120	
121	3.3 Water sorption analysis
122	
123	Partially crystalline systems with various ratios of amorphous and crystalline structures
124	(described above) were stored at evacuated desiccators (25±1°C) for 10 days over the
125	saturated solutions of P2O5, LiCl, CH3COOK, MgCl2, K2CO3, Mg(NO3)2, NaNO2, NaCl and
126	KCl (Sigma Chemical Co., St. Louis, MO. U.S.A.), which at equilibrium provided 0, 0.11,
127	0.23, 0.33, 0.44, 0.545, 0.66, 0.76 and 0.85 a _w , respectively. AQUALAB 4 (TE) (Decagon
128	Devices Inc., Pullman, WA., U.S.A.) water activity meter was used to measure water activity
129	for each material after storage. Samples were weighted at intervals of 0, 2, 4, 6, 8, 10, 24, 48,
130	72, 96 and 120 hours upon storage. Possible crystallization of amorphous trehalose was
131	assessed from the loss of sorbed water. The water content in each mixture was plotted as a
132	function of time, and the Guggenheim-Anderson-deBoer (GAB) relationship was fitted to

data to know water activity-water content dependence of amorphous trehalose systems (Eq.

135

1):

133

136
$$\frac{m}{m_0} = \frac{Cka_w}{(1 - ka_w)(1 - ka_w + Cka_w)}$$
 (1)

137

- where, m is water content, m₀ is the monolayer value and C and k were respectively
- 139 calculated from m_0 .
- 140 Equation 2 was used to determine the combined effects of amorphous and crystalline
- components (Bronlund & Paterson, 2004):

142

 $W_{mixture} = n_1 W_{crystalline} + n_2 W_{amorphous}$ (2)

144

- where, $W_{mixture}$ is the total equilibrium water content in the mixture; n_1 and n_2 are mass
- fractions of crystalline and amorphous components in the system $(n_1+n_2=1)$; $W_{crystalline}$ and
- $W_{amorphous}$ are water contents in crystalline and amorphous component.

148

149 3.4 Differential Scanning Calorimetry (DSC)

150

- Differential scanning calorimeter (DSC) (Mettler Toledo Schwerzenbach, Switzerland) was
- 152 used to measure the glass transition temperature, dehydration and spontaneous
- recrystallization of partially crystalline trehalose mixtures with 0, 0.11, 0.23, 0.33 and 0.44
- a_w. Samples of all mixtures were transferred to pre-weighted standard DSC aluminium pans
- 155 (40 µL, Mettler Toledo, Schwerzenbach, Switzerland) and hermetically sealed. An empty
- punctured pan was used as a reference. For anhydrous systems only, the lids of DSC
- 157 aluminium pans were punctured to allow evaporation of residual water upon the
- measurement. All samples were scanned with 5°C/min heating rate. The onset of T_g, heat
- capacity of endotherm and exotherm were determined by the STAR^e software version 8.10
- 160 (Mettler Toledo, Schwerzenbach, Switzerland).

161

162

3.5 Dynamical Mechanical Analyses (DMA)

- Dynamic mechanical analyzer (DMA) (Tritec 2000 DMA, Triton Technology Ltd.,
- 165 Grantham, Lincolnshire, UK) was used to measure mechanical properties (E'' loss
- modulus, E' storage modulus and $tan\delta = E''/E'$) of anhydrous and humidified partially
- 167 crystalline trehalose systems (described above for the DSC experiments). The DMA

instrument was balanced or set at zero to determine the zero displacement position before 168 starting. Approximately 60 g of ground samples were spread on a metal pocket-forming sheet 169 (Triton Technology Ltd., UK). This sheet was fixed between the stationary and drive shaft 170 clamps inside the measuring head of the DMA. Length, width and thickness were measured 171 for each sample. All results were obtained using 1.43.00 DMA software version. To control 172 temperature the DMA was connected to a liquid nitrogen tank (1L; Cryogun, Brymill 173 Cryogenic Systems, Labquip Ltd., Dublin, Ireland). Samples were scanned from ~ 50°C 174 below to over the α-relaxation region with cooling rate of 5°C/min and heating rate of 175 2°C/min using the single cantilever bending mode (Fan & Roos, 2016a, b; Maidannyk & 176 Roos, 2016). The α -relaxation temperatures (T_{α}) were determined from peaks of tan δ above 177 178 the glass transition.

Equation 3 was used to calculate the relaxation times (τ) of peak T_{α} , measured by DMA at

various frequencies (f) (Noel, Parker & Ring, 2000):

181

$$182 \tau = \frac{1}{2\pi f} (3)$$

183

184

3.6 Optical light microscopy

185

186

187

188

189

Microscope observation (OLYMPUS BX51 (Olympus Corporation, Tokyo, Japan), with magnification x20) was done on humidified (0, 0.11, 0.23 and 0.44 a_w) partially crystalline trehalose powders (described above for the DSC experiments) placed between cover glass and mounted Linkam 120 (TP 94) temperature controller stage. The samples were scanned from 5 to 150°C, with 2°C/min heating rate.

190 191

192

3.7 Calculation of WLF model constants and Structural Strength

193

The WLF equation in the form of (Eq. 4) was used to fit DMA and DSC data (Williams,

195 Landel & Ferry, 1955):

196

197
$$log_{10}\frac{\tau}{\tau_s} = log_{10}\frac{\eta}{\eta_s} = \frac{-c_1(T - T_g)}{c_2 + (T - T_g)}$$
 (4)

- where, τ is relaxation time, τ_s is reference relaxation time, η is viscosity, η_s is reference
- viscosity, T is temperature, T_g is glass transition temperature, C_1 and C_2 are constants.
- The WLF equation in the form of (Eq. 5) suggested that the plot of $1/lg(\tau/\tau_s)$ versus $1/(T-T_g)$
- 202 gives a linear correlation:

203

$$204 \quad \frac{1}{lg\frac{\tau}{\tau_s}} = \frac{1}{-c_1} - \frac{c_2}{c_1(T - T_g)} \tag{5}$$

205

- The WLF constants C_1 and C_2 were derived from the slope and intercept of the straight line
- 207 (Roos & Drusch, 2015).
- 208 Mathematically, structural strength parameter (S) is based on WLF relationship and can be
- calculated by Equation (6):

210

$$S = \frac{dC_2}{C_1 - d} \tag{6}$$

212

- where d is a parameter, showing the critical decrease in the number of logarithmic decades
- for the flow (e.g., 100 s to 0.01 s corresponds to d = 4; can be chosen for each system as an
- integer depending on the critical time for the process (Fig. 4)), C_1 and C_2 are "non-universal"
- 216 constants in the WLF equation.
- Equation 7 was used to predict structural strength at different water contents:

218

$$S = \frac{w_1 S_1 + k w_2 S_2}{w_1 + k w_2} \tag{7}$$

220

- where w_1 weight fraction of dry solid; w_2 weight fraction of water; k coefficient; S_1 –
- structural strength for anhydrous system; S_2 structural strength of pure water (S_2 = 6.0)
- 223 (Maidannyk & Roos, 2017).

