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1 **Application of percolation threshold to disintegration and dissolution of ibuprofen tablets**
2 **with different microcrystalline cellulose grades**

3

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ABSTRACT

This study investigated whether a step change in disintegration and dissolution behaviour was observed for tablets prepared below and above a percolation threshold. The percolation threshold investigated was previously calculated for ibuprofen/ microcrystalline cellulose (MCC) blends using percolation theory and compression data (Queiroz et al., 2019). The influence of MCC grade (air stream dried versus spray dried) on differences observed was also studied. Complementary to conventional disintegration and dissolution testing, Raman imaging determined drug distribution within tablets, and in-line particle video microscopy (PVM) and focused-beam reflectance measurement (FBRM) monitored tablet disintegration. Tablets were prepared containing 0 to 30% w/w ibuprofen. Raman imaging confirmed the percolation threshold by quantifying the number and equivalent circular diameters of ibuprofen domains on tablet surfaces. Across the percolation threshold a step change in dissolution behaviour occurred and tablets containing air stream dried MCC showed slower disintegration and dissolution compared to tablets containing spray dried MCC. Dissolution measurements confirmed experimentally a percolation threshold, determined using percolation theory and compression data. An increase in drug domains, due to cluster formation, and less efficient tablet disintegration contributed to slower ibuprofen release above the percolation threshold. Slower dissolution was measured for tablets containing air stream dried compared to spray dried MCC.

38 **1. INTRODUCTION**

39 Disintegration and dissolution profiles are critical quality attributes assessed to evaluate
40 drug release performance (Dressman and Krämer, 2005; Huang et al., 2011; Nickerson et al.,
41 2018). Disintegration is often the rate determining step for drug release, particularly for
42 poorly water soluble drugs (Caramella et al., 1988). Disintegration is the mechanical
43 fragmentation of the compressed tablet into small granules or agglomerates. Disintegration
44 is initiated by liquid penetration in the porous of the compact. Swelling is one of the most
45 accepted disintegration mechanisms, which is characterized by an enlargement of the
46 particles that builds up pressure to fragment the tablet matrix (Markl and Zeitler, 2017). The
47 bonding mechanism during compression and the bonding surface area have a direct impact
48 on tablet disintegration. Swelling depends on an optimal tablet porosity, such that the liquid
49 can enter the tablet matrix, however, the void spaces are too large enough to suppress the
50 swelling action of disintegrants (Desai et al., 2016).

51 The application of modelling approaches to enhance product knowledge has been
52 motivated by quality guidelines as an alternative to iterative testing approaches during
53 formulation development (International Council for Harmonisation, 2012, 2008, 2005a,
54 2005b; Kimura et al., 2013). The percolation threshold model has been used to explain how
55 particle-particle interactions of drug and diluents alters dissolution performance of
56 formulations containing different drug loadings (Bonny and Leuenberger, 1993, 1991). In
57 this context, the percolation threshold is the drug loading in which clusters of the drug span
58 throughout the entire volume of the tablet, i.e. an infinite cluster is formed. When this
59 cluster is formed properties of the blend may undergo significant changes (Leuenberger,
60 1999).

61 Previous studies determined the percolation threshold value from disintegration and
62 dissolution experimental data (Kimura et al., 2007a; Stillhart et al., 2017; Wenzel et al.,
63 2017). These studies experimentally determined critical loadings at which disintegration
64 times undergo a step change. These were then assumed to be the percolation thresholds.
65 However, Kimura, Betz and Leuenberger, 2007 recommended further studies to investigate
66 if the change in disintegration behaviour was linked to the formation of the infinite cluster
67 described by the percolation threshold theory (Kimura et al., 2007b). Since these earlier
68 studies, technological advancements have provided novel techniques to study drug
69 distribution in tablets and tablet disintegration behaviour. These techniques can be key to
70 providing data to support the percolation threshold concepts and the findings of previous
71 studies.

