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1 **Bacteriocins: Antibiotics in the Age of the Microbiome**

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15 Key words: Bacteriocin, Microbiome, Antibiotic, Probiotic, Antimicrobial Resistance

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23 Running title: Bacteriocins as therapeutic antimicrobials in the context of the
24 microbiome.

25 **Abstract**

26 Antibiotics have revolutionised the treatment of infectious disease and improved the
27 lives of billions of people worldwide over many decades. With the rise in
28 antimicrobial resistance (AMR) and corresponding lack of antibiotic development, we
29 find ourselves in dire need of alternative treatments. Bacteriocins are a class of
30 bacterially produced, ribosomally synthesised, antimicrobial peptides that may be
31 narrow or broad in their spectrums of activity. Animal models have demonstrated the
32 safety and efficacy of bacteriocins in treating a broad range of infections, however,
33 one of the principal drawbacks has been their relatively narrow spectra as compared
34 with small molecule antibiotics. In an era where we are beginning to appreciate the
35 role of the microbiota in human and animal health, the fact that bacteriocins cause
36 much less collateral damage to the host microbiome makes them a highly desirable
37 therapeutic. This review makes a case for the implementation of bacteriocins as
38 therapeutic antimicrobials, either alone or in combination with existing antibiotics to
39 alleviate the AMR crisis and to lessen the impact of antibiotics on the host
40 microbiome.

41 **Abbreviations used:** AAD, Antibiotic Associated Diarrhoea; AMR,
42 Antimicrobial Resistance; AOM, Acute Otitis Media; CDAD, *Clostridium difficile*
43 Associated Disorder; CDC, Centres for Disease Control and Prevention; EcN,
44 *Escherichia coli* Nissle; GI, Gastrointestinal tract; MIC, Minimum Inhibitory
45 Concentration ;VRE, Vancomycin Resistant *Enterococci*.

46 **Introduction**

47 Antimicrobial resistance (AMR) has been recognised as one of the major threats to
48 public health in the 21st century. In a report commissioned by the UK government in
49 2014, it was estimated that AMR could be responsible for 10 million deaths
50 worldwide by 2050, with a global financial cost of \$100 trillion (1). Meanwhile the
51 Centers for Disease Control and Prevention (CDC) estimates the annual cost of AMR
52 in the US to range from \$20 billion in direct healthcare costs to \$35 billion in
53 additional costs to society due to lost productivity (2). Apart from the human and
54 financial costs associated with AMR, there are also ethical considerations that need to
55 be addressed surrounding how we as a society respond and deal with the AMR crisis
56 (3). There are multiple reasons for the present AMR crisis, but significant factors
57 include the incorrect/indiscriminate administration and use of antibiotics and a dry
58 antibiotic development pipeline (4, 5). The CDC also recently estimated that in the
59 US approximately 50% of antibiotics are incorrectly prescribed. Moreover, the use of
60 antibiotics in agriculture has continued, despite undeniable evidence that this practice
61 adds to the antimicrobial resistance crisis. Resistance to a key “last-resort” antibiotic,
62 colistin, has been observed in the US, Europe and Asia (6-8). We have also seen the
63 rapid spread of resistance to another “last resort” class of antibiotics, the carbapenems
64 (9). With the emergence of these new resistant strains and the emergence of pan-
65 resistant bacteria, it is safe to say we have truly arrived in the much predicted post-
66 antibiotic era (10).

67

68 It is important that we acknowledge that broad-spectrum antibiotic therapy has
69 revolutionised the treatment of infectious diseases within the last century, but we must
70 also admit to unintended consequences of antibiotic use, such as potentially negative

71 effects on the host microbiome and their potential toxicity (5, 11). Although the field
72 of microbiome research is in its infancy relative to that of antibiotic therapy, evidence
73 strongly suggests that the composition of the microbiome can be an indicator of health
74 and is likely to be involved in many aspects of human health and disease (12). Strides
75 in DNA sequencing technology and bioinformatics have increased our understanding
76 of the role of the microbiome in a variety of disease states. Indeed the administration
77 of antibiotics in early life and the subsequent disruption of the microbiota may
78 contribute to risk of obesity in later life (13, 14). Furthermore, when subjected to
79 broad-spectrum antibiotic therapy, non-target commensal microbes may evolve and/or
80 acquire resistance mechanisms to evade the effects of the antibiotic, thereby
81 contributing to the antibiotic resistance crisis.

82

83 Bacteriocins represent a class of powerful antimicrobial peptides that may provide at
84 least part of a solution to the AMR crisis. We aim to demonstrate their efficacy in the
85 treatment of infectious disease and their reduced impact on the host microbiome by
86 comparison to broad-spectrum antibiotic therapy.

87

88 **Bacteriocins: potent antimicrobial peptides**

89 Many excellent reviews have been written about bacteriocins (11, 15, 16) , but in
90 brief they are a diverse group of peptides that may be classified into three distinct
91 groups; class I (modified), class II (unmodified or cyclic) and Class III (>10kDa
92 peptides). Apart from their potent antimicrobial activity (with minimum inhibitory
93 concentrations [MIC's] often in the nanomolar range) they have also been shown to
94 have antiviral (17), anticancer (18) and immunomodulatory properties (19).
95 Bacteriocins typically have a narrow spectrum of activity, but broad-spectrum

196 peptides are also present in this class of antimicrobials (e.g. nisin and lactacin 3147
197 inhibit a wide range of Gram-positive bacteria). As a result these peptides may be
198 suitable for treating infections of unknown aetiology, using broad-spectrum
199 bacteriocins, or may allow more precise targeting of known infectious agents using
200 highly active narrow spectrum bacteriocins. Bacteriocins are gene encoded, which
201 makes them amenable to genetic alterations to improve functional characteristics.
202 Furthermore, their toxicity is low and they may be administered as either purified
203 peptide or produced *in situ* by bacteriocin producing probiotic bacteria (11).
204 Bacteriocins are also known to interact with a variety of receptors, which are different
205 to those targeted by antibiotics, making cross-resistance less likely (20). Although a
206 more targeted approach may still ultimately lead to resistance development in the
207 infectious agent, it does reduce the likelihood of resistance development in
208 commensal populations outside of the target range of the bacteriocin. Resistance
209 mechanisms involving the class II receptors, the mannose phosphotransferase system
210 (Man-PTS), have been identified (21) along with a variety of resistance mechanisms
211 to the class I lantibiotics (22).

