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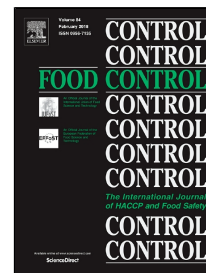
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- MRS was supplemented with phenylalanine or glycerol and fermented by *Lactobacillus reuteri* R29
- Accumulation of phenyllactic acid and reuterin was significantly increased
- Antifungal activity *in vitro* against common food spoilage fungi was significantly increased
- Antifungal performance of PLA was transferable in a bread system
- Antifungal activity of reuterin was not transferable in a bread system

Fundamental study on the improvement of the antifungal activity of *Lactobacillus reuteri* R29 through increased production of phenyllactic acid and reuterin

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ABSTRACT

Lactic acid bacteria have shown great potential as bio-preservative agents to maintain high food quality and safety. The strain *Lactobacillus reuteri* R29 is reported to have a broad spectrum of antifungal activity and suitability for application in food systems. Its antifungal activity is predominantly based on phenyllactic acid (PLA). Furthermore, it is potentially a producer of reuterin, a potent antimicrobial agent. This study focused on increasing the antifungal activity *in vitro* by supplementation of the growth medium with phenylalanine and glycerol to increase the yield of PLA and reuterin, respectively. For PLA, the addition of 1.5% phenylalanine (w/v) to MRS, resulted in significantly increased accumulation of PLA and antifungal performance against *Fusarium culmorum*. Supplementation of MRS with 500mM glycerol combined with a reduced glucose content (1.5%) showed the highest reuterin accumulation combined with fungal inhibition. To investigate the antifungal performance *in situ*, these cell-free supernatants (cfs) were applied in a bread system. The application of PLA-enriched cfs resulted in significantly extended (4 days) microbial shelf life compared to the control. The reuterin-enriched medium did not lead to significant shelf life extension. In conclusion, *Lactobacillus reuteri* R29 and its PLA-enriched cfs were found to be very promising alternatives for food bio-preservation.

Keywords: *Lactobacillus reuteri* R29, phenyllactic acid, reuterin, antifungal, bread making

1 Introduction

Lactic acid bacteria (LAB) are key microorganisms used for the production of various bakery products. They substantially improve nutritional and technological properties, as well as the flavour. However, current research is predominantly focused on their antimicrobial properties and potential applications as food bio-preservatives (Dalié, Deschamps, & Richard-Forget, 2010). The use of such LAB has the potential to replace chemical preservatives, such as calcium propionate, which would allow a “clean label” and hence lead to higher consumer acceptance (Pawlowska, Zannini, Coffey, & Arendt, 2012). In recent years, several strains of different genera have been found to express antimicrobial activity *in vitro* and *in situ* against food spoilage fungi (Axel et al., 2016; Crowley, Mahony, & Van Sinderen, 2013).

The antifungal activity of LAB is predominantly based on complex synergistic effects between several compounds. The production of these compounds is highly dependent on the strain and growth substrate. The exact interactions between the bacterial metabolites which create this synergistic effect are not yet fully understood. However, several metabolites that contribute to the antifungal activity have been identified and characterised. These metabolites, amongst others, include carboxylic acids (Ryan, Dal Bello, & Arendt, 2008) and 3-hydroxypropionaldehyde (3-HPA) also known as reuterin (Lüthi-Peng, Dileme, & Puhani, 2002).

Recently, the production and activity of 3-phenyllactic acid (PLA) has received great attention (Mu, Yu, Zhu, Zhang, & Jiang, 2012). This broad-spectrum antimicrobial compound originates in LAB fermented products from the catabolism of phenylalanine (Phe). Thereby, the Phe first undergoes a transamination by transferring the amino group onto a keto-acid acceptor. The synthesised

phenylpyruvic acid is then reduced to PLA by a dehydrogenase (Vermeulen, Gänzle, & Vogel, 2006). The antifungal activity of PLA is also dependent on a synergistic mechanism with other bacterial metabolites. Nevertheless, if the metabolic pathway yielding PLA is promoted, the antifungal activity is likely to increase. This could expand the field of application for LAB as bio-preservatives.

A very promising antimicrobial agent is the multicomponent system called reuterin, which results from the conversion of glycerol to 3-HPA (Engels et al., 2016). The antimicrobial activity of reuterin is, according to Schaefer et al. (2010), mainly based on its reactivity with free thiol groups of proteins, inducing oxidative stress in the target cells. It is understood that 3-HPA is released upon enzymatic dehydration of glycerol. Recent research conducted by Engels et al. (2016) suggests a rapid *in situ* conversion to acrolein, which is mainly responsible for the antimicrobial activity of the reuterin system. However, if sufficient amounts of carbohydrates, in particular glucose, are available, 3-HPA is further reduced to 1,3-propanediol which has no antimicrobial activity (Gänzle, 2015). Unfortunately, the research on reuterin so far has mainly focused on its industrial use as precursor for the synthesis of acrolein, not on food applications. To date, there are only two studies conducted by Gomez-Torres et al. (2014) and Ortiz-Rivera et al. (2017) available, investigating a possible application of reuterin as an antibacterial agent in a food system. As Gänzle (2015) observed, the applicability of reuterin, particularly in heat treated food systems, remains unclear. Further research is required, in particular to elucidate the effect of glycerol on bacterial growth and reuterin production.

A strain previously reported as demonstrating strong antifungal activity and suitability for food applications is *Lactobacillus reuteri* R29 (Axel et al., 2016; Oliveira et al.,

2015). Oliveira et al. (2015) demonstrated that the cell-free supernatant (cfs) of medium fermented for 48 h with this strain showed antifungal activity against *F. culmorum*, with PLA being the predominant antimicrobial compound produced. As it belongs to the species *L. reuteri*, the strain is likely to be a reuterin producer. To date there have been no studies carried out investigating the ability of *L. reuteri* R29 to produce and accumulate reuterin. Thus, the enhancement of the production and accumulation of antifungal compounds, namely reuterin and PLA, in the bacterial cfs represents a promising opportunity in particular, if this antifungal activity is applicable in food systems.

Therefore, the aim of this study was to increase the *in vitro* antifungal activity of *L. reuteri* R29 cfs. Different variations of the fermentation medium composition were investigated to increase the accumulation of PLA and reuterin. *F. culmorum* was used as indicator mould to determine the antifungal activity *in vitro*. Secondly, characterisations regarding the heat stability of the most efficient cfs were evaluated. Finally, antifungal cfs delivering the best results were applied in the bread system to obtain information regarding suitability for application in a food matrix. This study provides important information regarding the potential of *L. reuteri* R29 cfs as antifungal agent and its application as a bio-preservative agent, in the bread system.

2 Materials and Methods

2.1 Materials

Ingredients for microbiological media, chemicals and analytical standards used in this study were at least analytical grade and obtained from Sigma-Aldrich (Dublin, Ireland), unless otherwise stated. The analytical standards had a purity of $\geq 95\%$.

Wheat flour (bakers flour, Odlums, Dublin, Ireland), dry yeast (Puratos, Groot-Bijgaarden, Belgium), salt (Glacia British Salt Limited, Cheshire, UK), commercially available sunflower oil, sodium stearoyl lactylate (SSL) (Danisco, Denmark) and ascorbic acid (Storefast solutions, UK) were used for the baking trials.