72 Spectral imaging techniques have been used to provide in depth information related to drug
73 distribution in pharmaceutical tablets. These techniques can be used to investigate the
74 cluster formation predicted by the percolation theory. Fourier transform infrared
75 spectroscopy (ATR-FTIR), X-ray diffraction (XRD), and Raman spectroscopy are the main
76 techniques employed (Chan et al., 2005; Kazarian and Ewing, 2013; Miller and Havrilla,
77 2005; Zhang et al., 2005). Among those, advancements in Raman instruments has enabled
78 the technique to rapidly map drug distribution in tablets. Raman imaging instruments have
79 been designed to capture rich spectroscopic data which can be translated to provide high-
80 resolution chemical information for tablets (as low as 1 μm per pixel) and require short
81 acquisition times (approx. 15 min for a tablet of 13 mm diameter) (Ali et al., 2013).

82 Focused Beam Reflectance Method (FBRM) and in-line Particle Video Microscopy (PVM) are
83 innovative techniques that can give real-time in-situ information regarding disintegration
84 and dissolution performance of tablets. FBRM has been used to monitor the rate and the

85 degree of change in the number of particles and particle structures in a process (Barrett et
86 al., 2011; Gregory, 2009; Simon et al., 2019; Zhong et al., 2020). Measurement of the solid
87 particles using FBRM is performed without the need for sampling and performing off-line
88 analysis. The system gives particle count, dimension and shape information in real time by
89 monitoring changes in the system as they occur (Barrett and Glennon, 1999). PVM provides
90 real-time images of the system allowing the user to visually track changes in the solids over
91 time (Barrett and Glennon, 2002). The imaging window measures an area of approximately
92 800 μm by 1100 μm . PVM also records a Relative Backscatter Index (RBI) trend which can be
93 used to track changes in the shape and size of solid particles as well as changes in the solids
94 concentration (Werner et al., 2017). RBI is comparable to turbidity monitoring. Increased
95 RBI indicates a larger amount of solids (Hartwig and Hass, 2018). As tablet disintegration
96 progresses the number of particles in the slurry increase as larger particles fragment.
97 Therefore, as disintegration proceeds, more particles are captured in the image and the RBI
98 increases.

99 FBRM and PVM techniques are commonly used in crystallization studies (mass transfer from
100 solution to solid phase) (Barrett et al., 2011; Hartwig and Hass, 2018; Jiang et al., 2014; Liu
101 et al., 2011; Mitchell et al., 2011; Simon et al., 2019; Simone et al., 2015). FBRM has also
102 been utilized in previous studies for investigating tablet disintegration and dissolution
103 (Coutant et al., 2010; Han et al., 2009; Menning, 2016; Metzler et al., 2017). PVM has the
104 potential to monitor tablet disintegration and dissolution because changes in particle size
105 and shape in suspension are key features observed during tablet disintegration and
106 dissolution.

107 The aim of this study was to investigate if a step change in disintegration and dissolution
108 behaviour was observed for tablets produced with drug loadings below and above a

109 predetermined percolation threshold. The percolation threshold of these systems was
110 determined in an earlier study using the physical principals of blending and compaction
111 (Queiroz et al., 2019). The model system investigated was tablets produced from binary
112 blends of microcrystalline cellulose (MCC) and ibuprofen (IBU) at a range of ibuprofen mass
113 loadings. Tablets were prepared with two different MCC grades; one spray dried and one air
114 stream dried. The percolation threshold values determined were 19.08% w/w and 17.76%
115 w/w IBU for blends with air stream dried MCC and spray dried MCC, respectively (Queiroz et
116 al., 2019). A secondary study aim was to determine if the grade of MCC altered any changes
117 in disintegration and dissolution behaviour observed.

118 In the context of the previous and the present study, percolation threshold is a geometric
119 phase transition in which the concentration of ibuprofen particles is high enough to form a
120 cluster that spans throughout the entire volume of the tablet. When this ibuprofen particle
121 cluster is formed, it is anticipated that a step change in properties of the blend will occur.
122 For example, a reduction in flow, compaction and dissolution would be anticipated with
123 ibuprofen particle cluster formation, as ibuprofen has poor flowability and compressibility
124 properties compared to MCC (Al-Karawi et al., 2018; Liu et al., 2008), and is considerably
125 more hydrophobic (Kawabata et al., 2011).