212

213 **The microbiota perspective**

214 The term “superorganism” or “holobiont” has commonly been applied to describe the
215 relationship that exists between humans and its commensal microbes and viruses (23).
216 Understanding the role of the microbiota in health and protecting its diversity during
217 the treatment of infectious disease is a key element of why bacteriocins may be
218 suitable as alternatives to antibiotics.

219

120 The two-peptide lactobionic bacteriocin, Thuricin CD, is a narrow spectrum
121 bacteriocin. Thuricin CD is highly active against one of the main causative agent of
122 antibiotic associated diarrhoea (AAD), *Clostridium difficile*, which is responsible for
123 20-30% of AAD cases (24). Briefly, AAD is caused by a disruption of the microbiota
124 (often referred to as dysbiosis) following broad spectrum antibiotic treatment, and
125 notably has a recurrence rate of 15-60% (25). Thuricin CD was shown to exhibit
126 comparable activity to both vancomycin and metronidazole (two antibiotics used for
127 the treatment of AAD which has progressed to *C. difficile* associated disease, CDAD).
128 Importantly, it showed almost no effect on microbial diversity when compared to both
129 metronidazole and vancomycin in a distal colon model (26). The modified R-Type
130 bacteriocin, Av-CD291.2, has also been shown to prophylactically prevent
131 colonization of *C. difficile* in a mouse model without perturbing the microbiota (27).
132 There are other broad spectrum bacteriocins which are attractive therapeutic agents by
133 virtue of their activity against *C. difficile*, but while the broad spectrum lantibiotic
134 lactacin 3147 is effective at killing *C. difficile*, it has a significant impact on the
135 resident microbiome populations such as *Bifidobacterium*, *Lactobacillus* and
136 *Enterococcus* species (28). It has also been shown that a commercially available
137 product containing the lantibiotic nisin, Nisaplin[®], can eliminate a *C. difficile*
138 infection when added at a concentration of 20X MIC in a simulated human colon
139 model. However a significant decrease in the total microbiota count was observed,
140 with Gram-positives being adversely affected (29).

141

142 Notably, in recent years the emergence of Vancomycin Resistant *Enterococci* (VRE)
143 has become a great concern and therefore raises the issues surrounding the efficacy of
144 treating CDAD with vancomycin if it presents a risk to the general population and the

145 spread of antibiotic resistance. In this light, the treatment of CDAD with bacteriocins
146 could be a valuable alternative to vancomycin. When VRE development has taken
147 place, it has been shown that mice colonised with VRE can be decolonized through
148 the use of an *Enterococcus* probiotic containing a conjugation defective plasmid
149 which produces a bacteriocin named Bac-21 (30).

150

151 A defensin-like bacteriocin, bactofencin A, displays *in vitro* activity against *L.*
152 *monocytogenes* and *S. aureus* (31, 32). Although one might expect this medium to
153 broad spectrum antimicrobial peptide to cause drastic changes in the host
154 microbiome, this was in fact not the case and it was observed that the bactofencin
155 peptide only subtly modulated an *ex vivo* host microbiome (distal colon model) when
156 introduced as a bacteriocin producing probiotic or purified peptide. While the purified
157 peptide resulted in higher levels of beneficial microbes such as *Bifidobacterium*, it
158 was also associated with lower levels of *Clostridium*, which has been linked to
159 obesity and gut pathogenesis. Interestingly, although bactofencin does not show
160 inhibitory activity *in vitro* against strains from the genera *Clostridium*, *Fusobacterium*
161 and *Bacteroides*, the reduction of these populations in the bactofencin treated faecal
162 samples indicates that the consequence of bactofencin altering the overall microbiota
163 structure impacts, directly or indirectly, on these normally insensitive populations
164 when in the gut environment (33).

165

166 It has also been shown, using bacteriocin producing probiotic strains and their
167 isogenic mutants, that the production of bacteriocins can aid the colonisation of a
168 murine host (34). Sequencing data revealed that although bacteriocin production by
169 the probiotics did not affect bacterial diversity at the phylum level, broad spectrum

170 bacteriocins (enterocins and garvicin ML) had a more significant impact on the
171 genus/family diversity of the host microbiome than narrow spectrum bacteriocins
172 (sakacin A, plantaricins and pediocin PA-1).

173

174 **Bacteriocins in animal models**

175 Bacteriocins have been shown to be effective in the treatment of variety of infectious
176 bacteria using two delivery methods, either as purified peptides (Table 1) or when
177 delivered *in situ* by probiotics.

178

179 It has been hypothesized that there are three mechanisms by which bacteriocins
180 mediate their producers probiotic properties (42); (i) **Competitive Inhibition:**
181 bacteriocins may support colonization of the host through competitive inhibition of
182 the autologous microbiota, (ii) **Pathogen Inhibition:** bacteriocins may interact
183 directly on a pathogenic target, or (iii) **Immunomodulation:** bacteriocins may act as
184 signalling peptides, recruiting other bacteria or recruiting immune cells to the site of
185 infection to aid elimination of the pathogen (**Figure 1**).

186

187 ***Preventing infection***

188

189 The concept of oral replacement therapy is another interesting example of
190 prophylactic probiotic therapy, which has been investigated using the mutacin 1140
191 producing *streptococcus mutans* BCS3-L1. This bacteriocin producing strain is
192 suitable for replacement therapy as it has reduced cariogenic potential because it does
193 not produce lactic acid, mediated through the removal of its entire lactic acid
194 dehydrogenase operon (43). Another interesting probiotic that has shown promise in

195 the limitation of dental caries, plaque accumulation and acidification is *Streptococcus*
196 *salivarius* M18. This strain has 3 plasmid and 1 chromosomally encoded bacteriocins,
197 which is perhaps why it can colonise the oral cavity so effectively. It also produces
198 two enzymes, urease and dextranase, which reduce saliva acidity and counteract
199 plaque formation (44). In a clinical trial, both the safety and efficacy of this strain's
200 probiotic potential was demonstrated, and it was shown to significantly reduce plaque
201 formation in subjects who received the probiotic, over those who received the placebo
202 (45). Furthermore, the treatment of children who have a high risk of dental caries
203 development, with an oral formulation of the *S. salivarius* M18 probiotic (Carioblis®)
204 was shown to reduce the likelihood of new dental caries development (46).