2.2 Cultures, media and growth conditions

Fusarium culmorum TMW4.2043, *Aspergillus niger* FST4.21, *Penicillium expansum* FST 4.22 originated from the culture collection of School of Food and Nutritional Sciences, University College Cork (Cork, Ireland). Fungal spore suspensions were prepared according to the method described by (Oliveira, Mauch, Jacob, Waters, & Arendt, 2012).

Lactobacillus reuteri R29, originally isolated from human intestine, was obtained from the UCC culture collection (School of Food and Nutritional Sciences, University College Cork, Cork, Ireland). For long term storage, the culture was kept in commercial de Man Rogosa and Sharpe (MRS) broth containing 40% glycerol at -80°C. The culture was routinely refreshed on MRS agar plates (Fulka Chemie AG, Buchs, Switzerland) under anaerobic conditions at 37°C for 48 h.

2.3 Production of LAB cell-free supernatants

For production of 1-step fermented LAB cfs, a single colony, grown as described in 2.2, was inoculated in 5 mL MRS broth for 24 h at 37°C. Subsequently, a 1% inoculum into fresh fermentation medium (Table 1 and 2) was prepared and incubated at 37°C for the designated time. After fermentation, cell cultures were centrifuged (5,000 g, 15 min, 4°C) and the supernatant was sterile filtered through a 0.22 µm filter. The cfs obtained was stored at -20°C until further use.

The 2-step fermentation process was based on the method described by Doleyres et al. (2005) with the following modifications. After the first fermentation step, cells were washed and resuspended in sterile distilled water, containing 400 mM glycerol, at a final concentration of 10^9 CFU/mL. The suspension was incubated for 2 h at 30°C and the cfs collected as described above.

2.4 Evaluation of *in vitro* antifungal activity and determination of minimal inhibitory concentration (MIC₉₀)

Pre-trials were carried out to determine the ideal fungal spore concentration to investigate the increase of antifungal performance in the respective cell-free supernatants. Thereby, a concentration of 10^6 spores/mL as applied in the assay described were chosen for the subsequent experiments (data not shown). To determine the *in vitro* antifungal activity, 1 mL of *Fusarium culmorum*, *Aspergillus niger* or *Penicillium expansum* spore solution (containing approximately 10^6 spores) was transferred into a 2 mL microcentrifuge tube and centrifuged at 3,000 g (10 min, 4°C). The supernatant was discarded and the spore pellet resuspended in 1.0 mL of diluted LAB cfs (10%, v/v, in unfermented MRS). Subsequently, 200 µL aliquots were pipetted into a 96-well microplate (Sarstedt AG and Co, Nurembrecht, Germany). To control for the effect of condensation, each supernatant was also inoculated with 1 mL of sterile synthetic nutrient-poor bouillon, instead of fungal spore solution (blank). The microplate was sealed with an optically clear seal for Q-PCR (Thermo Scientific, Waltham, USA) and incubated in a Multiskan FC microplate-reader (Thermo Scientific, Waltham, USA) for 5 days at 25°C. The optical density was recorded at 620 nm (OD₆₂₀) every 2 h, with agitation in 4 s intervals. Values of each cfs inoculated with spore solution were corrected by the respective mediums blank

value. Antifungal activity was calculated as the percentage of OD reduction compared to the respective 0 h fermented medium and expressed as “percentage of growth inhibition”.

To determine the MIC₉₀ of PLA the substance was added to unfermented MRS-broth to a final concentration of 20,000 ppm and the pH adjusted to match the fermented medium. Due to the lack of a commercially available standard for reuterin, a serial dilution of the 2-step fermented supernatant was used to determine the MIC₉₀. The MIC₉₀ was defined as the minimal concentration of the respective antifungal compound required to achieve at least 90% reduction of fungal growth after 120 h under the assay conditions compared to a control medium without the antifungal compound.

2.5 Determination of antifungal fermentation products

2.5.1 Determination of carboxylic antifungal compounds

An Agilent 1260 high-performance liquid chromatography system equipped with an ultraviolet-diode array detector (UV/DAD) was used to quantify antifungal carboxylic compounds. Standard calibration curves for lactic and acetic acid (2 – 32 mM), as well as 13 phenolic compounds (catechol, hydroxyphenyllactic acid, 4-hydroxybenzoic acid, hydrocaffeic acid, caffeic acid, phloretic acid, hydroferulic acid, p-coumaric acid, ferulic acid, benzoic acid, salicylic acid, hydrocinnamic acid, methylcinnamic acid, vanillic acid and 3-phenyllactic acid) were prepared, using five different concentrations in duplicate. Calibration curves showed good linearity with correlation coefficients of ≥ 0.999 for all compounds. Extraction of antifungal organic acids (acetic acid and lactic acid) was carried out as described by Axel et al. (2016). For separation of the acids a Hi-Plex H Column (300×7.7 mm, 8 μ m; Agilent, Cork, Ireland), equipped with a guard column (50×7.7 mm, 8 μ m; Agilent, Cork, Ireland)

was used. Setting the UV/DAD at 210 nm, lactic and acetic acids in the cfs were determined after elution with 0.004 M sulphuric acid at 65 °C with a flow of 0.5 mL/min. Injection volume used was 20 µL.

Antifungal phenolic compounds were extracted, using a QuEChERS approach, as described by Peyer et al. (2016). Separation was carried out using a Gemini C₁₈ column (150 x 2.0 mm, 5 µm; Phenomenex, Macclesfield, UK), equipped with a guard column (AF0-8497; Phenomenex, Macclesfield, UK). The mobile phase consisted of A) H₂O with 0.1% FA and B) ACN with 0.1% FA. To ensure separation from other compounds, a gradient flow was performed (0 – 5 min – 90% A, 30 min – 80% A, 35 min – 60% A, 45 min – 5% A, 45 – 70 min – 90% A), using a flow rate of 0.2 mL/min at a temperature of 30°C. The volume injected was 10 µL and detection carried out at a wavelength of 210 nm.

If necessary, samples were diluted prior to extraction to ensure peak areas were within the calibration range. Identity of the peaks evaluated in the cfs samples was confirmed as PLA by comparison of the UV/VIS spectrum with the standard solution. The limit of detection (LOD) and the limit of quantification (LOQ) were determined from the signal/ noise (s/n) ratio. The LOD was set for s/n of 3:1 and the LOQ was set for s/n of 10:1. Recovery rates of the phenolic compounds were done with 3.0 mg/L of each analyte added in MRS and in chemically acidified MRS (pH 3 with 0.1 N HCl), and ranged from 89.1% (hydrocinnamic acid) to 118.1% (vanillic acid) of the total spiked amount.

2.5.3 Colorimetric assay for reuterin quantification

The concentration of reuterin in the cfs of the 1- and 2-step fermentations was determined using the colorimetric assay described by Lüthi-Peng et al. (2002).

Calibration was carried out, using acrolein as standard in distilled water (0.1 – 6.0 mM) and MRS broth (0.2 – 10 mM). Both calibration curves showed good linearity with correlation coefficients of >0.999.

2.6 Heat stability of antifungal fermentation products

Heat stability of the antifungal supernatants was determined by subjecting the cfs to 100°C for 1 h. Subsequently, samples were cooled immediately and kept in the fridge until further use. Control samples (not heat treated) were kept in the fridge at all times. Analysis of *in vitro* antifungal activity and quantification of antifungal compounds was carried out as described in sections 2.4 and 2.5, respectively but only *F. culmorum* was used as indicator mould.