126 In addition to traditional pharmacopeial disintegration and dissolution techniques, process
127 analytical technologies (PAT) FBRM and PVM were employed to better understand tablet
128 disintegration behaviour, and hence its influence on drug dissolution. Building on the
129 application of Raman imaging to qualitatively identity clusters of ibuprofen particles
130 (Queiroz et al., 2019), the present study demonstrated how Raman spectroscopy can be
131 used to quantitatively determine the size and the number of ibuprofen clusters formed on
132 tablets surfaces and hence confirm the percolation threshold determined from compaction

133 data. The MCC grades studied, have similar specifications; average particle size of 130 μm
 134 and similar bulk density (0.28 - 0.33 g/mL for the air stream dried and 0.25 – 0.37 g/mL for
 135 the spray dried). In the earlier study, Queiroz et al., reported morphological differences
 136 between both grades: air stream dried showed bigger particles with needle shaped
 137 geometry, while spray dried showed smaller particles with a more spherical-shaped
 138 geometry (Queiroz et al., 2019).

139

140 2. MATERIALS

141 Emcocel®90 (spray dried) and Vivapur®102 (air stream dried) were supplied by JRS Pharma
 142 (Weissenborn, Germany) and ibuprofen by Kemprotec (Cumbria, UK). The two MCC
 143 products studied were medium size standard grades with theoretical bulk density of 0.28 -
 144 0.33 g/mL for the air stream dried and 0.25 – 0.37 g/mL for the spray dried MCC. A range of
 145 particulate and bulk powder properties of the batches of ibuprofen, air stream dried MCC
 146 and spray dried MCC used in this study had been previously determined (Table 1) (Queiroz
 147 et al., 2019). Other materials used such as buffer components and HPLC mobile phase were
 148 all supplied by Sigma Aldrich, Ireland.

149 Table 1. Particulate and bulk powder properties of Air stream dried®, Spray dried MCC®, and
 150 ibuprofen. Average values are shown \pm standard deviation (Queiroz et al., 2019).

Property	Spray dried MCC®	Air stream dried®	Ibuprofen
D10 (μm) (n=5)	30.0 \pm 0.25	31.1 \pm 0.30	16.5 \pm 0.08
D50 (μm) (n=5)	111.6 \pm 0.73	118.0 \pm 1.60	54.9 \pm 0.21
D90 (μm) (n=5)	236.8 \pm 1.55	240.0 \pm 2.17	129.0 \pm 1.09
Surface area (m^2/g) (n=3)	1.32 \pm 0.01	1.37 \pm 0.01	0.22 \pm 0.02
True density (g/cm^3) (n=10)	1.58 \pm 0.00	1.57 \pm 0.00	1.12 \pm 0.00
Bulk density (g/cm^3) (n=3)	0.33 \pm 0.00	0.31 \pm 0.00	0.36 \pm 0.01

Relative density	0.21	0.20	0.32
Tapped density (g/cm ³) (n=3)	0.43 ± 0.01	0.40 ± 0.00	0.57 ± 0.01
Hausner Ratio	1.32	1.32	1.58
	(easy flowing)	(easy flowing)	(cohesive)
Flow function coefficient (n=3)	7.0 ± 0.91	6.9 ± 0.00	3.9 ± 0.11
	(easy flowing)	(easy flowing)	(cohesive)

151

152 3. METHODS

153 3.1 Tablet manufacture and characterization

154 Binary blends of MCC and IBU were prepared containing a range of IBU concentrations: 2.5,
 155 5, 7.5, 10, 12.5, 15, 20, and 30 % w/w. Each blend, total weight 300 g, was prepared using a
 156 cube mixer KB 15 (Erweka, Heusenstamm, Germany) at 30 rpm for a duration of 30 min.
 157 Flat, round, 8 mm tablets were manufactured using a 10 punches Piccola rotary tablet press
 158 (Riva, Buenos Aires, Argentina) rotating at 20 rpm. The tablet hardness was controlled to
 159 120 ± 10 N, tablet weight variation to 270 ± 10 mg, and the room air humidity to 50 ± 5 %
 160 and temperature to 19 ± 2 °C.