205

206 It has been demonstrated that dosing mice orally with the bacteriocin producer *Lb.*
207 *salivarius* UCC118 three days prior to infection with *L. monocytogenes* resulted in a
208 significant reduction in subsequent infection by *L. monocytogenes* (47). Nisin Z and
209 pediocin AcH have also been shown to reduce and prevent the colonisation of a
210 mouse model with vancomycin resistant *Enterococci* (VRE), where the
211 bacteriocinogenic probiotic was administered 8 days prior to infection (48). It has also
212 been demonstrated using a porcine model that *Salmonella enterica* serovar
213 *typhimurium* shedding is reduced and disease symptoms of infection are alleviated
214 when a mix of five probiotic strains was administered 6 days before infection (49).
215 One of the probiotics, *L. salivarius*, produces salivaricin P, which can kill the other 4
216 strains in the probiotic mix. Interestingly, this bacteriocinogenic strain dominated in
217 the ileum (the primary attachment site of the infecting *Salmonella*) whereas it was
218 only detected as a minor component in the faeces of the same animals. This suggests
219 that bacteriocin production may play a role where colonisation can occur along the

gastrointestinal (GI) tract (50). The concept of using prophylactic probiotics to competitively colonise a pathogens niche could be an effective strategy in agriculture to reduce antibiotic usage. If, as expected, regulations limiting the use of antibiotics in agriculture come into force, probiotics may be invaluable alternative.

Acute otitis media (AOM) is a type of inflammatory disease of the middle ear, characterised typically by rapid inflammation, potential tympanic membrane perforation, along with fullness and erythema. It has been reported that the levels of normal α -haemolytic *Streptococcus* colonising the nasopharynx of otitis prone children is much lower than in healthy individuals and that recolonization can significantly reduce the episodes of AOM (51, 52). It has been demonstrated treating otitis prone children with a history of AOM, with a nasal spray containing safe *Streptococcus salivarius* 24SMB (a strain which produces a bacteriocin-like substance), reduces the incidences of AOM over the placebo treated group (53).

Treating infection

Helicobacter pylori infection and colonisation results in a variety of disease states and may even lead to the development of gastric carcinoma. More recently, the prevalence of antibiotic resistant *H. pylori* has been increasing, creating a need for a new therapeutic agent (54). It has been shown in mice that eradication of *H. pylori* was achieved using a bacteriocinogenic probiotic treatment of *P. acidilactici* BA28 (55). Using a mixture of cranberry juice and the bacteriocin producing probiotic culture *Lactobacillus johnsonii* str. La1 supernatant, the carriage of *H. pylori* was also reduced in children after three weeks of treatment (56).

245

246 One barrier to the use of probiotics as a therapeutic is their ability to survive and
247 colonise the area of infection. It has been shown that *Pediococcus acidilactici* UL5
248 and *Lactococcus lactis* ATCC 11454 can produce the bacteriocins pediocin PA-1 and
249 nisin, respectively, *in situ* under simulated upper gastric conditions (57). Interestingly,
250 the *in vitro* activity of a bacteriocin does not always correspond to the *in vivo* activity,
251 where the bacteriocin is sometimes more or less active in an animal model, as is the
252 case with mersacidin which is more active *in vivo* than *in vitro* (58).

253

254 **Bacteriocins against Gram-negatives**

255 Comparatively speaking, Gram-negative bacteria are relatively insensitive to
256 bacteriocins compared to their Gram-positive counterparts, largely owing to the outer
257 membrane which acts as a physical barrier. Until recently the treatment of Gram-
258 negative infections with bacteriocins has not been favoured due to the efficacy of
259 conventional antibiotics in the treatment of these infections. The rise of antibiotic
260 resistant Gram-negative bacteria to the last line of antibiotics (6) means the treatment
261 of Gram-negative infections using bacteriocins can no longer be ignored .

262

263 Widespread use requires a solution to the relative insensitivity of Gram-negative
264 microorganisms. One possibility is to use bacteriocins in combination with other
265 antimicrobial agents, including conventional antibiotics. Although conventional
266 antibiotics will have an impact on the host microbiota (as previously discussed),
267 certain bacteriocin/antibiotic combinations can be synergistic (59-62) and therefore
268 lead to a reduced dose of both antimicrobial agents needed to treat an infection,

269 thereby lowering the potential effect on the host microbiome, the cytotoxic effects on
270 the host and may potentially reduce the development of resistance.

271

272 Success of antibiotics is also hindered by Gram-negative bacteria residing within
273 biofilms, where they are highly resistant to antibiotic treatments. Bacteriocin/
274 antibiotic combinations have shown great promise in overcoming biofilm mediated
275 resistance for important Gram-negative pathogens such as *Pseudomonas aeruginosa*
276 (63) and *Escherichia coli* (64).

277

278 Although this review mainly focuses on Gram-positive bacteriocins, it is important to
279 also to identify Gram-negative bacteriocins which may have potential therapeutic
280 significance. Microcins are ribosomally-synthesized peptides commonly produced by
281 Gram-negative bacteria which are active against Gram-negative strains, and are an
282 interesting alternative to Gram-positive bacteriocins. They have been shown to
283 display potent antimicrobial activity *in vitro* (65, 66) and more recently also *in vivo*
284 (67). It has been demonstrated that the microcin producer *E. coli* Nissle 1917 (EcN)
285 can prevent colonisation of competing *Enterobacteriaceae* in the gut, whilst still
286 having a minimal impact on the diversity of the gut microbiota. However, EcN
287 microcins exhibit their mechanism of action by targeting specific siderophore
288 receptors on other *Enterobacteriaceae* which are only displayed during iron
289 starvation, making their spectrum of activity quite narrow. Additionally to its
290 prophylactic applications, EcN has also been demonstrated to reduce inflammation
291 and weight loss associated with *Salmonella* infections. Another microcin produced by
292 *E. coli* G3/10, microcin S, has been shown to inhibit other *E. coli* strains and

293 furthermore can prevent the adherence of Enteropathogenic *E. coli* (EPEC) to
294 intestinal epithelial cells (68).