2.7 Application of cfs in bread system and shelf life analysis

For baking trials, water in the recipe was replaced by the respective LAB cell-free supernatant. For preparation of breads 63% cfs, 3% yeast, 3% oil, 2% salt, 0.5% SSL and 0.1% ascorbic acid (each based on flour weight) were used. Reactivation of the yeast and dough preparation was carried out according to Heitmann et al. (2015). The dough was then divided into 65 g portions, placed in non-stick baking tins (dimensions: top inside, 50 mm x 90 mm; bottom outside, 45 mm x 85 mm; inside depth, 30 mm; Sasa UK, Enfield Middlesex, UK) and proofed for 75 minutes (30°C, 85% RH). Doughs were transferred into the oven (MIWE, condo, Arnstein, Germany) and baked for 14 minutes at 210°C (top and bottom). Following baking the bread loaves were removed from the tins immediately and cooled to room temperature for at least 120 min before further use.

The microbial shelf life test was carried out according to Heitmann et al. (2015). The amount of fungal spoilage was visually recorded over 13 days for each slice, using a calliper to determine the fungal colony size. Based on the percentage of the total surface area of the slice where fungal outgrowth occurred, each slice was categorised every day as A) mould-free, B) <10% covered with mould, C) 11 – 25% covered with mould, D) 26 – 50% covered with mould or E) >50% covered with mould.

2.8 Statistical analysis

Baking trials and all analyses were carried out from three independent fermentations, analysing each sample in duplicate. Statistical analysis was performed using Minitab 17 software. Data points were checked for outliers (Grubb's test) and evaluation of significant differences was performed using one-way analysis of variances (ANOVA). All differences were considered significant at $P < 0.05$. Where F-values were significant, pairwise comparisons were carried out with the help of Tuckey Post Hoc test to describe the statistical significance between the different fermentation media and times.

3 Results and Discussion

3.1 Effect of phenylalanine on antifungal activity and production of phenyllactic acid

Fungal isolates of *F. culmorum*, *Aspergillus niger* and *Penicillium expansum* were used as indicator moulds to evaluate the impact of Phe supplementation on the antifungal activity *in vitro* and PLA production. The results obtained are summarised in Table 1. The sole addition of up to 2% Phe (w/v) to MRS broth had no noteworthy

effect on the fungal growth in the unfermented medium (0 h fermentation). The PLA quantities for all 0 h fermented samples were below the limit of detection. Thus, all media used for inoculation of the different fungal spores were generally suitable for their growth and any inhibition obtained in the fermented samples was due to the antifungal metabolites produced by *L. reuteri* R29. Concentrations of antifungal carboxylic and phenolic acids were determined for all samples analysed. However, no significant differences ($p < 0.05$), apart from the PLA concentration, was found between any of the samples fermented for equal amounts of time (data not shown).

After 24 h of fermentation, all samples were found to contain substantial amounts of PLA. In addition, the PLA quantities were in direct correlation to the amount of Phe initially added. This confirms the findings of Rodríguez et al. (2012), that more PLA is produced when more Phe is available during fermentation. The PLA level of the unsupplemented MRS at this point (24 h) was 38.4 ± 1.3 ppm. When challenged with the fungal spores (10^7 spores/mL) the antifungal activity of the cfs was relatively poor. After 120 h no noteworthy inhibition was detected against either of the three fungal isolates. The cfs obtained through fermentation of MRS+1.5% Phe and MRS+2% Phe resulted after 120 h of incubation in significantly ($p < 0.05$) better inhibition of fungal spore germination, compared to cfs produced from unsupplemented MRS. But, it needs to be mentioned that the supernatants presented stronger inhibition against *F. culmorum* than against *A. niger* and *P. expansum*. Hence, it can be concluded that *F. culmorum* is the most sensitive of the three fungal species towards PLA. Similarly, the concentration of PLA in the cfs was found to be 3 times higher compared to the control. Addition of 0.5% or 1.0% Phe was much less efficient in terms of PLA accumulation and antifungal activity.

269 Finally, after 48 h of fermentation, all cfs showed the best spore germination
270 inhibition for the respective medium. The fermentation of unsupplemented MRS
271 resulted in limited inhibition of fungal spore germination over 120 h of incubation.
272 Compared to the unfermented medium, spore inhibition rates of $18.4\pm1.4\%$,
273 $2.2\pm0.4\%$ and $3.1\pm0.4\%$ were found against *F. culmorum*, *A. niger* and *P. expansum*,
274 respectively. In contrast, the addition of 1.0% Phe to the medium led to substantially
275 improved antifungal activity ($60.3\pm3.2\%$) over 120 h when challenged against *F.*
276 *culmorum*. The most efficient medium composition, in terms of antifungal
277 performance, was the addition of 1.5% Phe. This cfs resulted in $84.1\pm3.2\%$ inhibition
278 of spore germination against *F. culmorum* after 120 h. When challenged against the
279 other fungal genera, spore germination inhibition was found to be less substantial,
280 yet significantly increased compared to the unsupplemented medium. In contrast, the
281 supplementation with higher amounts of Phe did not further improve the antifungal
282 performance. The addition of less than 1.0% Phe showed no positive effect on the
283 antifungal performance. In general, the quantities of PLA determined for each cfs
284 correlate well with its respective antifungal activity. Thus, up to the addition of 1.5%
285 Phe, the amounts of PLA increased in direct correlation with the level of Phe initially
286 added. But no further increase could be achieved by supplementation with higher
287 quantities of Phe. Furthermore, the concentration of PLA after 48 h of fermentation
288 was always the highest for the respective medium, compared with shorter
289 fermentation times. This also correlates with the findings of Oliveira et al. (2015) who
290 concluded that PLA is produced by resting cells. The reason behind this is the
291 promotion of amino acid metabolism under nitrogen poor conditions (Hazelwood,
292 Daran, van Maris, Pronk, & Dickinson, 2008), which yields the antifungal carboxylic
293 acids, like PLA. Thus, after consuming the majority of nitrogen during the initial

growth phase, the resting bacteria have to survive in a relatively nitrogen poor environment. This leads to increased catabolism of Phe and hence the high release of PLA.

Furthermore, it was evident that the antifungal activity of cfs obtained from *L. reuteri* R29 was clearly related to the amounts of PLA accumulated in the medium. For all supernatants tested, a significant increase in PLA yield also led to improved antifungal performance. These results are in correlation with previous findings of Oliveira et al. (2015), who reported the antimicrobial activity of PLA and its role as lead antifungal substance produced by *Lactobacillus reuteri* R29. Conversely, Axel et al. (2016) found that it is not necessarily the strain with the highest production of antifungal compounds that shows the best antifungal activity, demonstrating the key role of the type of the antifungal compounds and their synergistic effects. Nevertheless, the increased accumulation of antifungal active compounds by *L. reuteri* R29 improves the overall antifungal performance of this particular strain (Valerio, Di Biase, Lattanzio, & Lavermicocca, 2016). Thus, the increased accumulation of PLA, as the lead antifungal compound of *L. reuteri* R29, led to improved antifungal activity of the cfs. This clearly demonstrates the great potential of microbial produced PLA as an antifungal substance.