161 Tablet porosity was determined using Equation 1. Tablet envelope density was determined
 162 dividing the mass by the volume of each tablet. The blend true density values were
 163 previously determined (Queiroz et al., 2019).

$$Porosity = 100 \times \left[1 - \left(\frac{tablet\ envelope\ density}{blend\ true\ density} \right) \right] \quad (Equation\ 1)$$

164 3.2 Raman imaging analysis of tablet surface

165 Drug and excipient distributions on external surfaces and surfaces of internal sections of
 166 tablets were investigated by Raman imaging analysis using a RA802 Pharmaceutical analyser

167 (Renishaw, New Mills, UK). First, reference spectra of air stream dried MCC, spray dried
168 MCC, and ibuprofen were acquired. Then, tablets of the blends of ibuprofen and MCC were
169 screened using the StreamLine™ fast imaging method that acquired around 76,000 spectra
170 over the entire surface of each tablet, with a pixel size of 10µm/20µm, and those spectra
171 were averaged to a single resulting spectrum. The total time of measurement for each
172 individual tablet was 15 min. Images of the drug distribution on the surface of the tablet
173 were generated by non-negative least squares (NNLS) component analysis. Domains of each
174 substance were determined based on the reference spectra acquired for the pure
175 substances. Domains of each substance in the generated images were analysed using
176 Particle Analysis in Renishaw's WiRE software. This software resolves the image domains
177 and determines particle metrics. The numbers of domains of ibuprofen on the entire surface
178 of each tablet and their average equivalent circle diameters were determined.

179 Principal component analysis (PCA) was performed using the average spectra obtained from
180 Raman imaging analysis. Unscrambler X (Camo Analytics, Oslo, Norway) was used to
181 perform the PCA with full cross validation, using the algorithm Singular Value
182 Decomposition (SVN).

183 3.3 Disintegration analysis

184 In vitro disintegration time was determined in water at $37\text{ }^{\circ}\text{C} \pm 2\text{ }^{\circ}\text{C}$, using a tablet
185 disintegration tester ZT42 (Erweka, Edison, USA) which complies with Ph. Eur. 2.9.1
186 (Disintegration of tablets and capsules) (Council of Europe, 2019). Each tablet was placed
187 inside of one basket which were continuously and automatically agitated vertically in the
188 disintegration medium. The disintegration process was observed until the tablets

189 disintegrated into small enough particles that could escape the basket so that no substantial
190 material remained in the basket. Analysis was performed in triplicate.

191 3.4 FBRM and PVM analysis

192 FBRM (FBRM G600) and PVM (PVM V19) (Mettler Toledo, Leicester, England) were used to
193 monitor tablet disintegration using a Mettler Toledo Easymax™ 102 system. The
194 disintegration medium was phosphate buffer (pH 7.2). The system used consisted of 100 mL
195 glass vessels with automated internal temperature and agitation control. System specific
196 PTFE (polytetrafluoroethylene) lids allowed for integration of the FBRM and PVM probes. A
197 visual check of the system was possible through an inspection window at the front of the
198 system.

199 The working volume of the system was 50 mL. Experiments were performed at 37 °C and
200 the agitation rate was 250 rpm using an upward pumping, pitch blade impeller for a
201 minimum of 10 minutes after the tablet was added to the vessel. The powder or tablet was
202 added to the glass vessel under agitation. PVM and FBRM monitoring was performed
203 throughout the duration of the experiment; FBRM data was recorded every 2 seconds and
204 two PVM images were recorded every second.

205 3.5 Dissolution studies

206 The dissolution studies were carried out using a DT 600 dissolution tester of Ph. Eur. 2.9.3
207 (paddle) (Erweka, Edison, USA). A volume of 500 mL of phosphate buffer pH 7.2 equilibrated
208 at 37 °C was used as the dissolution medium and the paddle rotation was kept at 50 rpm.
209 Solubility of ibuprofen in the given conditions is 3.74 mg/ml (Dabbagh and Taghipour, 2007).
210 The experiment was conducted using sink conditions; the theoretical concentration of
211 ibuprofen in the dissolution medium following complete dissolution of 30 % w/w ibuprofen

212 tablets was 0.16 mg/ml. Following addition of the tablet sample to the dissolution medium,
213 samples of 0.5 ml volume were withdrawn at 1, 5, 15, 30, 60 and 120, 180, and 240 min
214 intervals in order to determine the dissolution profiles. An additional sample was taken at
215 the 24 h time point to determine the total amount of drug in each tablet tested. At the 24 h
216 time point the tablet had completely disintegrated and complete IBU dissolution was
217 assumed.

218 All samples were filtered with 0.45 µm filter and 0.5 mL of fresh, pre-warmed medium was
219 immediately added to the system in order to correct the volume to the sample volume
220 withdrawn. Samples were analysed by HPLC. The % cumulative amount IBU released was
221 calculated and plotted against time.