295

296 **Overcoming the limitations/outlook**

297 In previous decades significant emphasis was placed on functional characteristics of
298 bacteriocins, such as spectrum of activity, pH and temperature stability, which were
299 essential for the use of bacteriocins in food applications. For their use as therapeutics
300 additional characteristics such as proteolytic resistance, stability and solubility of
301 bacteriocins will also be important.

302

303 With advancements in the field of bioengineering many intrinsic limitations have
304 been overcome, and it has been shown using the prototypic lantibiotic, nisin, that
305 bioengineering strategies can improve functional qualities such as antimicrobial
306 activity (69-72), solubility (73, 74) diffusion properties (75) and effectiveness against
307 Gram-negative bacteria (71). Indeed, similar bioengineering strategies could be
308 applied to other bacteriocins once suitable expression systems have been developed.
309 Although the sensitivity of bacteriocins to proteolytic cleavage was previously
310 regarded as a desirable trait when using these peptides as food preservatives, it does
311 represent a major concern with regard to their administration and widespread use,
312 both orally and intravenously. Bioengineering strategies could be once again used to
313 manipulate peptide residues so they are no longer recognisable by host proteases and
314 therefore are not proteolytically cleaved, thereby improving peptide functional
315 qualities (76). Notably, the therapeutic application of the prototypic bacteriocin, nisin,
316 has been in part hampered by its sensitivity to host proteases (77). Other approaches
317 include prospecting for bacteriocins that display innate resistance to proteases, as was

318 achieved with pseudomycoicidin (78), which is naturally resistant to trypsin due to the
319 presence of a thioether ring structure. The field of bioinformatics and the use of such
320 programs as BAGEL 3.0 (79) and antiSMASH (80) could be a fundamental aspect of
321 this prospecting, as these bacteriocin amino acid prediction tools from genome
322 sequences may also allow researchers to identify protease resistant peptides before
323 investing large amounts of time and effort in characterising such bacteriocins. Finally,
324 understanding bacteriocin pharmacodynamics and pharmacokinetics are also essential
325 to their safe implementation as therapeutics, which has been under investigated in
326 comparison to other aspects of bacteriocin research. If bacteriocins are indeed to
327 become an alternative to conventional antibiotics, a greater emphasis must be placed
328 on research surrounding these host-drug interactions, such as was achieved with
329 MU1140 (81). Addressing these limitations of bacteriocin research to date could
330 provide a turning point for the flagging interest of the pharmaceutical industry and
331 make bacteriocins an attractive therapeutic alternative to current antibiotics (10).

332

333 Although there is considerable evidence that narrow spectrum bacteriocins have a
334 minimal effect on the host microbiome by comparison to current broad-spectrum
335 antibiotics, it should also be recognised that more work in this regard is needed to
336 strengthen the argument for the use of bacteriocins as antibiotics, along with
337 overcoming the previously outlined limitations. Ultimately, we believe given the safe
338 history of use of bacteriocins in food and the large body of literature surrounding this
339 field, that they are useful candidates for antimicrobial therapeutics as the AMR crisis
340 continues to worsen.

341 **Summary**

- 342 • Antimicrobial resistance (AMR) is a major threat to public health requiring
343 immediate attention.
- 344 • Bacteriocins are potent antimicrobial peptides, active in the nanomolar range and
345 have a reduced impact on the host microbiota.
- 346 • Bacteriocins may be used to treat a broad range of infections and can be delivered
347 as purified peptides or as bacteriocinogenic probiotics.
- 348 • Combining antibiotics and bacteriocins is a strategy to reduce the negative
349 impacts on the host microbiota and also alleviate the AMR crisis.
- 350 • Overcoming the current limitations of bacteriocin-based therapeutics should be a
351 key goal of bacteriocin research in the future.

352 **Declarations of interest**

353 The authors declare that the research was conducted in the absence of any commercial
354 or financial relationships that could be construed as a potential conflict of interest.

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361 **Author Contribution statement**

362 KE drafted the manuscript. RR and CH revised and approved the final manuscript.

1. O'Neill J. Antimicrobial Resistance: tackling a crisis for the health and wealth of nations. The Review on Antimicrobial Resistance. 2014.
2. Centers for Disease Control and prevention. Antibiotic Resistance Threats in the United States 2013. Washington: CDC. 2013.
3. Littmann J, Viens AM. The Ethical Significance of Antimicrobial Resistance. Public Health Ethics. 2015;8(3):209-24.
4. Arias CA, Murray BE. Antibiotic-Resistant Bugs in the 21st Century — A Clinical Super-Challenge. New England Journal of Medicine. 2009;360(5):439-43.
5. Riley MA, Robinson SM, Roy CM, Dorit RL. Rethinking the composition of a rational antibiotic arsenal for the 21st century. Future medicinal chemistry. 2013;5(11):1231-42.
6. Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. The Lancet Infectious diseases. 2016;16(2):161-8.
7. Ye H, Li Y, Li Z, Gao R, Zhang H, Wen R, et al. Diversified mcr-1-Harboured Plasmid Reservoirs Confer Resistance to Colistin in Human Gut Microbiota. mBio. 2016;7(2).

- 388 8. Meinersmann RJ, Ladely SR, Plumblee JR, Cook KL, Thacker E. Prevalence
389 of mcr-1 in US Food-Animal Cecal Contents. *Antimicrobial agents and*
390 *chemotherapy*. 2016.
391
- 392 9. Potter RF, D'Souza AW, Dantas G. The rapid spread of carbapenem-resistant
393 *Enterobacteriaceae*. *Drug Resistance Updates*. 2016;29:30-46.
394
- 395 10. Fair RJ, Tor Y. Antibiotics and Bacterial Resistance in the 21st Century.
396 *Perspectives in Medicinal Chemistry*. 2014;6:25-64.
397
- 398 11. Cotter PD, Ross RP, Hill C. Bacteriocins - a viable alternative to antibiotics?
399 *Nature reviews Microbiology*. 2013;11(2):95-105.
400
- 401 12. Guinane CM, Cotter PD. Role of the gut microbiota in health and chronic
402 gastrointestinal disease: understanding a hidden metabolic organ. *Therapeutic*
403 *Advances in Gastroenterology*. 2013;6(4):295-308.
404
- 405 13. Cox LM, Blaser MJ. Antibiotics in early life and obesity. *Nat Rev Endocrinol*.
406 2015;11(3):182-90.
407
- 408 14. Petschow B, Dore J, Hibberd P, Dinan T, Reid G, Blaser M, et al. Probiotics,
409 prebiotics, and the host microbiome: the science of translation. *Annals of the New*
410 *York Academy of Sciences*. 2013;1306:1-17.