224

225 3.8 Data analysis

226

- All experiments were performed in triplicate. Mean data of the water sorption analyses, DSC,
- DMA were calculated from 3 replicates with standard deviations expressed in error bars.

230 4. Results and discussion

232 4.1 Water sorption analysis

233

- Table 1 and Figure 1 shows experimental water content of partially crystalline trehalose
- systems, over the whole range of water activities at $25\pm1^{\circ}$ C.
- 236 Crystalline structures humidified at low water activities (0-0.76 a_w) adsorb very little amount
- of water while at high water activities (0.85-0.99 a_w) the amount of sorbed water increases
- exponentially (Table 1) due to capillary condensation and dissolution (Bronlund & Paterson,
- 239 2004).
- Steady-state water contents of each mixture at 120 hours were used in GAB model. The GAB
- model was fitted to experimental data of 100:0 amorphous:crystalline systems below 0.55 a_w
- 242 (Labuza, 1984; Maidannyk & Roos, 2016). However, at higher water activities (≥0.55 a_w), the
- 243 crystallization of trehalose occurred and extrapolated sorption data from GAB relationship
- 244 give misleading results for sorbed water contents (Potes, 2014). Fractional isotherm approach
- 245 (Eq. 2) allowed calculating water contents at high water activities. Using previously
- published data for pure amorphous trehalose (Maidannyk & Roos, 2016; Maidannyk,
- Nurhadi & Roos, 2017) and experimental data for pure crystalline trehalose, the GAB model
- was fitted to obtain sorption isotherms at high water activities (Fig. 1). Amorphous structure
- adsorbs around 100 times more water than crystalline structure (Table 1). That's why the
- 250 presence of a small amount of amorphous component in crystalline structure results in
- significant difference to the water sorption isotherm of material. Water sorption isotherm and
- Eq. 2 allows predicting water content for partially crystalline trehalose at all water activities
- and all amorphous:crystalline ratios.

254255

4.2 Differential Scanning Calorimetry and Microscopy

- 257 The onset of calorimetric glass transition temperature (Tg) was obtained by DSC for each
- anhydrous and humidified (0-0.44a_w) partially crystalline trehalose system (Table 2).
- 259 Detected values of T_g belong to amorphous component in partially crystalline systems. The
- obtained data for pure anhydrous trehalose is in agreement with previous study (Green &
- 261 Angell, 1989; Roos 1993; Miller & de Pablo, 2000; Maidannyk & Roos, 2016; 2017 a). As
- expected crystalline fraction has no significant effect on the $T_{\rm g}$ of the amorphous fraction in

the partially crystalline systems (Table 2). Glass transition is that of the amorphous fraction which is shown by the proportional decrease of heat capacity (C_p) over the T_g region. A slight decrease of T_g value with increase of crystalline component content in the systems is in agreement with water sorption data (Table 1). Water as a good and strong plasticizer significantly increases mobility of molecules of the amorphous component, which results in the decrease of T_g values with increasing water content (Maidannyk & Roos, 2016). For example, in the present study T_g of 100:0 amorphous:crystalline system decreased from $112^{\circ}C$ (at 0.44 a_w) due to water plasticization. Microscope observation, carried out on the same samples placed between cover glasses,

Microscope observation, carried out on the same samples placed between cover glasses, showed that crystalline and amorphous components of trehalose became more transparent during heating to above T_g . However, at the same time the external morphology of the structure was not changed until the temperature about 135° C, that the "solid" sample appears to change morphological shape ("edge roundness") with an increasing mobility of the phase, although still firm (sticky) (Sussich et al, 1998) (Figure 2). These observations were the same for anhydrous and humidified (0.11, 0.23, 0.33 and 0.44a_w) partially crystalline trehalose systems.

Partially crystalline trehalose systems with different ratios of amorphous and crystalline

components (described above for DSC experiments) were scanned by DMA in multi-

4.3 Dynamical Mechanical Analyses (DMA)

frequency mode. The rapid changes in mechanical properties, which related to the amount of energy converted to heat during relaxation can be detected by DMA, while DSC directly detects the difference in the amount of heat (Gonnet, Guillet, Sirakov, Fulchiron & Seytre, 2002; Roos & Drusch, 2015). Figure 3 shows typical frequently-dependent thermogram obtained by DMA. The α -relaxation temperature values were determined from the temperature peak of $\tan\delta$, at \sim 20-30°C above the onset of the T_g . Systems with high amount of crystalline component showed slightly lowered α -relaxation temperature peak with less intensity, due to significant difference between crystalline and amorphous particle sizes (crystalline particles needs less mechanical energy than amorphous and α -relaxations occurs at lower temperature). Also crystalline phase may slightly decreased α -relaxation temperature due to sliding effect inside the partially crystalline system (Cano-Chauca, Stringheta, Ramos & Cal-Vidal, 2005). The big differences (\sim 6-10°C) were observed for anhydrous partially crystalline trehalose

- systems, while for humidified systems (0.44 a_w) these differences were not significant (~2-
- 298 5°C) and α-relaxation occurred at similar temperatures at all amorphous and crystalline ratios
- 299 (Table 3).
- Water as a good plasticizer, increases free volume in the systems as well as molecular
- mobility of amorphous components (Slade et al., 1991; Royall et al., 2005; Meinders and van
- Vliet, 2009) that result in significant decrease (~85°C for 100:0 amorphous:crystalline) of the
- T_{α} for every partially crystalline system with increasing water content. These results are in
- agreement with previous studies (Fan & Roos, 2016 a, b; Maidannyk & Roos, 2016; 2017;
- 305 Maidannyk, Nurhadi & Roos, 2017).