222 HPLC analysis was performed using an Agilent 1200 series HPLC system with an UV/Vis
223 detector (Agilent Technologies, Santa Clara, USA). A reversed-phase column (Gemini C-18,
224 250 × 4 mm x 5 µm, Phenomenex Ltd. UK), mobile phase of acetonitrile and water (60:40,
225 pH adjusted to 2.5) at a flow rate of 1.5 ml/min and injection volume of 20 µL were
226 employed. The wavelength for Ibuprofen detection was set at 215 nm and retention time
227 was 7 min.

228 **4. RESULTS**

229 4.1 Tablet Characterisation

230 The average content of ibuprofen was determined for all tablets analysed and compared to
231 the theoretical content (Table 2). Greatest variance between actual and theoretical content
232 was measured for the 30 % w/w ibuprofen loading. Drug content uniformity was also
233 determined with the percentage relative standard deviation less than 7 % for all drug
234 loading. Tablet porosity was also determined (Table 2) as it can influence tablet

235 disintegration and dissolution (Ibrahim, 1985; Yassin et al., 2015). Porosity of tablets
236 decreased as ibuprofen content increased. The porosity of tablets containing spray dried
237 MCC was slightly greater than tablets containing the air stream dried MCC at all drug
238 loadings except for 15%.

239

240

Table 2. Tablet ibuprofen theoretical and average actual content \pm % relative standard deviation (n=5), blend true density and tablet porosity

Theoretical drug concentration (% w/w)	Theoretical drug content (mg)	Air stream dried MCC			Spray dried MCC				
		Actual drug content (mg)	\pm	% RSD	Porosity (%)	Actual drug content (mg)	\pm	% RSD	
2.5 %	6.75	6.63	\pm	2.60 %	30.3	6.82	\pm	1.57 %	32.3
5 %	13.50	13.40	\pm	2.21 %	30.9	13.17	\pm	2.79 %	31.1
10 %	27.00	27.33	\pm	2.81 %	28.5	27.06	\pm	1.38 %	29.7
15 %	40.50	41.68	\pm	1.78 %	28.6	41.94	\pm	5.02 %	28.5
20 %	54.00	54.13	\pm	4.26 %	26.1	55.04	\pm	2.01 %	28.4
30 %	81.00	78.33	\pm	6.77 %	23.1	85.34	\pm	2.00 %	24.10

Commented [CA1]: New table with True Density of blends

241

242 4.2 Raman imaging analysis of tablet surface

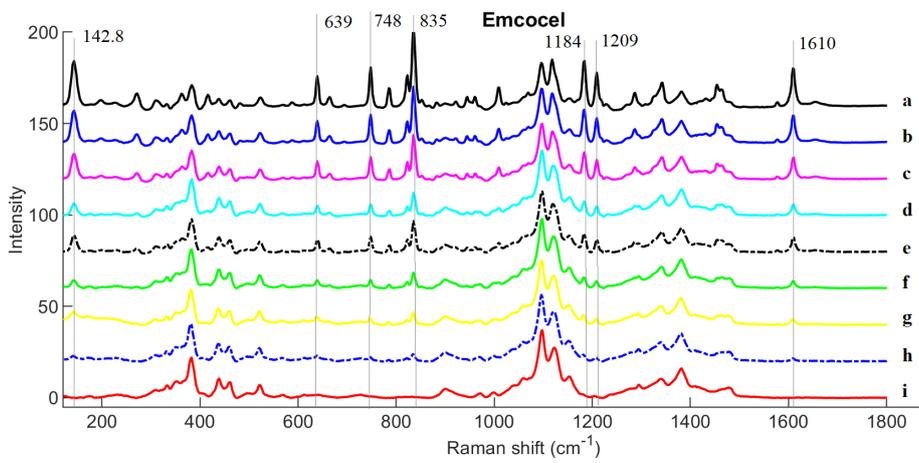
243 Raman spectroscopy did not show differences between the characteristic bands of the MCC
244 tablets (air stream dried and spray dried MCC only), indicating similar chemical identity of
245 microcrystalline cellulose between grades. In respect to the ibuprofen loading, spectral
246 peaks related to ibuprofen increased in intensity when ibuprofen loading was increased
247 (Figure 1).