- 411 15. Rea M, Ross RP, Cotter P, Hill C. Classification of Bacteriocins from Gram-
412 Positive Bacteria. In: Drider D, Rebuffat S, editors. Prokaryotic Antimicrobial
413 Peptides: Springer New York; 2011. p. 29-53.
414
- 415 16. Alvarez-Sieiro P, Montalbán-López M, Mu D, Kuipers OP. Bacteriocins of
416 lactic acid bacteria: extending the family. *Applied Microbiology and Biotechnology*.
417 2016;100(7):2939-51.
- 418 17. Al Kassaa I, Hober D, Hamze M, Chihib NE, Drider D. Antiviral potential of
419 lactic acid bacteria and their bacteriocins. *Probiotics and Antimicrobial Proteins*.
420 2014;6(3-4):177-85.
421
- 422 18. Kamarajan P, Hayami T, Matte B, Liu Y, Danciu T, Ramamoorthy A, et al.
423 Nisin ZP, a Bacteriocin and Food Preservative, Inhibits Head and Neck Cancer
424 Tumorigenesis and Prolongs Survival. *PloS one*. 2015;10(7):e0131008.
425
- 426 19. de Pablo MA, Gaforio JJ, Gallego AM, Ortega E, Galvez AM, Alvarez de
427 Cienfuegos Lopez G. Evaluation of immunomodulatory effects of nisin-containing
428 diets on mice. *FEMS immunology and medical microbiology*. 1999;24(1):35-42.
429
- 430 20. Cotter PD. An 'Upp'-turn in bacteriocin receptor identification. *Molecular*
431 *microbiology*. 2014;92(6):1159-63.
432
- 433 21. Kjos M, Nes IF, Diep DB. Mechanisms of Resistance to Bacteriocins
434 Targeting the Mannose Phosphotransferase System. *Applied and Environmental*
435 *Microbiology*. 2011;77(10):3335-42.

436

437 22. Draper LA, Cotter PD, Hill C, Ross RP. Lantibiotic resistance. Microbiology
438 and molecular biology reviews : MMBR. 2015;79(2):171-91.

439

440 23. Goodacre R. Metabolomics of a superorganism. The Journal of nutrition.
441 2007;137(1 Suppl):259s-66s.

442

443 24. Rea MC, Alemayehu D, Ross RP, Hill C. Gut solutions to a gut problem:
444 bacteriocins, probiotics and bacteriophage for control of Clostridium difficile
445 infection. Journal of medical microbiology. 2013;62(9):1369-78.

446

447 25. McFarland LV. Antibiotic-associated diarrhea: epidemiology, trends and
448 treatment. Future microbiology. 2008;3(5):563-78.

449

450 26. Rea MC, Dobson A, O'Sullivan O, Crispie F, Fouhy F, Cotter PD, et al. Effect
451 of broad- and narrow-spectrum antimicrobials on Clostridium difficile and microbial
452 diversity in a model of the distal colon. Proceedings of the National Academy of
453 Sciences of the United States of America. 2011;108 Suppl 1:4639-44.

454

455 27. Gebhart D, Lok S, Clare S, Tomas M, Stares M, Scholl D, et al. A modified R-
456 type bacteriocin specifically targeting Clostridium difficile prevents colonization of
457 mice without affecting gut microbiota diversity. mBio. 2015;6(2).

458

- 459 28. Rea MC, Clayton E, Connor PM, Shanahan F, Kiely B, et al.
460 Antimicrobial activity of lacticin 3147 against clinical *Clostridium difficile* strains.
461 *Journal of medical microbiology*. 2007;56(7):940-6.
462
- 463 29. Le Lay C, Fernandez B, Hammami R, Ouellette M, Fliss I. On *Lactococcus*
464 *lactis* UL719 competitiveness and nisin (Nisaplin®) capacity to inhibit *Clostridium*
465 *difficile* in a model of human colon. *Frontiers in Microbiology*. 2015;6:1020.
466
- 467 30. Kommineni S, Bretl DJ, Lam V, Chakraborty R, Hayward M, Simpson P, et
468 al. Bacteriocin production augments niche competition by enterococci in the
469 mammalian gastrointestinal tract. *Nature*. 2015;526(7575):719-22.
470
- 471 31. O'Shea EF, O'Connor PM, O'Sullivan O, Cotter PD, Ross RP, Hill C.
472 Bactofencin A, a New Type of Cationic Bacteriocin with Unusual Immunity. *mBio*.
473 2013;4(6).
474
- 475 32. O'Shea EF, Gardiner GE, O'Connor PM, Mills S, Ross RP, Hill C.
476 Characterization of enterocin- and salivaricin-producing lactic acid bacteria from the
477 mammalian gastrointestinal tract. *FEMS microbiology letters*. 2009;291(1):24-34.
478
- 479 33. Guinane CM, Lawton EM, O'Connor PM, O'Sullivan O, Hill C, Ross RP, et
480 al. The bacteriocin bactofencin A subtly modulates gut microbial populations.
481 *Anaerobe*. 2016;40:41-9.
482