306

4.4 Applications of WLF equation and Strength

308

- 309 Williams-Landel-Ferry or WLF model (Eq. 4) is a popular equation, which can describe
- 310 dependence of relaxation time and viscosity on the temperature close to onset T_g. Original
- relationship (Williams et al., 1955) showed that glass formers possessed similar decreases in
- relaxation times over the temperature range of T_g to $T_g + 100$ K. Because of that authors
- offered "universal" values for WLF constants: $C_1 = 17.44$ and $C_2 = 51.6$ for many different
- 314 materials (inorganic and organic). However, using WLF equation with "non-universal"
- 315 constants C₁ and C₂ provides more realistic modelling with good fit to experimental data
- 316 (Ferry, 1980; Peleg, 1992; Slade & Levine, 1995; Peleg & Chinachoti, 1996; Roos & Drusch,
- 317 2015;). In the present study, WLF constants C₁ and C₂ were calculated by Eq. 5 and
- summarized in table 4. C₁ and C₂ constants were determined with 0.9651, 0.9210, 0.9116,
- 319 0.9491 and 0.7622 R² for 100:0; 80:20; 60:40; 40:60 and 20:80 amorphous:crystalline ratios
- at RH 0%; 0.9707, 0.8933, 0.9281, 0.9681 and 0.9013 at RH 11%; 0.8079, 0.8694, 0.7596,
- 321 0.9664 and 0.7279 at RH 23%; 0.9987, 0.9491, 0.9506, 0.9627 and 0.7639 at RH 33%;
- 322 0.7440, 0.9742, 0.9715, 0.9807 and 0.9698 at RH 44% respectively.
- In calculations, the viscosity and relaxation time of supercooled liquid approached 10¹² Pa s
- and 100 s respectively. Upon heating, values of viscosity and relaxation time were decreasing
- down to 10⁵ Pa s and 10⁻¹⁴ s respectively (Angell, 1991; Angell, Ngai, McKenna, McMillan &
- 326 Martin, 2000; Roos & Drusch, 2015; Maidannyk & Roos, 2016). Using 10¹² Pa s and 100 s as
- 327 reference values of viscosity and structural relaxation time respectively, Roos and co-workers
- 328 (Roos et al., 2015; Fan & Roos, 2016 a ,b; Maidannyk & Roos, 2016; 2017; Maidannyk,
- Nurhadi & Roos, 2017) adapted WLF equation in a simple and convenient form, named
- "Strength" concept. Structural strength measures solid flow characteristics of amorphous

331	materials above $T_{\rm g}$ and links the material state with a practically important time factor. As
332	large and critical changes in structural relaxation time upon heating happened between 100 s
333	and 0.01 s (between 2 and -2 in logarithmic scale) (Fig. 4), the corresponding temperature
334	difference, T-T _g , was defined as strength, S (Eq. 6) (Roos et al., 2015; Fan & Roos, 2016 a ,b;
335	Maidannyk & Roos, 2016; 2017; Maidannyk, Nurhadi & Roos, 2017).
336	S was obtained (at $d = 4$) for partially crystalline trehalose systems at all ratios (Table 5).
337	Strength curves, which show structural strength dependence on water content, were
338	calculated by Eq. 7, which allows predicting S value at all water contents (from 0 to 100
339	g/100g of dry Solids) (Fig. 5). The k value was 15.2, 26.0, 17.1, 21.0 and 26.0 for 100:0,
340	80:20, 60:40, 40:60 and 20:80 amorphous:crystalline trehalose systems respectively.
341	Table 5 and Figure 5 shows that for anhydrous systems (RH 0%) the S value significantly
342	decreased ($\Delta S = 9.1$ °C) with increasing crystalline component in a system, while for
343	humidified systems (RH 44%) these differences were not so pronounced ($\Delta S = 1.7$ °C). This
344	result is similar to our previous findings and can be explained by molecular mobility of
345	amorphous component and by strong plasticization properties of water (Roos et al., 2015; Fan
346	& Roos, 2016 a, b; Maidannyk & Roos, 2016; 2017; Maidannyk, Nurhadi & Roos, 2017).
347	Even small variation in material structure significantly changes the structural strength in a
348	system (Fig. 6). Amorphous systems show more "strong" behavior compare to "weak" highly
349	crystalline trehalose systems. However, this difference is significant only for anhydrous
350	systems, while humidified systems shows very similar values of strength for all partially
351	crystalline system. Hence, the presence of water is a very important factor, which
352	significantly influences a structural strength and structure of food material. Despite the fact
353	that for humidified systems, the values of S are similar, they are still decreasing linearly (with
354	$R^2\ 0.9879,\ 0.9310,\ 0.9045,\ 0.8860,\ 0.8816,\ 0.8789,\ 0.8767,\ 0.8750,\ 0.8743\ and\ 0.8737\ at\ 0\%,$
355	10%, 20%, 30%, 40%, 50%, 60%, 70%, 80% and 90% of water respectively) with increasing
356	crystalline content in partially crystalline systems (Fig. 7).
357	Knowing structural strength of pure amorphous component and reference partially crystalline
358	systems allows calculation of S value for systems at any crystalline content at any water
359	content, as well as prediction of crystalline content in unknown partially crystalline system.
360	Therefore, the present study showed that strength concept is a suitable model for determining
361	crystalline content in partially crystalline system and allows controlling stability and quality
362	in various food systems.

363

364

5. Conclusions

365

This study was focused on applying and further development of the structural strength 366 concept. For this, strength analysis was applied in partially crystalline trehalose system. 367 Water sorption analysis allows controlling water contents in different partially crystalline 368 mixtures at low and high water activities. The glass transition and α -relaxation temperatures 369 were detected by DSC and DMA respectively. The values of T_g and T_α as well as structural 370 371 strength are significantly decreasing with increasing water content in a system. At very low content of amorphous component in a system, the determination of strength is practically 372 complex. However, strength analysis showed good consistency in predicting the crystalline 373 content in unknown systems at different water contents. The structural strength shows linear 374 decrease with increase of crystalline content in a system and significant decrease with 375 increase of the water content in a system. Hence structural strength concept can be used in 376 adaptation of processing and characterization of various food materials as well as in quality 377 and stability control during production and storage. 378

379

380

6. Acknowledgments

381

- 382 This work was supported by the Food Institutional Research Measure (FIRM) project
- 383 "Formulation and Design for Food Structure and Stability" funded by the Department of
- Agriculture, Food and Marine (11-F-001), coordinated by prof. Y.H. Roos, UCC, Ireland.

385

386 7. References

- Angell, C.A., Ngai, K.L., McKenna, G.B., McMillan, P.F., Martin, S.W., (2000).
- Relaxation in glassforming liquids and amorphous solids. *Journal of Applied Physics* 88(6),
- 390 *3113-3157*.
- Bhandari, B., Bansal, N., Zhang, M. and Schuck, P. (2013). Handbook of food powders:
- processes and properties. Elsevier. Woodhead Publishing. Limited. Oxford, pp. 660.
- Bronlund, J., Paterson, T., (2004). Moisture sorption isotherms for crystalline, amorphous
- and predominantly crystalline lactose powders. *International Dairy Journal* 14(3), 247-254.
- Buckton, G., Darcy, P., (1995). The use of gravimetric studies to assess the degree of
- 396 crystallinity of predominantly crystalline powders. *International Journal of Pharmaceutics*
- 397 *123(2)*, *265-271*.