It has to be mentioned however, that MIC₉₀ values of synthetic PLA in acidified MRS against all three fungi tested are more than 10 times higher compared to the amounts detected in the bacterial supernatants showing fungal inhibition. MIC₉₀ values determined are 6,000 ppm against *F. culmorum* and 15,000 ppm against *A. niger* and *P. expansum* (data not shown). Considering the high concentration of 10⁷ fungal spores/mL these values are in good correlation with the findings of

Lavermicocca, Valerio, & Visconti (2003). After all, these findings highlight that PLA, although being of major importance for antifungal performance, is not the only component responsible. Instead the synergy of PLA with other bacterial metabolites resulting in antifungal performance is evident.

In conclusion, the addition of Phe to the bacterial growth medium resulted in a supernatant which exhibited improved antifungal activity *in vitro* against fungal spores belonging to the genera of *Fusarium*, *Aspergillus* and *Penicillium*. This effect was clearly related to the higher PLA accumulation during LAB fermentation. The capacity to transfer this functionality to a food system remains unclear and will be investigated further in sections 3.3 and 3.4. This would allow the exploitation of the antifungal nature of LAB, particularly in systems that are unsuitable for direct bacterial fermentation. Thus, these results highlight novel and promising perspectives for the use of LAB as bio-preservatives.

As the 48 h fermentation of MRS broth supplemented with 1.5% phenylalanine was found to be the best antifungal medium, it was carried forward for further investigation, as discussed in sections 3.3 and 3.4. The medium and supernatant will henceforth be referred to as PMRS and Pcfs, respectively.

3.2 Effect of glycerol and sugar content on antifungal activity and reuterin production

Another promising antimicrobial compound with possible application in food systems and produced by certain strains of *Lactobacillus reuteri*, is the multicomponent system reuterin, with 3-hydroxy propionaldehyde (3-HPA) being the main active component. However, to date no study has demonstrated the ability of *Lactobacillus reuteri* R29 to produce reuterin via the bioconversion of glycerol. Various

combinations of glycerol and glucose in the growth medium were tested, in order to achieve the highest possible reuterin yield and the best antifungal activity *in vitro* against *F. culmorum*. Results for the different cfs, fermented for 0, 24 and 48 h, respectively are summarised in Table 2.

Firstly, the results show that all the unfermented media (0 h fermentation) provide a suitable substrate for spore germination of *F. culmorum*. The growth curve obtained for each respective medium was used as reference, in order to evaluate the antifungal activity of the respective fermentation cfs. Furthermore, concentrations of antifungal carboxylic and phenolic acids were determined for all samples, showing no substantial differences between any of the 24 and 48 h fermented samples were found.

After 24 h of fermentation, the cfs obtained from unmodified MRS contained no detectable amounts of reuterin, as MRS broth contains no glycerol. However, MRS supplemented with 250 – 1000 mM glycerol and a glucose content of 1.0 – 1.5% led to cfs containing various concentrations of reuterin. Values determined ranged between 12.5 ± 0.1 mM (250 mM glycerol, 1.0% glucose) and 27.5 ± 0.6 mM (500 mM glycerol, 1.5% glucose). Any further increase of glucose in the medium resulted in significantly lower reuterin concentrations, as the data for the media containing 2% glucose (Table 2) demonstrate. This suggests that 1.5% glucose in the initial medium is the ideal amount required for *L. reuteri* R29 to accumulate reuterin. At this level of glucose, the bacteria are able to grow to high cell density in the medium, resulting in a high glycerol conversion to reuterin. However, this concentration is low enough to avoid expression of the 3-HPA reductase responsible for the reductive effect on antifungal activity by removing reuterin from the system through its

conversion to 1,3-propanediol. Comparison of reuterin contents for media containing similar amounts of glycerol but varying glucose contents suggests that in presence of sufficient amounts of glucose the 3-HPA reductase is expressed already during the first 24 h of fermentation, preventing the accumulation of reuterin. The ideal amount of added glycerol was found to be 500 mM, as it resulted in the highest reuterin concentrations for each respective level of glucose in the medium. The amount of reuterin produced by *L. reuteri* R29 correlates well with the *in vitro* antifungal activity against fungal spores of *F. culmorum*, *A. niger* and *P. expansum*. As mentioned in section 3.1 the cfs obtained from normal MRS broth showed, due to the high spore concentration, very low levels of inhibition against the three fungi after 120 h of incubation. In contrast, the supernatants containing reuterin presented substantially better spore germination inhibition, demonstrating the antifungal potential of reuterin. The cfs with the highest reuterin concentration (500 mM glycerol, 1.5% glucose) showed total inhibition of *F. culmorum* against 10^7 spores/mL over 96 h of incubation (data not shown), and, after 120 h still had $69 \pm 2\%$ fungal inhibition. Similarly to the Pcfs, inhibition of *Aspergillus* and *Penicillium* spores was found to be less substantial but still significant compared to the control (unsupplemented MRS). In conclusion, it is evident from these results that a higher reuterin concentration results in enhanced antifungal performance of the cfs.

In contrast, after 48 h of fermentation, the amounts of reuterin and the antifungal performance for each supernatant decreased substantially, compared to the respective 24 h fermented medium. This may be due to the extremely high reactivity of reuterin. As MRS broth is a complex medium, it provides plenty of potential reaction opportunities for reuterin. Furthermore, if the fermentation time is too long the reuterin acts as a self-inhibiting antimicrobial compound against the LAB in the

medium (Lüthi-Peng et al., 2002). Although *L. reuteri* R29 was shown to be able to produce and accumulate reuterin, exact control of the fermentation parameters, such as time and medium composition is crucial. As a consequence, cfs obtained after 48 h of fermentation showed no significant difference in antifungal performance compared to the unsupplemented control.

Overall, the most efficient modification in terms of reuterin yield and *in vitro* antifungal activity against *F. culmorum* was obtained by 24 h fermentation of MRS supplemented with 500 mM glycerol and 1.5% glucose, instead of the conventional 2% glucose. This concentration appears to be the best compromise to allow sufficient bacterial growth and at the same time inhibit the expression of 3-HPA reductase. This correlates well with the findings of Lüthi-Peng et al. (2002). Consequently, the modification of the bacterial growth medium to induce reuterin production and accumulation was shown to be a promising method of improving the suitability of *L. reuteri* R29 as a bio-preservative agent.

In addition, the possibility of a combined increase in PLA and reuterin accumulation during LAB fermentation was attempted. However, this resulted in a total loss of antifungal activity (data not shown) of the cfs, due to the high reactivity of reuterin with the free amino-group of the Phe, needed as pre-cursor for PLA (Hazelwood et al., 2008).

As reuterin has industrial relevance as pre-cursor for the production of acrolein, several studies have been conducted to increase the reuterin production by LAB. These studies solved the problem regarding the carbohydrate content by application of a 2-step fermentation process. Although this process leads to high reuterin accumulation, with the cfs of the first fermentation step, also antifungal compounds

produced by *L. reuteri* R29 are removed. Thus, it is uncertain if this process ultimately leads to effective fungal inhibition when applied *in vitro*. Therefore, the 2-step fermentation method of Doleyres et al. (2005) was adopted and tested for both, yield of reuterin and *in vitro* antifungal activity. Here, a reuterin content of up to 93.1±2.0 mM was achieved. Although this is much lower than the previously reported 176 mM (Doleyres et al., 2005) it equates to a more than 3 fold increase compared to the yield in the 1 step fermentation procedure (27.5±0.6 mM). This relatively low level of reuterin, compared to Doleyres et al. (2005), is most likely due to the use of a different strain of *Lactobacillus reuteri*. When the cfs was applied *in vitro* in the antifungal assay, it resulted in a total inhibition of all fungi tested over 120 h of incubation. Consequently, the significantly increased yield of reuterin in the 2-step fermentation method fully compensated for the loss of carboxylic acids. In fact, the cfs produced by this method had the highest *in vitro* antifungal activity of all the treatments investigated in this study.