- 483 34. Umu ÖCO, Bäuerl C, Oostindjer M, Pope PB, Hernández PE, Pérez-Martínez
484 G, et al. The Potential of Class II Bacteriocins to Modify Gut Microbiota to Improve
485 Host Health. PloS one. 2016;11(10):e0164036.
- 486
- 487 35. De Kwaadsteniet M, Doeschate KT, Dicks LM. Nisin F in the treatment of
488 respiratory tract infections caused by Staphylococcus aureus. Letters in Applied
489 Microbiology. 2009;48(1):65-70.
- 490
- 491 36. van Staden AD, Brand AM, Dicks LM. Nisin F-loaded brushite bone cement
492 prevented the growth of Staphylococcus aureus in vivo. Journal of applied
493 microbiology. 2012;112(4):831-40.
- 494
- 495 37. Kim SY, Shin S, Koo HC, Youn JH, Paik HD, Park YH. In vitro antimicrobial
496 effect and in vivo preventive and therapeutic effects of partially purified lantibiotic
497 lactacin NK34 against infection by Staphylococcus species isolated from bovine
498 mastitis. Journal of dairy science. 2010;93(8):3610-5.
- 499
- 500 38. Champion A, Casey PG, Field D, Cotter PD, Hill C, Ross RP. In vivo activity
501 of Nisin A and Nisin V against Listeria monocytogenes in mice. BMC Microbiology.
502 2013;13(1):23.
- 503
- 504 39. Rihakova J, Cappellet J-M, Hue I, Demnerova K, Fédérighi M, Prévost H, et
505 al. In Vivo Activities of Recombinant Divercin V41 and Its Structural Variants
506 against Listeria monocytogenes. Antimicrobial agents and chemotherapy.
507 2010;54(1):563-4.

508

509 40. Mota-Meira M, Morency H, Lavoie MC. In vivo activity of mutacin B-Ny266.
510 Journal of Antimicrobial Chemotherapy. 2005;56(5):869-71.

511

512 41. Kruszezwska D, Sahl H-G, Bierbaum G, Pag U, Hynes SO, Ljungh Å.
513 Mersacidin eradicates methicillin-resistant *Staphylococcus aureus* (MRSA) in a
514 mouse rhinitis model. Journal of Antimicrobial Chemotherapy. 2004;54(3):648-53.

515

516 42. Dobson A, Cotter PD, Ross RP, Hill C. Bacteriocin production: a probiotic
517 trait? Applied and Environmental Microbiology. 2012;78(1):1-6.

518

519 43. Hillman JD, Mo J, McDonell E, Cvitkovitch D, Hillman CH. Modification of
520 an effector strain for replacement therapy of dental caries to enable clinical safety
521 trials. Journal of applied microbiology. 2007;102(5):1209-19.

522

523 44. Burton JP, Wescombe PA, Macklaim JM, Chai MHC, MacDonald K, Hale
524 JDF, et al. Persistence of the Oral Probiotic *Streptococcus salivarius* M18 Is Dose
525 Dependent and Megaplasmid Transfer Can Augment Their Bacteriocin Production
526 and Adhesion Characteristics. PloS one. 2013;8(6):e65991.

527

528 45. Burton JP, Drummond BK, Chilcott CN, Tagg JR, Thomson WM, Hale JD, et
529 al. Influence of the probiotic *Streptococcus salivarius* strain M18 on indices of dental
530 health in children: a randomized double-blind, placebo-controlled trial. Journal of
531 medical microbiology. 2013;62(Pt 6):875-84.

532

533 46. Di Pierro F, Zanvit A, Nobili P, Risso P, Fornaini C. Cariogram outcome after
534 90 days of oral treatment with *Streptococcus salivarius* M18 in children at high risk
535 for dental caries: results of a randomized, controlled study. Clinical, cosmetic and
536 investigational dentistry. 2015;7:107-13.
537

538 47. Corr SC, Li Y, Riedel CU, O'Toole PW, Hill C, Gahan CGM. Bacteriocin
539 production as a mechanism for the antiinfective activity of *Lactobacillus salivarius*
540 UCC118. Proceedings of the National Academy of Sciences. 2007;104(18):7617-21.
541

542 48. Millette M, Cornut G, Dupont C, Shareck F, Archambault D, Lacroix M.
543 Capacity of human nisin- and pediocin-producing lactic Acid bacteria to reduce
544 intestinal colonization by vancomycin-resistant enterococci. Applied and
545 Environmental Microbiology. 2008;74(7):1997-2003.
546

547 49. Casey PG, Gardiner GE, Casey G, Bradshaw B, Lawlor PG, Lynch PB, et al.
548 A Five-Strain Probiotic Combination Reduces Pathogen Shedding and Alleviates
549 Disease Signs in Pigs Challenged with *Salmonella enterica* Serovar Typhimurium.
550 Applied and Environmental Microbiology. 2007;73(6):1858-63.
551

552 50. Walsh MC, Gardiner GE, Hart OM, Lawlor PG, Daly M, Lynch B, et al.
553 Predominance of a bacteriocin-producing *Lactobacillus*
554 *salivarius* component of a five-strain probiotic in the porcine ileum and
555 effects on host immune phenotype. FEMS Microbiology Ecology. 2008;64(2):317.
556

- 557 51. Marchisio P, Claut L, Rognoni A, Esposito S, Passali D, Bellussi L, et al.
558 Differences in nasopharyngeal bacterial flora in children with nonsevere recurrent
559 acute otitis media and chronic otitis media with effusion: implications for
560 management. *The Pediatric infectious disease journal*. 2003;22(3):262-8.
561
- 562 52. Roos K, Hakansson EG, Holm S. Effect of recolonisation with "interfering"
563 alpha streptococci on recurrences of acute and secretory otitis media in children:
564 randomised placebo controlled trial. *BMJ (Clinical research ed)*. 2001;322(7280):210-
565 2.
566
- 567 53. Santagati M, Scillato M, Patanè F, Aiello C, Stefani S. Bacteriocin-producing
568 oral streptococci and inhibition of respiratory pathogens. *FEMS Immunology and*
569 *Medical Microbiology*. 2012;65(1):23-31.
570
- 571 54. Thung I, Aramin H, Vavinskaya V, Gupta S, Park JY, Crowe SE, et al.
572 Review article: the global emergence of *Helicobacter pylori* antibiotic resistance.
573 *Alimentary pharmacology & therapeutics*. 2016;43(4):514-33.
574
- 575 55. Kaur B, Garg N, Sachdev A, Kumar B. Effect of the Oral Intake of Probiotic
576 *Pediococcus acidilactici* BA28 on *Helicobacter pylori* Causing Peptic Ulcer in
577 C57BL/6 Mice Models. *Applied biochemistry and biotechnology*. 2014;172(2):973-
578 83.
579