- 398 Cano-Chauca, M., Stringheta, P. C., Ramos, A. M., Cal-Vidal, J. (2005). Effect of the
- 399 carriers on the microstructure of mango powder obtained by spray drying and its functional
- 400 characterization. *Innovative Food Science & Emerging Technologies*, 6(4), 420-428.
- 401 Crowe, L.M., Crowe, J.H., Rudolph, A., Womersley, C., Appel, L., (1985). Preservation
- of freeze-dried liposomes by trehalose. Archives of Biochemistry and Biophysics 242(1), 240-
- 403 247.
- 404 Fan, F., Roos, Y.H., (2016a). Structural relaxations of amorphous lactose and lactose-whey
- 405 protein mixtures. *Journal of Food Engineering 173, 106-115*.
- 406 Fan, F., Roos, Y.H., (2016 b). Structural strength and crystallization of amorphous lactose in
- 407 food model solids at various water activities. Innovative Food Science & Emerging
- 408 Technologies 40 (2017): 27-34.
- 409 Gonnet, J.M., Guillet, J., Sirakov, I., Fulchiron, R. Seytre, G., (2002). "In-situ"
- 410 monitoring of the non-isothermal crystallization of polymers by dielectric spectroscopy.
- 411 *Polymer Engineering Science* 42(6), 1159-1170.
- 412 Green, J.L., Angell, C.A., (1989). Phase relations and vitrification in saccharide-water
- solutions and the trehalose anomaly. *The Journal of Physical Chemistry 93(8)*, 2880-2882.
- 414 Labuza, T.P., (1984). Moisture sorption: Practical aspects of isotherm measurement and use.
- 415 American Association of Cereal Chemists, St Paul, U.S.A., pp. 158.
- Lehto, V.-P., Tenho, M., Vähä-Heikkilä, K., Harjunen, P., Päällysaho, M., Välisaari, J.,
- Niemelä, P., Järvinen, K., (2006). The comparison of seven different methods to quantify
- 418 the amorphous content of spray dried lactose. *Powder Technology* 167(2), 85-93.
- 419 Mackin, L., Zanon, R., Park, J.M., Foster, K., Opalenik, H., Demonte, M., (2002).
- Quantification of low levels (< 10%) of amorphous content in micronised active batches
- 421 using dynamic vapour sorption and isothermal microcalorimetry. *International Journal of*
- 422 *Pharmaceutics 231(2), 227-236.*
- 423 Maidannyk, V., Roos, Y., (2016). Modification of the WLF model for characterization of
- 424 the relaxation time-temperature relationship in trehalose-whey protein isolate systems.
- 425 *Journal of Food Engineering 188, 21-31.*
- 426 Maidannyk, V. A., and Y. H. Roos., (2017). Water sorption, glass transition and "strength"
- of lactose–Whey protein systems. *Food Hydrocolloids* 70, 76-87.
- 428 Maidannyk, V. A., Nurhadi, B., & Roos, Y. H. (2017). Structural strength analysis of
- amorphous trehalose-maltodextrin systems. Food Research International, 96, 121-131.
- 430 Meinders, M.B., van Vliet, T., (2009). Modeling water sorption dynamics of cellular solid
- food systems using free volume theory. *Food Hydrocolloids 23(8) 2234-2242*.

- 432 Miller, D.P., de Pablo, J.J., (2000). Calorimetric solution properties of simple saccharides
- and their significance for the stabilization of biological structure and function. *The Journal of*
- 434 *Physical Chemistry B* 104(37), 8876-8883.
- 435 Moates, G., Noel, T., Parker, R., Ring, S., (2001). Dynamic mechanical and dielectric
- characterisation of amylose–glycerol films. *Carbohydrate Polymers* 44(3), 247-253.
- Noel, T.R., Parker, R., Ring, S.G., (2000). Effect of molecular structure and water content
- on the dielectric relaxation behaviour of amorphous low molecular weight carbohydrates
- above and below their glass transition. Carbohydrate Research 329(4), 839-845.
- Nurhadi, B., Roos, Y., (2016). Dynamic water sorption for the study of amorphous content
- of vacuum-dried honey powder. *Powder Technology 301, 981-988.*
- Peleg, M., Chinachoti, P., (1996). On modeling changes in food and biosolids at and around
- 443 their glass transition temperature range. Critical Review in Food Science and Nutritional
- 444 *36(1-2), 49-67.*
- Roos, Y., Karel, M., (1990). Differential scanning calorimetry study of phase transitions
- affecting the quality of dehydrated materials. *Biotechnology Progress* 6(2), 159-163.
- 447 Roos, Y.H., Drusch, S., (2015). Phase transitions in foods. Academic Press, Elsevier,
- 448 *Waltham MA., U.S.A.*, pp. 368.
- Roos, Y.H., Fryer, P.J., Knorr, D., Schuchmann, H.P., Schroën, K., Schutyser, M.A.,
- 450 Trystram, G., Windhab, E.J., (2015). Food engineering at multiple scales: case studies,
- challenges and the future—a European perspective. *Food Engineering Reviews, 1-25.*
- 452 Saleki-Gerhardt, A., Ahlneck, C., Zografi, G., (1994). Assessment of disorder in
- 453 crystalline solids. *International Journal of Pharmaceutics 101(3), 237-247*.
- Sebhatu, T., Angberg, M., Ahlneck, C., (1994). Assessment of the degree of disorder in
- 455 crystalline solids by isothermal microcalorimetry. *International Journal of Pharmaceutics*
- 456 *104(2)*, *135-144*.
- 457 Slade, L., Levine, H., Reid, D.S., (1991). Beyond water activity: recent advances based on
- an alternative approach to the assessment of food quality and safety. Critical Reviews in Food
- 459 Science & Nutrition 30(2-3), 115-360.
- Sussich, F., Urbani, R., Princivalle, F., Cesàro, A., (1998). Polymorphic amorphous and
- 461 crystalline forms of trehalose. Journal of the American Chemical Society 120(31), 7893-
- 462 *7899*.
- 463 Uritani, M., Takai, M., Yoshinaga, K., (1995). Protective effect of disaccharides on
- restriction endonucleases during drying under vacuum. Journal of Biochemistry 117(4), 774-
- 465 *779*.