This 2-step fermented supernatant was further used to determine the MIC₉₀ of reuterin, as no commercial reuterin standards are available. Values found were 4 mM against *F. culmorum* and 8 mM against *A. niger* and *P. expansum* (data not shown). These values are in good correlation with the findings of Chung, Axelsson, Lindgren, & Dobrogosz (1989). Furthermore, it highlights the great potential of reuterin as antimicrobial and antifungal agent.

In conclusion, *Lactobacillus reuteri* R29 was shown to have the ability to metabolise glycerol to reuterin. This can be achieved using a 1 or 2 step fermentation procedure. Both approaches resulted in very high rates of fungal inhibition, when challenged against 10⁷ spores/mL over 120 h (Table 2). Thus, the modification of the

fermentation process of *L. reuteri* R29 towards the production of reuterin shows the potential for bio-preservation. But the need to control parameters such as glucose and glycerol content, and fermentation time carefully, to allow accumulation of reuterin has to be kept in mind. For a successful application in a food system, the high reactivity of reuterin might impose further challenges. This applies in particular, if heat treatment is involved, as this would further increase the reactivity of reuterin. Furthermore, there is the risk of acrolein formation due to thermal dehydration (Gänzle, 2015).

3.3 Heat stability of PLA and reuterin

In order to get a more detailed picture with respect to the possible applications of the various antifungal cell-free supernatants obtained in sections 3.1 and 3.2, three selected media were subjected to heat treatment. In particular for use in food systems, a certain heat resistance would broaden the range of possible applications immensely. Therefore, the impact of heat treatment (1 h at 100°C) on PLA and reuterin content, and on the antifungal performance *in vitro* was determined, using *F. culmorum* as indication strain.

For the Pcfs, no significant reduction of inhibitory capacity against *F. culmorum* was found. Even after 120 h the heat treated supernatant resulted in 80±3% of fungal growth inhibition, compared to 84±3% for the not heat treated cfs (Figure 1A). Chromatographic analysis of the heat treated supernatant further revealed no substantial loss in PLA, compared to the unheated one (data not shown). Consequently, this cfs can be considered as heat stable with regards to the antifungal performance and the concentration of its lead antifungal substance, PLA. This result is in good correlation with the findings of Cortés-Zavaleta, López-Malo,

Hernández-Mendoza, & García (2014). Thus, the fermentation of Phe enriched medium by *L. reuteri* R29 appears to be very promising for the production of antifungal cfs and use as a bio-preservative in food systems.

For the reuterin-containing supernatants different results were obtained, depending on the fermentation method. For the cfs produced with the 2-step fermentation procedure, no substantial reduction in antifungal performance (Figure 1B) or reuterin content (Figure 1C) was observed. However, heat treatment of the cfs produced by the 1-step fermentation procedure resulted in a complete loss of antifungal activity (Figure 1D). Likewise, the concentration of reuterin in the cfs decreased from 27.5 ± 0.6 mM to 0.5 ± 0.1 mM, due to the heating process (Figure 1C). The reason for these differences is in the constituents of the respective supernatant, as heating primarily increases the reactivity of reuterin (Vollenweider, Evers, Zurbriggen, & Lacroix, 2010). The 2-step fermentation produces a cfs that consists only of water, glycerol and reuterin, and thus no adequate reactive partner is present to sequester reuterin. Hence, the antifungal activity of this cfs was not notably effected by the heating process. On the other hand, the 1-step fermentation produces a supernatant, containing all the ingredients of MRS broth and several metabolites produced by the bacteria which are potential reactive partners for the reuterin. Consequently, the increase in reactivity due to heating resulted in a substantial decrease in reuterin content and hence severely reduced antifungal activity.

To examine this topic further and evaluate the antifungal activity *in situ*, the reuterin enriched (1- and 2-step fermentation process) and the Pcfs were applied into a model system for food production that involves heat treatment, the bread making process.

3.4 Effect of PLA and reuterin on bread microbial shelf life

In order to investigate the antifungal activity *in situ*, it was attempted to transfer and exploit the increased antifungal activity (*in vitro*) in the food processing chain, using bread making as an example. Therefore, the water in the dough preparation was replaced with the cell-free supernatants and the effect on microbial shelf life was tested by challenge against environmental fungi. Thus, use of the *in vitro* fermented cfs enabled comparison of the *in situ* antifungal performance of the different supernatants without the influence of other ingredients on the bacterial growth.

As demonstrated in previous studies the application of LAB fermented media into a cereal food matrix is possible (Le Lay et al., 2016; Peyer et al., 2016; Russo et al., 2015; Saladino et al., 2016). Furthermore, the routinely use of LAB as starter cultures for sourdough bread shows that the acidity has no substantial negative impact on the product quality and taste (Pawlowska et al., 2012). Hence, application of bacterial cfs, despite the strong acidic pH, as promising perspective for natural extension of microbial shelf-life.

As shown in Figure 2A, the use of cfs obtained by fermentation of normal MRS (24 h) resulted in a microbial shelf life of 5 days. After 13 days of storage 83% of the bread slices contained mould colonies (category B), whereas the remaining 17% were still completely mould free. A very similar result was achieved for the same medium, fermented for 48 h (Figure 2B). The microbial shelf life was determined to 4 days and after 13 days of storage 17% of the slices were still mould free. However, the mould was found to spread slightly faster, than with the 24 h fermented supernatant, as there were 6.5% of the slices in the “C” category (11 – 24% mouldy) and the rest in “B”. This outcome is interesting, as the antifungal activity *in vitro* was

evident after 48 h of fermentation. However, as the breads were challenged against environmental fungi and not just one specific strain, variations in the antifungal performance are possible. In particular, as the antifungal performance is based on a synergistic mechanism, the amount of organic acids produced cannot serve as sole indicator for antifungal activity. Thus, it is possible that the mixture of antifungal compounds present after 24 h is more effective against certain environmental fungi than after 48 h. These samples, served as controls in order to evaluate if the increased antifungal activity as evident in *in vitro* trials could also translated *in situ*.

In comparison, the Pcfs (24 h, Figure 2C) did not significantly increase the shelf life, compared to the respective control (MRS/24 h). The microbial shelf life was determined to be 5 days and after 13 days, 19% of the slices were still mould free. At the same time, 6.5% of the slices were mouldy to more than 10%. These differences to the 24 h fermented control are not statistically significant ($p < 0.05$). Thus, the 24 h fermentation of Phe enriched MRS did not increase the antifungal performance *in situ*. In contrast, after 48 h of fermentation, the Pcfs led to substantially increased shelf life (Figure 2D) when compared to the respective control. The shelf life increased from 4 to 8 days (100%), due to the use of Pcfs. In addition, the spread of fungal outgrowth was found to be notably retarded and the number of mouldy slices after 13 days of monitoring was, with 53%, significantly reduced also. This correlates well with the results of the *in vitro* assays for the Pcfs against *F. culmorum*. These results strengthen the conclusions previously made by Crowley et al. (2013), who reported that LAB are, due to their antimicrobial acids, very promising candidates as food bio-preservatives. In addition, this result demonstrates the broad antifungal activity of PLA against various spoilage organisms *in situ*. Thus, the results obtained in this study further prove the suitability of *L. reuteri* R29 as natural food

preservative, using bread making as an example of such an application. The use of *L. reuteri* R29 as bio-preservative, in particular with emphasis on high PLA production appears to be a promising alternative to conventional preservatives.