- 580 56. Gotteland M, Andrews M, Toledo M, Muñoz L, Caceres P, Anziani A, et al.
581 Modulation of *Helicobacter pylori* colonization with cranberry juice and *Lactobacillus*
582 *johnsonii* La1 in children. *Nutrition*. 2008;24(5):421-6.
583
- 584 57. Fernandez B, Hammami R, Savard P, Jean J, Fliss I. *Pediococcus acidilactici*
585 UL5 and *Lactococcus lactis* ATCC 11454 are able to survive and express their
586 bacteriocin genes under simulated gastrointestinal conditions. *Journal of applied*
587 *microbiology*. 2014;116(3):677-88.
588
- 589 58. Chatterjee S, Chatterjee DK, Jani RH, Blumbach J, Ganguli BN, Klesel N, et
590 al. Mersacidin, a new antibiotic from *Bacillus*. In vitro and in vivo antibacterial
591 activity. *The Journal of antibiotics*. 1992;45(6):839-45.
592
- 593 59. Naghmouchi K, Le Lay C, Baah J, Drider D. Antibiotic and antimicrobial
594 peptide combinations: synergistic inhibition of *Pseudomonas fluorescens* and
595 antibiotic-resistant variants. *Resistance in Microbiology*. 2012;163(2):101-8.
596
- 597 60. Naghmouchi K, Baah J, Hober D, Jouy E, Rubrecht C, Sane F, et al.
598 Synergistic effect between colistin and bacteriocins in controlling Gram-negative
599 pathogens and their potential to reduce antibiotic toxicity in mammalian epithelial
600 cells. *Antimicrobial agents and chemotherapy*. 2013;57(6):2719-25.
601
- 602 61. Rishi P, Preet Singh A, Garg N, Rishi M. Evaluation of nisin-beta-lactam
603 antibiotics against clinical strains of *Salmonella enterica* serovar Typhi. *The Journal*
604 *of antibiotics*. 2014;67(12):807-11.

605

606 62. Draper LA, Cotter PD, Hill C, Ross RP. The two peptide lantibiotic lacticin
607 3147 acts synergistically with polymyxin to inhibit Gram negative bacteria. BMC
608 Microbiology. 2013;13:212.

609

610 63. Field D, Seisling N, Cotter PD, Ross RP, Hill C. Synergistic Nisin-Polymyxin
611 Combinations for the Control of Pseudomonas Biofilm Formation. Frontiers in
612 Microbiology. 2016;7(1713).

613

614 64. Al Atya AK, Abriouel H, Kempf I, Jouy E, Auclair E, Vachee A, et al. Effects
615 of Colistin and Bacteriocins Combinations on the In Vitro Growth of Escherichia coli
616 Strains from Swine Origin. Probiotics and Antimicrobial Proteins. 2016.

617

618 65. Patzer SI, Baquero MR, Bravo D, Moreno F, Hantke K. The colicin G, H and
619 X determinants encode microcins M and H47, which might utilize the catecholate
620 siderophore receptors FepA, Cir, Fiu and IroN. Microbiology (Reading, England).
621 2003;149(9):2557-70.

622

623 66. Nolan EM, Walsh CT. Investigations of the MceIJ-catalyzed posttranslational
624 modification of the microcin E492 C-terminus: linkage of ribosomal and
625 nonribosomal peptides to form "trojan horse" antibiotics. Biochemistry.
626 2008;47(35):9289-99.

627

628 67. Sassone-Corsi M, Nuccio S-P, Liu H, Hernandez D, Vu CT, Takahashi AA, et
629 al. Microcins mediate competition among Enterobacteriaceae in the inflamed gut.
630 Nature. 2016;540(7632):280-3.
631

632 68. Zschüttig A, Zimmermann K, Blom J, Goesmann A, Pöhlmann C, Gunzer F.
633 Identification and Characterization of Microcin S, a New Antibacterial Peptide
634 Produced by Probiotic Escherichia coli G3/10. PloS one. 2012;7(3):e33351.
635

636 69. Healy B, Field D, O'Connor PM, Hill C, Cotter PD, Ross RP. Intensive
637 mutagenesis of the nisin hinge leads to the rational design of enhanced derivatives.
638 PloS one. 2013;8(11):e79563.
639

640 70. Molloy EM, Field D, PM OC, Cotter PD, Hill C, Ross RP. Saturation
641 mutagenesis of lysine 12 leads to the identification of derivatives of nisin A with
642 enhanced antimicrobial activity. PloS one. 2013;8(3):e58530.
643

644 71. Field D, Begley M, O'Connor PM, Daly KM, Hugenholtz F, Cotter PD, et al.
645 Bioengineered nisin A derivatives with enhanced activity against both Gram positive
646 and Gram negative pathogens. PloS one. 2012;7(10):e46884.
647

648 72. Field D, Connor PM, Cotter PD, Hill C, Ross RP. The generation of nisin
649 variants with enhanced activity against specific gram-positive pathogens. Molecular
650 microbiology. 2008;69(1):218-30.
651

- 652 73. Rollema HS, Kuipers OP, Both P, de Vos WM, Siezen RJ. Improvement of
653 solubility and stability of the antimicrobial peptide nisin by protein engineering.
654 *Applied and Environmental Microbiology*. 1995;61(8):2873-8.
655
- 656 74. Yuan J, Zhang ZZ, Chen XZ, Yang W, Huan LD. Site-directed mutagenesis of
657 the hinge region of nisinZ and properties of nisinZ mutants. *Applied Microbiology*
658 *and Biotechnology*. 2004;64(6):806-15.
659
- 660 75. Rouse S, Field D, Daly KM, O'Connor PM, Cotter PD, Hill C, et al.
661 Bioengineered nisin derivatives with enhanced activity in complex matrices.
662 *Microbial biotechnology*. 2012;5(4):501-8.
663
- 664 76. Field D, Cotter PD, Hill C, Ross RP. Bioengineering Lantibiotics for
665 Therapeutic Success. *Frontiers in Microbiology*. 2015;6(1363).
666
- 667 77. Field D, Cotter PD, Ross RP, Hill C. Bioengineering of the model lantibiotic
668 nisin. *Bioengineered*. 2015;6(4):187-92.
669
- 670 78. Basi-Chipalu S, Dischinger J, Josten M, Szekat C, Zweynert A, Sahl H-G, et
671 al. Pseudomycoicidin, a Class II Lantibiotic from *Bacillus pseudomycoides*. *Applied*
672 *and Environmental Microbiology*. 2015;81(10):3419-29.
673
- 674 79. van Heel AJ, de Jong A, Montalban-Lopez M, Kok J, Kuipers OP. BAGEL3:
675 Automated identification of genes encoding bacteriocins and (non-)bactericidal

676 posttranslationally modified peptides. Nucleic acids research. 2013;41(Web Server
677 issue):W448-53.