	ACCEPTED MANUSCRIPT
466	Willart, J., Dujardin, N., Dudognon, E., Danède, F., Descamps, M., (2010).
467	Amorphization of sugar hydrates upon milling. Carbohydrate Research 345(11), 1613-1616.
468	Williams, M.L., Landel, R.F., Ferry, J.D. (1955). The temperature dependence of
469	relaxation mechanisms in amorphous polymers and other glass-forming liquids. Journal of
470	the American Chemical Society 77(14), 3701-3707.
471	Xie, G., Timasheff, S.N., (1997). The thermodynamic mechanism of protein stabilization by
472	trehalose. Biophysical Chemistry 64(1), 25-43.
473	

Table 1. Water content of partially crystalline trehalose systems (ratios of components 100:0, 80:20, 60:40, 40:60, 20:80, 0:100) stored at different water activities (0.11, 0.23, 0.33, 0.44, 0.55, 0.65, 0.76, 0.85, 0.99) for 120 hours at 25±1°C.

Experimental water content for partially crystalline (amorphous:crystalline										
$\mathbf{a}_{\mathbf{w}}$	trehalose systems (g/100 g of solids)									
	100:0	80:20	60:40	40:60	20:80	0:100				
0.11	2.2±0.1	1.7±0.1	1.3±0.1	0.9±0.1	0.4±0.1	0.010±0.01				
0.23	4.1 ± 0.4	3.3 ± 0.1	2.5 ± 0.3	1.7 ± 0.2	0.8±0.2	0.014±0.03				
0.33	5.9 ± 0.4	4.7 ± 0.2	3.7 ± 0.1	2.4 ± 0.2	1.2±0.2	0.019±0.03				
0.44	9.0 ± 0.2	6.9 ± 0.2	5.3±0.1	3.4 ± 0.3	1.8±0.2	0.023 ± 0.05				
0.55	9.5±0.1	7.6 ± 0.1	5.6 ± 0.3	3.8±0.3	1.9±0.3	0.029 ± 0.04				
0.65	9.4 ± 0.3	7.6 ± 0.2	5.7±0.1	3.8±0.1	1.9±0.3	0.037 ± 0.02				
0.76	8.8 ± 0.4	7.2 ± 0.3	5.5 ± 0.2	3.6±0.2	1.8±0.2	0.049 ± 0.05				
0.85	9.5 ± 0.4	7.6 ± 0.1	5.5 ± 0.2	3.7±0.2	1.9 ± 0.3	0.066 ± 0.02				
0.99	9.7±0.3	7.8±0.3	5.7±0.3	3.8±0.3	2.3±0.2	0.598±0.09				

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Table 2. The onset of calorimetric glass transition temperatures (T_g) of anhydrous $(0 \ a_w)$ and humidified $(0.11\text{-}0.44 \ a_w)$ partially crystalline trehalose systems (amorphous:crystalline) stored for 120 hours at $25\pm1^{\circ}C$.

Glass transition temperature (T_g) of partially crystalline (amorphous:crystalline) trehalose systems, ${}^{\rm o}{\rm C}$

\mathbf{a}_{w}	100:0	80:20	60:40	40:60	20:80
0	112±3	112±2	111±3	111±2	111±2
0.11	62±1	61±3	60±2	59±2	59±2
0.23	43±2	43±1	42±3	42±1	41±1
0.33	32±2	32±1	32±2	31±3	31±2
0.44	15±2	15±3	15±2	15±1	14±3

Table 3. α -Relaxation temperature (T_{α}) detected by DMA for partially crystalline trehalose systems with different amorphous:crystalline ratios, equilibrated at different relative humidities (RH).