Despite showing less promising results *in vitro* after heat treatment, the reuterin containing 1- and 2-step fermentations were also applied in the baking process. The respective 24 h and 48 h 1-step fermented supernatants (Figures 2E and F, respectively) were found to result in a shelf life increase of 1 day, compared to their respective controls. Also the spread of the fungal outgrowth appeared to be slower. However, these improvements were not statistically significant ($p < 0.05$). Due to the high reactivity of reuterin, in particular at high temperatures as shown in section 3.3, it is likely that the reuterin reacted with other dough constituents. Thus, when the microbial challenge test was carried out, no noteworthy levels of active reuterin were present. This indicates that the breads obtained from this supernatant should perform similar to the control in terms of microbial shelf life, as the normally produced antifungal acids were present in both control and 1-step fermentation. Hence, the outcome of the shelf life test, where no significant difference to the control was found, is in good correlation with the previous results, as discussed in section 3.3.

Finally, the 2-step fermented cfs showed the least antifungal activity of all samples, when applied to the baking process (Figure 2G). The shelf life decreased substantially by 2 days, compared to the 24 h MRS control. The fungal spread was also much faster than in any other sample, resulting in 42% of the bread slices being covered to more than 50% by fungi (category "E"), after 13 days of monitoring. Thus, the cfs obtained from the 2-step fermentation was found to be completely unsuitable

for *in situ* microbial preservation in bread. The explanation for this, again, is related to the high reactivity of reuterin, in particular at high temperatures. Although this supernatant showed heat stability when tested *in vitro*, the dough matrix is very complex and thus provides plenty of reaction opportunities for the reuterin. In particular, free thiol groups which are the main target in the inactivated microbes present in the flour can interfere with the reuterins antimicrobial performance. Despite the fact that levels of free thiol groups in a dough formulation as it was used here are 10 – 100 times below the reuterin concentrations (Kohler, 2003; Reinbold, Rychlik, Asam, Wieser, & Koehler, 2008) they are likely to compromise the antifungal activity significantly (Engels et al., 2016). Furthermore, the production process of this cfs eliminates the other metabolites produced by *L. reuteri* R29, such as antifungal acids. As a consequence, by losing the reuterin due to reactions with other dough constituents, no active antifungal compounds were present anymore. Hence, the outcome of an even further decreased shelf life, compared to the control. The results of this study clearly demonstrate that reuterin is not suitable as food preservative if it is subjected to heating. However, it may still be suitable for unheated foods like salami or cheese, for which it has already been successfully applied (Gomez-Torres et al., 2014; Ortiz-Rivera et al., 2017).

4. CONCLUSIONS

In conclusion, this study demonstrates three possibilities for improving the efficiency of antifungal LAB *in vitro*, using *Lactobacillus reuteri* R29 as an example. Further understanding regarding production and stability of antifungal compounds was obtained. In particular, the key role of PLA for the antifungal performance of *L. reuteri* R29 became evident. However, from the MIC₉₀ values of synthetic PLA it also

became evident that microbial PLA just in synergy with other bacterial metabolites can serve as efficient antifungal agent. The results achieved *in vitro*, could only partly be transferred into the bread making process. Reuterin, due to its high reactivity, in particular at high temperatures (Vollenweider et al., 2010), did not lead to satisfactory results *in situ*. In contrast, the supplementation with Phe, to increase the production of PLA, was found to be very efficient in both *in vitro* and *in situ*. Hence, Phe supplemented fermentation media should be considered as promising options to improve the antimicrobial performance of LAB during production of food, such as bread or beverages. The proteolytic activity reported for *L. reuteri* R29 (Axel et al., 2016) also proposes the possibility to achieve this antifungal effect upon supplementation with Phe rich proteins. This work shows the potential for further exploitation of LAB as bio-preservatives, particularly in environments unsuitable for bacterial fermentation, for example, during grain storage. This demonstrates the potential to enlarge the field of application for the antimicrobial properties of LAB. Further research is required on the *in situ* production of PLA, including its stability and influence on sensory parameters. In addition, application of such methodologies investigated in this study in further food systems will serve to increase our knowledge in this increasingly pertinent area.

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REFERENCES

- Axel, C., Brosnan, B., Zannini, E., Peyer, L. C., Furey, A., Coffey, A., & Arendt, E. K. (2016). Antifungal activities of three different *Lactobacillus* species and their production of antifungal carboxylic acids in wheat sourdough. *Applied Microbiology and Biotechnology*, 100(4), 1701–1711. <http://doi.org/10.1007/s00253-015-7051-x>
- Chung, T. C., Axelsson, L., Lindgren, S. E., & Dobrogosz, W. J. (1989). In Vitro Studies on Reuterin Synthesis by *Lactobacillus reuteri* In Vitro Studies on Reuterin Synthesis by *Lactobacillus*. *Microbial Ecology in Health and Disease*, 2, 137–144.
- Cortés-Zavaleta, O., López-Malo, A., Hernández-Mendoza, A., & García, H. S. (2014). Antifungal activity of lactobacilli and its relationship with 3-phenyllactic acid production. *International Journal of Food Microbiology*, 173(December 2013), 30–35. <http://doi.org/10.1016/j.ijfoodmicro.2013.12.016>
- Crowley, S., Mahony, J., & Van Sinderen, D. (2013). Current perspectives on antifungal lactic acid bacteria as natural bio-preservatives. *Trends in Food Science and Technology*, 33(2), 93–109. <http://doi.org/10.1016/j.tifs.2013.07.004>
- Dalié, D. K. D., Deschamps, A. M., & Richard-Forget, F. (2010). Lactic acid bacteria - Potential for control of mould growth and mycotoxins: A review. *Food Control*, 21(4), 370–380. <http://doi.org/10.1016/j.foodcont.2009.07.011>
- Doleyres, Y., Beck, P., Vollenweider, S., & Lacroix, C. (2005). Production of 3-hydroxypropionaldehyde using a two-step process with *Lactobacillus reuteri*. *Applied Microbiology and Biotechnology*, 68(4), 467–474.

<http://doi.org/10.1007/s00253-005-1895-4>

Engels, C., Schwab, C., Zhang, J., Stevens, M., Bieri, C., Ebert, M.-O., ... Sturla, S. (2016). Acrolein contributes strongly to antimicrobial and heterocyclic amine transformation activities of reuterin. in revision. *Nature Publishing Group*, (August), 1–13. <http://doi.org/10.1038/srep36246>

Gänzle, M. G. (2015). Lactic metabolism revisited: Metabolism of lactic acid bacteria in food fermentations and food spoilage. *Current Opinion in Food Science*, 2(Figure 2), 106–117. <http://doi.org/10.1016/j.cofs.2015.03.001>

Gomez-Torres, N., Vila, M., Gaya, P., & Garde, S. (2014). Prevention of late blowing defect by reuterin produced in cheese by a *Lactobacillus reuteri* adjunct. *Food Microbiology*, 42(February), 82–88. <http://doi.org/10.1016/j.fm.2014.02.018>

Hazelwood, L. H., Daran, J.-M. G., van Maris, A. J. A., Pronk, J. T., & Dickinson, J. R. (2008). The Ehrlich Pathway for Fusel Alcohol Production: a Century of Research on *Saccharomyces cerevisiae* Metabolism. *Applied and Environmental Microbiology*, 74(8), 2259–2266. <http://doi.org/10.1128/AEM.02625-07>

Heitmann, M., Zannini, E., & Arendt, E. K. (2015). Impact of different beer yeasts on wheat dough and bread quality parameters. *Journal of Cereal Science*, 63, 49–56. <http://doi.org/10.1016/j.jcs.2015.02.008>

Kohler, P. (2003). Concentrations of Low and High Molecular Weight Thiols in Wheat Dough As Affected by Different Concentrations of. *Journal of Agricultural and Food Chemistry*, 51, 4948–4953.