678

679 80. Blin K, Medema MH, Kottmann R, Lee SY, Weber T. The antiSMASH
680 database, a comprehensive database of microbial secondary metabolite biosynthetic
681 gene clusters. Nucleic acids research. 2016.

682

683 81. Ghobrial O, Derendorf H, Hillman JD. Pharmacokinetic and
684 pharmacodynamic evaluation of the lantibiotic MU1140. Journal of pharmaceutical
685 sciences. 2010;99(5):2521-8.

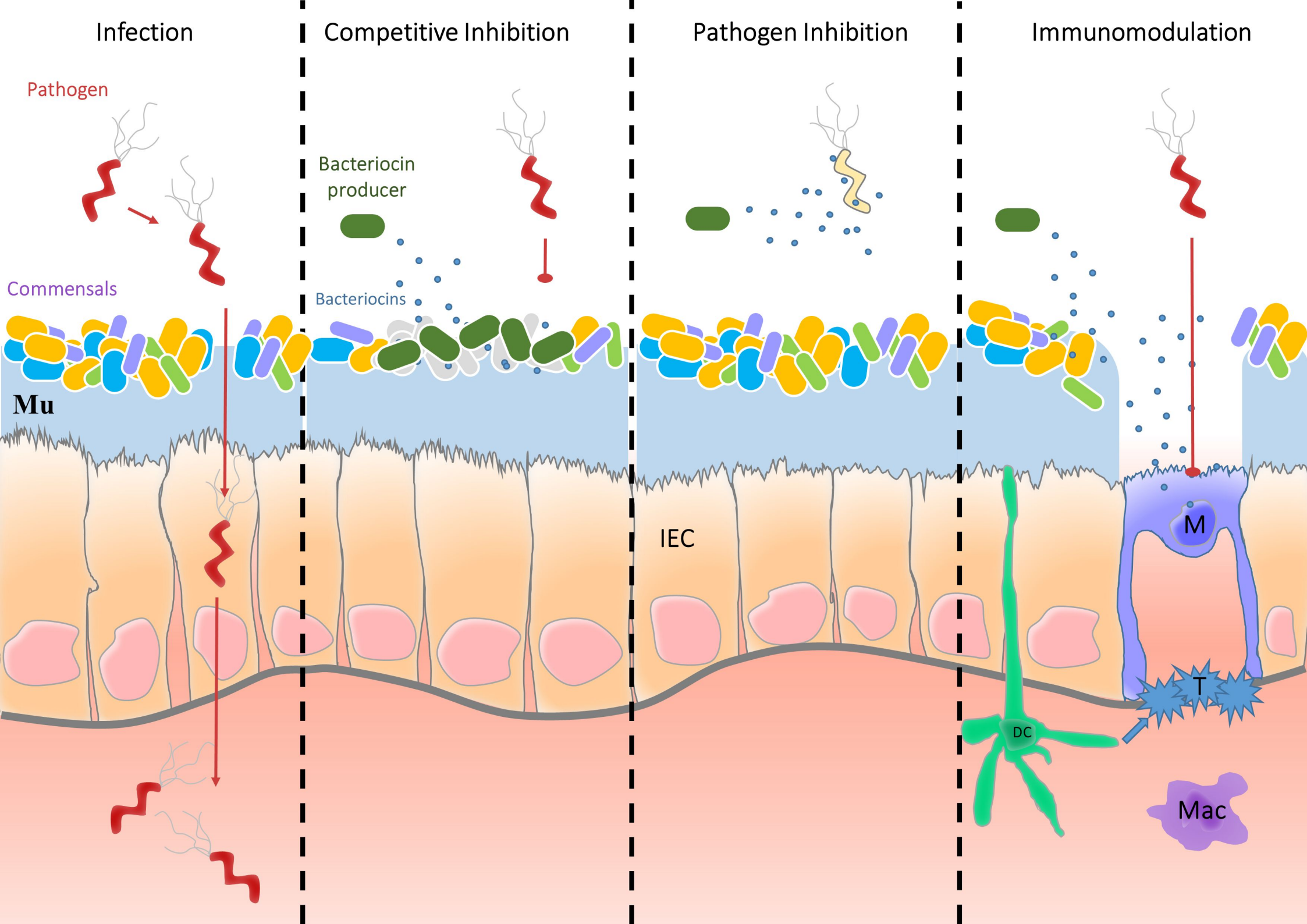
686 **Figure 1: Bacteriocinogenic probiotics can be utilized either prophylactically or**
687 **therapeutically to treat an infection.** M, M cell; Mac, macrophage; Mu, mucous; T,
688 T cell; IEC, intestinal epithelial cell; DC, dendritic cell.

Infection

Pathogen

Commensals

Mu



Peptide	Strain inhibited	Model	Purity	Reference
Nisin F	<i>S. aureus</i>	Immunosuppressed Wistar rats	Semi-pure	(35)
	<i>S. aureus</i>	Brushite cement in BALB/c mice	Semi-pure	(36)
Lacticin NK34	<i>S. aureus</i> / <i>S. simulans</i>	ICR mice	Semi-pure	(37)
Nisin V	<i>L. monocytogenes</i>	BALB/c mice	Pure	(38)
Divercin V41	<i>L. monocytogenes</i>	BALB/c mice	Pure	(39)
Mutacin B-Ny266	<i>S. aureus</i>	Unknown	Pure	(40)
Mersacidin	Methicillin- resistant <i>Staphylococcus</i> <i>aureus</i> (MRSA)	BALB/cA mice	pure	(41)

Table 1. Bacterial infections in animal models successfully treated using purified bacteriocins.