RH 0%	RH 0% Amorphous:Crystalline trehalose							
		100:0	80:20	60:40	40:60	20:80		
f, Hz	$\log \tau$, s	T _α , °C	T _a , °C	T_{α} , ${}^{\circ}C$	T_{α} , ${}^{\circ}C$	T_{α} , °C		
0.1	0.20	123±2	122±2	118±2	117±2	116±1		
0.5	-0.49	125±1	123±1	119±1	118±1	116±2		
1.0	-0.80	127 ± 2	125±2	120 ± 2	119±2	117±2		
2.0	-1.10	128 ± 2	126±3	121±1	120±2	118±1		
5.0	-1.50	128 ± 3	127±1	123±3	121±2	119±3		
10.0	-1.80	130 ± 1	128±2	124±2	122±1	120 ± 2		
RH 11%			Amorphous:C	rystalline tre	ehalose			
		100:0	80:20	60:40	40:60	20:80		
f, Hz	logτ, s	T _α , °C	T _α , °C	T_{α} , °C	T _α , °C	T _α , °C		
0.1	0.20	71 ± 2	68±2	66±2	64 ± 2	64 ± 2		
0.5	-0.49	73 ± 2	69±2	67±2	65 ± 2	65 ± 2		
1.0	-0.80	75 ± 1	70±3	68±2	66 ± 2	65±3		
2.0	-1.10	77 ± 2	71±2	69±1	67 ± 2	66±1		
5.0	-1.50	78 ± 2	73±1	70±2	68±3	67±2		
10.0	-1.80	79±3	75±2	72±2	69±3	68±2		
RH 23%			Amorphous:C	•	ehalose			
		100:0	80:20	60:40	40:60	20:80		
f, Hz	loge s	$\mathbf{T} \circ \mathbf{C}$	$\mathbf{T} \circ \mathbf{C}$	$\mathbf{T} \circ \mathbf{C}$	T_{α} , °C	T _α , °C		
1, 11Z	logτ, s	T _α , °C	T _α , °C	T _α , °C	1α, C	1α, C		
0.1	0.20	$\frac{1_{\alpha}, \mathbf{C}}{52\pm 1}$	51±2	50±3	49±1	47±2		
-					-			
0.1	0.20	52±1	51±2	50±3	49±1	47±2		
0.1 0.5 1.0 2.0	0.20 -0.49 -0.80 -1.10	52±1 53±1 53±2 54±2	51±2 52±3 52±2 53±2	50±3 50±2 51±2 52±3	49±1 50±2 51±3 51±1	47±2 47±1 48±2 48±2		
0.1 0.5 1.0 2.0 5.0	0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1	51±2 52±3 52±2 53±2 54±3	50±3 50±2 51±2 52±3 53±1	49±1 50±2 51±3 51±1 52±1	47±2 47±1 48±2 48±2 49±2		
0.1 0.5 1.0 2.0 5.0 10.0	0.20 -0.49 -0.80 -1.10	52±1 53±1 53±2 54±2	51±2 52±3 52±2 53±2 54±3 55±3	50±3 50±2 51±2 52±3 53±1 53±2	49±1 50±2 51±3 51±1 52±1 52±2	47±2 47±1 48±2 48±2		
0.1 0.5 1.0 2.0 5.0	0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1	51±2 52±3 52±2 53±2 54±3	50±3 50±2 51±2 52±3 53±1 53±2	49±1 50±2 51±3 51±1 52±1 52±2	47±2 47±1 48±2 48±2 49±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33%	0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1 57±2	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20	50±3 50±2 51±2 52±3 53±1 53±2 Tystalline tree 60:40	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60	47±2 47±1 48±2 48±2 49±2 50±2 20:80		
0.1 0.5 1.0 2.0 5.0 10.0	0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1 57±2	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C	50±3 50±2 51±2 52±3 53±1 53±2 Trystalline tre	49±1 50±2 51±3 51±1 52±1 52±2 ehalose	47±2 47±1 48±2 48±2 49±2 50±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33%	0.20 -0.49 -0.80 -1.10 -1.50 -1.80	52±1 53±1 53±2 54±2 55±1 57±2	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20	50±3 50±2 51±2 52±3 53±1 53±2 Tystalline tree 60:40	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60	47±2 47±1 48±2 48±2 49±2 50±2 20:80		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33%	0.20 -0.49 -0.80 -1.10 -1.50 -1.80	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T _a , °C	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _α , °C	50±3 50±2 51±2 52±3 53±1 53±2 Tystalline tree 60:40 T _α , °C	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60 T _α , °C	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T_α, °C		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33%	0.20 -0.49 -0.80 -1.10 -1.50 -1.80	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T _a , °C 38±2	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _a , °C 38±1	50±3 50±2 51±2 52±3 53±1 53±2 Trystalline tree 60:40 T _a , °C 37±1	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60 T _a , °C 36±1	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _a , °C 36±1 36±1 37±1		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33% f, Hz 0.1 0.5 1.0 2.0	0.20 -0.49 -0.80 -1.10 -1.50 -1.80 logτ, s 0.20 -0.49 -0.80 -1.10	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T _a , °C 38±2 40±2 41±2 42±3	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _α , °C 38±1 39±2 40±1 41±2	50±3 50±2 51±2 52±3 53±1 53±2 Trystalline tree 60:40 T _α , °C 37±1 39±1 39±2 41±1	49±1 50±2 51±3 51±1 52±1 52±2 chalose 40:60 T _α , °C 36±1 37±2 38±2 39±1	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _α , °C 36±1 36±1 37±1 38±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33% f, Hz 0.1 0.5 1.0 2.0 5.0	0.20 -0.49 -0.80 -1.10 -1.50 -1.80 logτ, s 0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T ₀ , °C 38±2 40±2 41±2 42±3 43±3	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _a , °C 38±1 39±2 40±1 41±2 42±2	50±3 50±2 51±2 52±3 53±1 53±2 Trystalline tree 60:40 T _a , °C 37±1 39±1 39±2 41±1 41±2	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60 T _a , °C 36±1 37±2 38±2 39±1 39±2	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _a , °C 36±1 36±1 37±1 38±2 38±2 38±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33% f, Hz 0.1 0.5 1.0 2.0	0.20 -0.49 -0.80 -1.10 -1.50 -1.80 logτ, s 0.20 -0.49 -0.80 -1.10	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T _a , °C 38±2 40±2 41±2 42±3	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _α , °C 38±1 39±2 40±1 41±2	50±3 50±2 51±2 52±3 53±1 53±2 Trystalline tree 60:40 T _α , °C 37±1 39±1 39±2 41±1	49±1 50±2 51±3 51±1 52±1 52±2 chalose 40:60 T _α , °C 36±1 37±2 38±2 39±1	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _α , °C 36±1 36±1 37±1 38±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33% f, Hz 0.1 0.5 1.0 2.0 5.0	0.20 -0.49 -0.80 -1.10 -1.50 -1.80 logτ, s 0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T ₀ , °C 38±2 40±2 41±2 42±3 43±3	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _a , °C 38±1 39±2 40±1 41±2 42±2	50±3 50±2 51±2 52±3 53±1 53±2 rystalline tre 60:40 T _a , °C 37±1 39±1 39±2 41±1 41±2 41±2 41±2	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60 T _α , °C 36±1 37±2 38±2 39±1 39±2 40±2	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _a , °C 36±1 36±1 37±1 38±2 38±2 38±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33% f, Hz 0.1 0.5 1.0 2.0 5.0 10.0	0.20 -0.49 -0.80 -1.10 -1.50 -1.80 logτ, s 0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T ₀ , °C 38±2 40±2 41±2 42±3 43±3	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _α , °C 38±1 39±2 40±1 41±2 42±2 43±1	50±3 50±2 51±2 52±3 53±1 53±2 Erystalline tree 60:40 T _α , °C 37±1 39±1 39±2 41±1 41±2 41±2	49±1 50±2 51±3 51±1 52±1 52±2 Shalose 40:60 T _α , °C 36±1 37±2 38±2 39±1 39±2 40±2	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _a , °C 36±1 36±1 37±1 38±2 38±2 38±2		
0.1 0.5 1.0 2.0 5.0 10.0 RH 33% f, Hz 0.1 0.5 1.0 2.0 5.0 10.0	0.20 -0.49 -0.80 -1.10 -1.50 -1.80 logτ, s 0.20 -0.49 -0.80 -1.10 -1.50	52±1 53±1 53±2 54±2 55±1 57±2 100:0 T _o , °C 38±2 40±2 41±2 42±3 43±3 44±2	51±2 52±3 52±2 53±2 54±3 55±3 Amorphous:C 80:20 T _a , °C 38±1 39±2 40±1 41±2 42±2 43±1 Amorphous:C	50±3 50±2 51±2 52±3 53±1 53±2 rystalline tre 60:40 T _a , °C 37±1 39±1 39±2 41±1 41±2 41±2 41±2	49±1 50±2 51±3 51±1 52±1 52±2 chalose 40:60 T _a , °C 36±1 37±2 38±2 39±1 39±2 40±2 nalose	47±2 47±1 48±2 48±2 49±2 50±2 20:80 T _α , °C 36±1 36±1 37±1 38±2 38±2 39±2		

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0.5	-0.49	19±1	19±2	19±1	19±1	18±2			
1.0	-0.80	20±2	20±1	20±2	20±2	19±1			
2.0	-1.10	21±1	21±1	21±2	21±2	20±1			
5.0	-1.50	23±1	22 ± 1	23±1	22 ± 1	20±1			
10.0	-1.80	25 ± 1	24 ± 2	23 ± 2	23±1	22 ± 2			