Lavermicocca, P., Valerio, F., & Visconti, A. (2003). Antifungal Activity of

- 654 Phenyllactic Acid against Molds Isolated from Bakery Products, 69(1), 634–640.
655 <http://doi.org/10.1128/AEM.69.1.634>
- 656 Le Lay, C., Mounier, J., Vasseur, V., Weill, A., Le Blay, G., Barbier, G., & Coton, E.
657 (2016). In vitro and in situ screening of lactic acid bacteria and propionibacteria
658 antifungal activities against bakery product spoilage molds. *Food Control*, 60,
659 247–255. <http://doi.org/10.1016/j.foodcont.2015.07.034>
- 660 Lüthi-Peng, Q., Dileme, F. B., & Puhon, Z. (2002). Effect of glucose on glycerol
661 bioconversion by *Lactobacillus reuteri*. *Applied Microbiology and Biotechnology*,
662 59(2–3), 289–296. <http://doi.org/10.1007/s00253-002-1002-z>
- 663 Mu, W., Yu, S., Zhu, L., Zhang, T., & Jiang, B. (2012). Recent research on 3-
664 phenyllactic acid, a broad-spectrum antimicrobial compound. *Applied*
665 *Microbiology and Biotechnology*, 95(5), 1155–1163.
666 <http://doi.org/10.1007/s00253-012-4269-8>
- 667 Oliveira, P. M., Brosnan, B., Furey, A., Coffey, A., Zannini, E., & Arendt, E. K. (2015).
668 Lactic acid bacteria bioprotection applied to the malting process. Part I: Strain
669 characterization and identification of antifungal compounds. *Food Control*, 51,
670 433–443. <http://doi.org/10.1016/j.foodcont.2014.07.004>
- 671 Oliveira, P. M., Mauch, A., Jacob, F., Waters, D. M., & Arendt, E. K. (2012).
672 Fundamental study on the influence of *Fusarium* infection on quality and
673 ultrastructure of barley malt. *International Journal of Food Microbiology*, 156(1),
674 32–43. <http://doi.org/10.1016/j.ijfoodmicro.2012.02.019>
- 675 Ortiz-Rivera, Y., Sanchez-Vega, R., Gutierrez-Mendez, N., Leon-Felix, J., Acosta-
676 Muniz, C., & Sepulveda, D. R. (2017). Production of reuterin in a fermented milk

product by *Lactobacillus reuteri* : Inhibition of pathogens , spoilage
microorganisms , and lactic acid bacteria. *Journal of Dairy Science*, 100(6),
4258–4268. <http://doi.org/10.3168/jds.2016-11534>

Pawlowska, A. M., Zannini, E., Coffey, A., & Arendt, E. K. (2012). “Green
Preservatives”: Combating Fungi in the Food and Feed Industry by Applying
Antifungal Lactic Acid Bacteria. *Advances in Food and Nutrition Research*,
66(November 2016), 217–238. [http://doi.org/10.1016/B978-0-12-394597-](http://doi.org/10.1016/B978-0-12-394597-6.00005-7)
6.00005-7

Peyer, L. C., Axel, C., Lynch, K. M., Zannini, E., Jacob, F., & Arendt, E. K. (2016).
Inhibition of *Fusarium culmorum* by carboxylic acids released from lactic acid
bacteria in a barley malt substrate. *Food Control*, 69, 227–236.
<http://doi.org/10.1016/j.foodcont.2016.05.010>

Reinbold, J., Rychlik, M., Asam, S., Wieser, H., & Koehler, P. (2008). Concentrations
of total glutathione and cysteine in wheat flour as affected by sulfur deficiency
and correlation to quality parameters. *Journal of Agricultural and Food
Chemistry*, 56(16), 6844–6850. <http://doi.org/10.1021/jf800880n>

Rodríguez, N., Salgado, J. M., Cortés, S., & Domínguez, J. M. (2012). Antimicrobial
activity of d-3-phenyllactic acid produced by fed-batch process against
Salmonella enterica. *Food Control*, 25(1), 274–284.
<http://doi.org/10.1016/j.foodcont.2011.10.042>

Russo, P., Arena, M. P., Fiocco, D., Capozzi, V., Drider, D., & Spano, G. (2015).
Lactobacillus plantarum with broad antifungal activity: A promising approach to
increase safety and shelf-life of cereal-based products. *International Journal of*

Food Microbiology. <http://doi.org/10.1016/j.ijfoodmicro.2016.04.027>

Ryan, L. A. M., Dal Bello, F., & Arendt, E. K. (2008). The use of sourdough fermented by antifungal LAB to reduce the amount of calcium propionate in bread. *International Journal of Food Microbiology*, 125(3), 274–278. <http://doi.org/10.1016/j.ijfoodmicro.2008.04.013>

Saladino, F., Luz, C., Manyes, L., Fernández-Franzón, M., & Meca, G. (2016). In vitro antifungal activity of lactic acid bacteria against mycotoxigenic fungi and their application in loaf bread shelf life improvement. *Food Control*, 67, 273–277. <http://doi.org/10.1016/j.foodcont.2016.03.012>

Schaefer, L., Auchtung, T. A., Hermans, K. E., Whitehead, D., Borhan, B., & Britton, R. A. (2010). The antimicrobial compound reuterin (3-hydroxypropionaldehyde) induces oxidative stress via interaction with thiol groups. *Microbiology*, 156(6), 1589–1599. <http://doi.org/10.1099/mic.0.035642-0>

Valerio, F., Di Biase, M., Lattanzio, V. M. T., & Lavermicocca, P. (2016). Improvement of the antifungal activity of lactic acid bacteria by addition to the growth medium of phenylpyruvic acid, a precursor of phenyllactic acid. *International Journal of Food Microbiology*, 222(January), 1–7. <http://doi.org/10.1016/j.ijfoodmicro.2016.01.011>

Vermeulen, N., Gánzle, M. G., & Vogel, R. F. (2006). Influence of peptide supply and cosubstrates on phenylalanine metabolism of *Lactobacillus sanfranciscensis* DSM20451T and *Lactobacillus plantarum* TMW1.468. *Journal of Agricultural and Food Chemistry*, 54(11), 3832–3839. <http://doi.org/10.1021/jf052733e>

Vollenweider, S., Evers, S., Zurbruggen, K., & Lacroix, C. (2010). Unraveling the

723 hydroxypropionaldehyde (HPA) system: An active antimicrobial agent against
724 human pathogens. *Journal of Agricultural and Food Chemistry*, 58(19), 10315–
725 10322. <http://doi.org/10.1021/jf1010897>