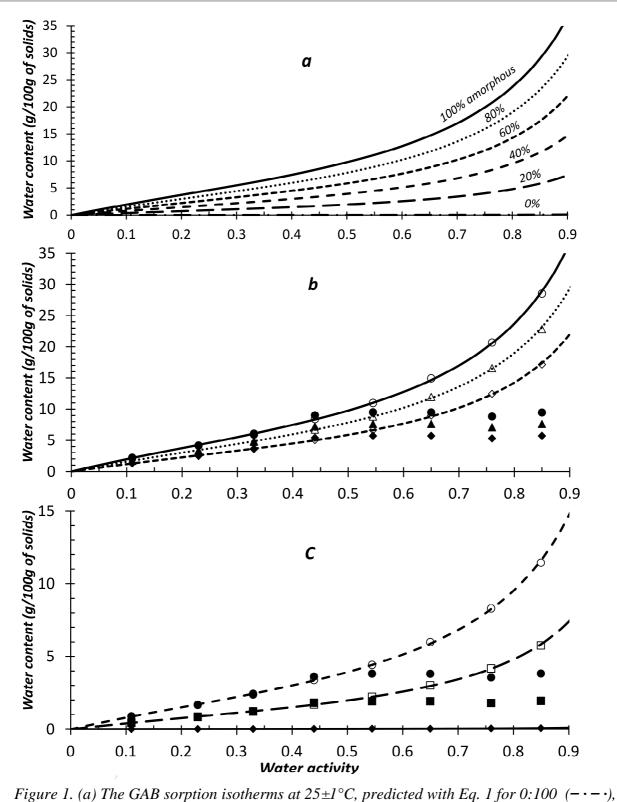
Table 4. Calculated WLF ($\tau_s = 100s$; $\eta_s = 10^{12}$ Pa s) constants C_1 and C_2 for partially crystalline trehalose systems, stored at various relative humidities (RH).

Amorphous:	RH 0%		RH 11%		RH 23%		RH 33%		RH 44%	
Crystalline Trehalose	$-C_1$, s	-C ₂ , •C	$-C_1$, s	-C ₂ , •C	$-C_1$, s	<i>-C</i> ₂ , <i>•C</i>	$-C_{I}$, s	-C ₂ , •C	$-C_1$, s	<i>-C</i> ₂ , <i>•C</i>
100:0	5.0±0.3	40.7±0.5	23.7±0.5	123.8±2.4	3.8±0.9	25.6±1.7	31.8±2.6	111.4±2.3	-10.0±0.7	-14.6±1.1
80:20	5.5±0.3	38.7±0.6	27.4±1.5	104.0 ± 3.1	3.2±1.1	21.0±1.5	11.9±1.3	43.6±1.2	-9.6±0.5	-12.5±1.0
60:40	15.1±0.3	61.0±0.5	20.9±0.7	70.8±1.3	3.8±1.4	22.1±0.7	24.0±2.1	71.5±1.6	-9.5±0.9	-12.3±0.7
40:60	11.9±0.2	43.6±1.2	36.4±1.1	101.1±1.7	2.8±0.6	17.4±1.7	10.7±0.9	33.9±2.9	-11.1±1.3	-15.0±0.9
20:80	44.6±0.2	109.6±2.7	9.1±0.6	28.4±2.1	4.6±1.3	18.8±1.0	8.9±1.1	25.9±1.4	-13.4±1.6	-18.8±2.0

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Table 5. "Strength" S (calculated at d=4) for partially crystalline trehalose systems at all ratios (amorphous:crystalline) stored at different relative humidities (RH).

Amorphous: Crystalline Trehalose	RH 0%	RH11%	RH 23%	RH 33%	RH 44%
100:0	18.1±2.9	17.9 ± 2.1	13.2±1.8	12.5±1.2	9.7±1.1
80:20	16.2 ± 2.5	13.2 ± 1.9	11.6±1.3	10.9±1.1	8.9 ± 1.4
60:40	12.8 ± 2.3	11.4±1.1	11.4 ± 1.1	10.2±2.1	9.0 ± 0.4
40:60	10.9 ± 1.7	10.0 ± 1.2	10.3 ± 0.9	9.2 ± 1.3	8.4 ± 0.5
20:80	9.0 ± 0.9	8.7 ± 0.4	8.7 ± 0.5	8.1±0.7	8.0 ± 0.7



20:80 (— —), 40:60 (——), 60:40 (——), 80:20 (——) and 100:0 (——) partially crystalline trehalose; comparison of modeled (lines), experimental data (solid symbols) and predicted by Eq. 1 (empty symbols): (b) for 100:0 (\bullet , \circ), 80:20 (\bullet , $^{\triangle}$), 60:40 (\bullet , $^{\triangle}$) partially crystalline

systems; (c) for 40:60 (\bullet , \circ), 20:80 (\bullet , \circ), 0:100 (\bullet) partially crystalline systems.

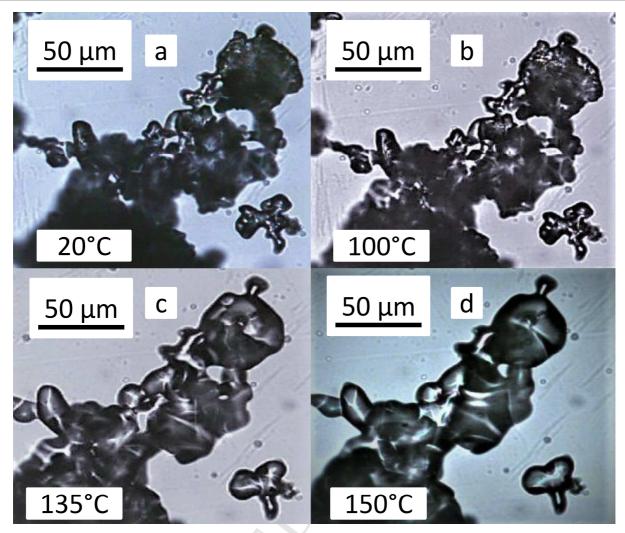


Figure 2. Microscope observation of 60:40 amorphous:crystalline trehalose, stored at 0.11 a_w , scanned at 2 K min⁻¹ at the temperature of (a) $20^{\circ}C$; (b) $100^{\circ}C$; (c) $135^{\circ}C$ and (d) $150^{\circ}C$.

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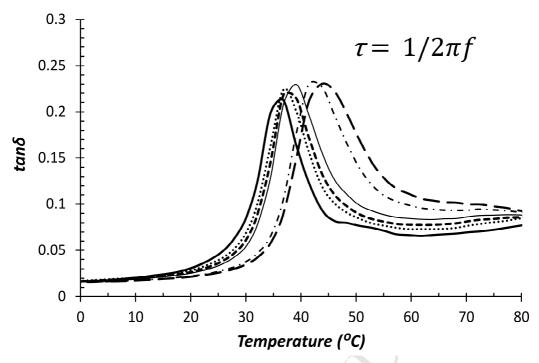


Figure 3. Temperature dependence of tangents δ for humidified (0.23 a_w) 80:20 amorphous:crystalline trehalose system determined by DMA in multi-frequency mode 0.1 Hz (---), 0.5 Hz (----), 1 Hz (----), 2 Hz (----), 5 Hz (-----), 10 Hz (-----).

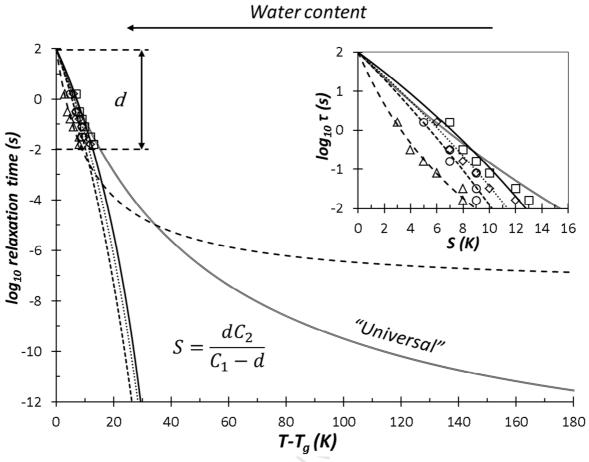


Figure 4. Modified WLF curves (lines), experimental data (symbols) and "strength" plot (inset) for 60:40 amorphous:crystalline trehalose system humidified at different relative humidities (0 (—, $^{\circ}$), 11 (…, $^{\diamond}$), 33 (---, $^{\circ}$), 44% RH (---, $^{\triangle}$)) with different water content (0, 1.3, 3.7, 5.3 (g/100g of dry solids), respectively).

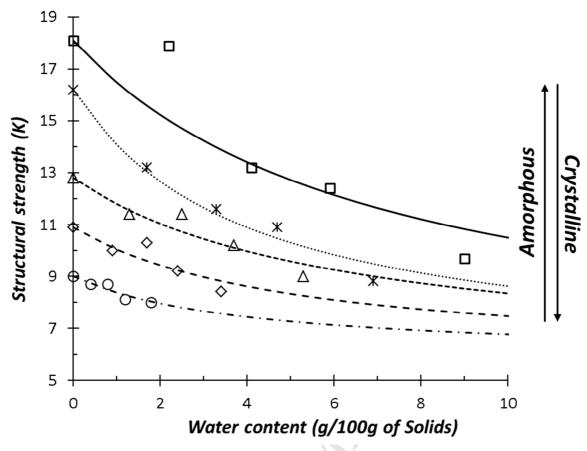


Figure 5. Strength curves, modeled by Eq. 7 (lines) and experimental (symbols) S of partially crystalline trehalose systems (100:0 (—, $^{\circ}$), 80:20 (·······, $^{\times}$), 60:40 (---, $^{\triangle}$), 40:60 (---, $^{\diamond}$), and 20:80 (---, $^{\circ}$) amorphous:crystalline ratios) at different water contents (g/100g of dry solids).

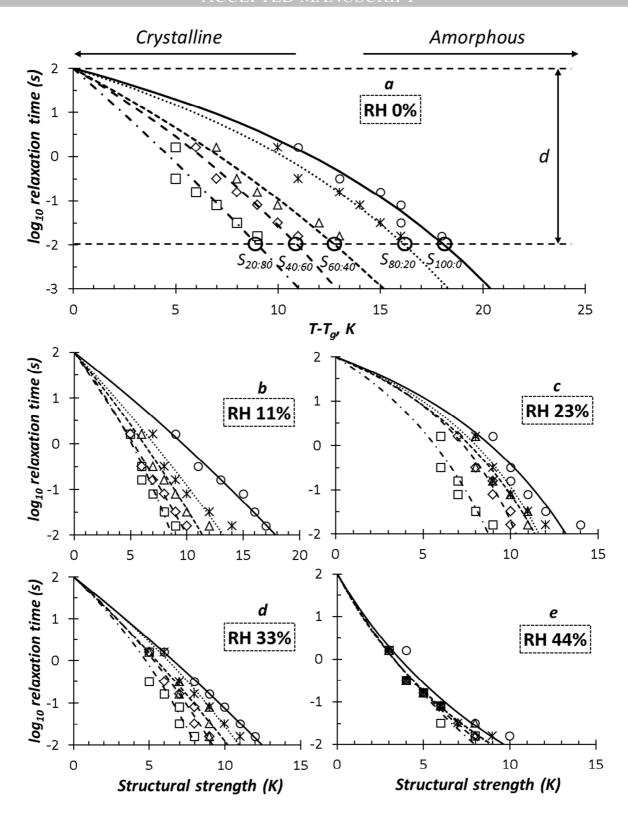
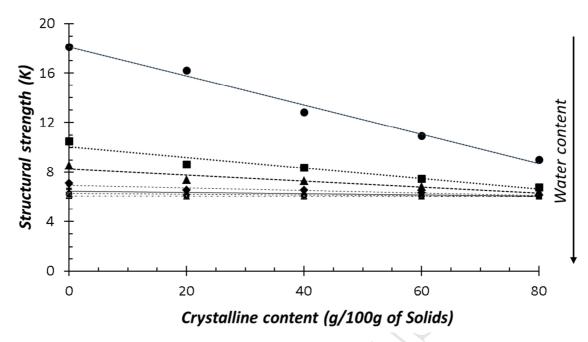


Figure 6. Modified WLF curves (lines) and experimental data (symbols) for partially crystalline trehalose (100:0 (—, \circ), 80:20 (······, \mathcal{K}), 60:40 (---, $^{\triangle}$), 40:60 (---, $^{\triangle}$) and 20:80 (---, $^{\circ}$) amorphous:crystalline ratios) stored 120 h (at 25±1°C) at different relative humidities (a (RH 0%), b (RH 11%), c (RH 23%), d (RH 33%) and e (RH 44%).



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Figure 7. Structural strength versus crystalline component content of partially crystalline trehalose (%) at different water contents (the data predicted by Eq. 7): 0% (\bullet), 10% (\bullet), 20% (\triangle), 40% (\diamond), 50% (X), 60% (X), 70% (\diamondsuit), 80% (\circ) and 90% (\triangle).

The concept of strength was developed using partially crystalline trehalose systems.
Strength shows linear decreasing with increasing of crystalline content in a system.
Fractional water sorption analysis was used in partially crystalline systems.
DSC shows crystalline phase-independent glass transition temperature.

The water content dependence on strength was shown.