726 **Table 1: Concentration (ppm) of phenyllactic acid (PLA) and antifungal activity over 120 h (% of growth inhibition) of cell-free supernatants of**
727 ***Lactobacillus reuteri* R29 grown in MRS for 0, 24 or 48 h at 37°C in presence of various amounts of phenylalanine.#**

Medium	fermentation time [h]	PLA [ppm]	antifungal activity after 120 hours of incubation [% growth inhibition against 10 ⁷ spores/mL]		
			<i>P. expansum</i>	<i>A. niger</i>	<i>F. culmorum</i>
MRS	0	_a	_a	_a	_a
	24	38.4±1.3 ^b	1.2±0.2 ^b	0.2±0.1 ^a	1.5±0.2
	48	85.3 ±1.1	3.1±0.4 ^c	2.2±0.4 ^b	18.4±1.4 ^c
MRS + 0.5% Phe	0	_a	_a	_a	_a
	24	41.6±1.8 ^b	1.4±0.1 ^b	3.2±0.5 ^{bc}	3.4±2.2 ^{bc}
	48	103.2±2.3	3.6±0.3 ^c	3.7±1.4 ^c	14.1±2.8 ^c
MRS + 1.0% Phe	0	_a	_a	_a	_a
	24	57.5±1.5	2.2±0.3	2.8±1.4 ^b	6.1±2.5 ^c
	48	237.1±1.4	16.5±1.3	24.7±3.7	50.3±3.2
MRS + 1.5% Phe	0	_a	_a	_a	_a
	24	116.4±1.3 ^c	8.3±0.8	11.0±0.5	25.4±3.1 ^d
	48	361.2±2.4 ^d	32.7±0.2 ^d	63.5±2.1 ^d	84.1±3.2 ^e
MRS + 2.0% Phe	0	_a	_a	_a	_a
	24	113.2±1.8 ^c	14.1±2.5	16.0±1.1	27.1±2.1 ^d
	48	363.8±2.0 ^d	33.1±0.6 ^d	62.9±1.6 ^d	83.5±2.2 ^e

728 # Results shown are mean values ± confidence interval. Values in one column followed by the same lower case letter are not significantly different, values
729 without letter of significance are significantly different from all other values in the same column (p<0.05)

Table 2: Concentration of reuterin (mM) and antifungal activity (% growth inhibition) against *Penicillium expansum*, *Aspergillus niger* and *Fusarium culmorum* of cell-free supernatants of *Lactobacillus reuteri* R29 grown in various MRS formulations, containing 1.0 – 2.0% of glucose and 250 – 1000mM of glycerol, for 0, 24 and 48h (1-step fermentation process) and for the 2-step fermentation process (2h in water/glycerol).#

Medium fermented by <i>L. reuteri</i> R29	fermentation time [h]	reuterin [mM]	antifungal activity [% growth inhibition against 10 ⁷ <i>F. culmorum</i> spores/mL]		
			<i>P. expansum</i>	<i>A. niger</i>	<i>F. culmorum</i>
Control (MRS)	0	–a	–a	–a	–a
	24	–a	0.2±0.1	0.5±0.1 ^b	0.3±0.2 ^a
	48	–a	3.1±0.7 ^b	8.2±0.4 ^c	18.4±1.4 ^b
MRS + 250mM glycerol; 1.0% glucose	0	–a	–a	–a	–a
	24	12.5±0.1	18.3±1.7 ^d	22.5±0.2 ^d	39.5±2.7
	48	–a	2.1±0.4 ^b	8.9±0.6 ^c	8.4±1.2 ^c
MRS + 250mM glycerol; 1.5% glucose	0	–a	–a	–a	–a
	24	20.5±1.1 ^b	17.0±1.2 ^d	24.5±1.3 ^d	55.8±3.1 ^d
	48	0.4±0.1	3.5±0.4 ^b	3.5±0.6 ^e	8.7±2.1 ^c
MRS + 250mM glycerol; 2% glucose	0	–a	–a	–a	–a
	24	0.9±0.2	4.5±0.2 ^e	11.1±1.9 ^f	15.7±2.6 ^e
	48	–a	2.2±0.3 ^b	1.8±0.5	14.6±0.1 ^e
MRS + 500mM glycerol; 1.0% glucose	0	–a	–a	–a	–a
	24	18.6±0.3	35.6±1.6	21.8±0.1 ^d	58.7±2.3 ^d
	48	0.2±0.1 ^a	2.4±0.2 ^b	3.3±0.3 ^e	8.8±0.6 ^c
MRS + 500mM glycerol; 1.5% glucose	0	–a	–a	–a	–a
	24	27.5±0.6 ^c	55.0±2.1	64.9±1.8	69.1±0.4
	48	1.2±0.3 ^d	3.1±1.1 ^b	4.5±0.4 ^e	17.9±2.1 ^{be}
MRS + 500mM glycerol; 2% glucose	0	–a	–a	–a	–a
	24	1.3±0.2 ^d	8.4±0.5	7.6±0.9	19.1±1.5 ^e
	48	–a	2.8±0.7 ^b	2.2±0.5 ^g	16.3±0.4 ^e
MRS + 1000mM glycerol; 1.0% glucose	0	–a	–a	–a	–a
	24	21.5±1.4 ^b	14.7±1.0	13.4±0.2 ^f	57.1±0.5 ^d
	48	0.3±0.1 ^a	–a	0.2±0.1 ^b	–a
MRS + 1000mM glycerol; 1.5% glucose	0	–a	–a	–a	–a
	24	26.6±0.3 ^c	26.5±1.1	28.5±1.5 ^d	52.9±2.4 ^d
	48	0.5±0.4 ^a	1.0±0.1 ^c	1.4±0.2 ^h	18.6±1.9 ^{be}
MRS + 1000mM glycerol; 2% glucose	0	–a	–a	–a	–a
	24	1.4±0.1 ^d	3.6±0.7 ^e	2.6±0.1 ^g	18.1±1.6 ^{be}
	48	–a	0.8±0.2 ^c	1.2±0.2 ^h	17.5±1.2 ^{be}
Medium (2-step fermentation process)	fermentation time [h]	reuterin [mM]	antifungal activity [% growth inhibition against 10 ⁷ <i>F. culmorum</i> spores/mL]		
			<i>P. expansum</i>	<i>A. niger</i>	<i>F. culmorum</i>

distilled water + 400mM glycerol	0 (control)	^a	^a	^a	^a
	2 (sample)	93.1±2.0	100±0	100±0	100±0

735 # Results shown are mean values ± confidence interval. Values in one column followed by the same
736 lower case letter are not significantly different, values without letter of significance are significantly
737 different from all other values in the same column (p<0.05)

738

739 **Figure 1:** *In vitro* antifungal activity against *F. culmorum* in A) MRS + 1.5% Phe (48h
740 fermented), B) 2-step fermentation supernatant, C) impact of heat treatment on
741 reuterin content of the cell-free supernatants and D) MRS + 500mM glycerol; 1.5%
742 glucose (24h fermented).

743 **Figure 2:** Shelf life of wheat bread against environmental mould during a 13-day
744 storage period is indicated as the percentage of the total surface area of each slice,
745 where fungal growth occurred. Mould-free slices (A), <10% mouldy (B), 10-24%
746 mouldy (C), 25-49% mouldy (D) and >50% mouldy (E). Mean values are shown
747 (n=3); error bars indicate standard deviations.

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