248 Raman images for tablets of all drug loadings were previously published (Queiroz et al.,
249 2019). In this study the number of ibuprofen domains on surfaces of each tablet was
250 determined from these Raman images, as described in section 3.2 The number of ibuprofen
251 domains decreases for the tablets with drug loading above 15 % w/w (Table 3), despite an
252 increase in the overall intensity peaks related to ibuprofen (Figure 1). Above the percolation
253 threshold the domains of ibuprofen start to connect to the neighbouring ibuprofen
254 domains. Thus, one single domain with larger area is counted, instead of numerous smaller
255 neighbouring domains. In the case of the compacted tablets, it results in a change in the
256 drug distribution from dispersion of drug particles in a matrix of MCC (large number of small
257 drug domains) to distribution of MCC in a matrix of drug (smaller number of larger drug
258 domains). The resulting larger domains were characterized by continuously increased
259 equivalent circular diameter of ibuprofen domains with a more pronounced increase
260 between the concentrations of 15% and 20% w/w ibuprofen (Table 3). These results build
261 on the results in the earlier study which qualitatively confirmed the percolation threshold
262 values determined by visual appearance, which can be subjective. In this study the
263 quantitative data obtained related to the number and size of ibuprofen domains provides a
264 less subjective confirmation of the percolation threshold value. It is important to consider

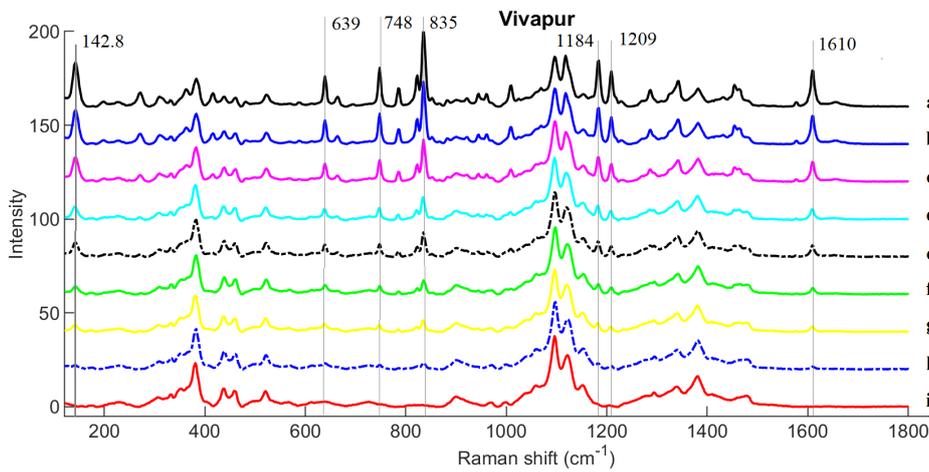
265 the presence of these larger clusters of drug as they can alter the tablet disintegration and
266 drug dissolution behaviour.

267

A



B



268 Figure 1. Average Raman spectra of the surface of (A) Spray dried MCC® and (B) Air stream
269 dried® tablets containing a range of ibuprofen loadings: (a) 30 %, (b) 20 %, (c) 15 %, (d) 12.5

270 %, (e) 10 %, (f) 7.5 %, (g) 5 %, (h) 2.5 %, and (i) 0 % ibuprofen w/w. Vertical lines indicate
271 characteristic peaks of ibuprofen.

272

273 Table 3. The number of ibuprofen domains (N) and the equivalent circular diameter (ECD) of
 274 ibuprofen domains on the surface of Spray dried MCC® and Air stream dried® tablets
 275 containing a range of ibuprofen loadings (2.5 to 30% w/w ibuprofen). The number of
 276 domains (N) and the equivalent circular diameter values were determined from images
 277 generated using Raman image analysis.

% w/w Ibuprofen	Air stream dried®		Spray dried MCC®	
	N	ECD of ibuprofen domains (µm)	N	ECD of ibuprofen domains (µm)
2.5 %	158	69.2	130	62.0
5 %	264	70.4	218	59.6
7.5%	264	76.7	343	74.4
10 %	462	79.5	475	71.8
12.5%	377	74.3	469	77.1
15 %	513	90.5	483	88.6
20 %	377	103.5	376	103.4
30 %	112	153.7	72	138.9

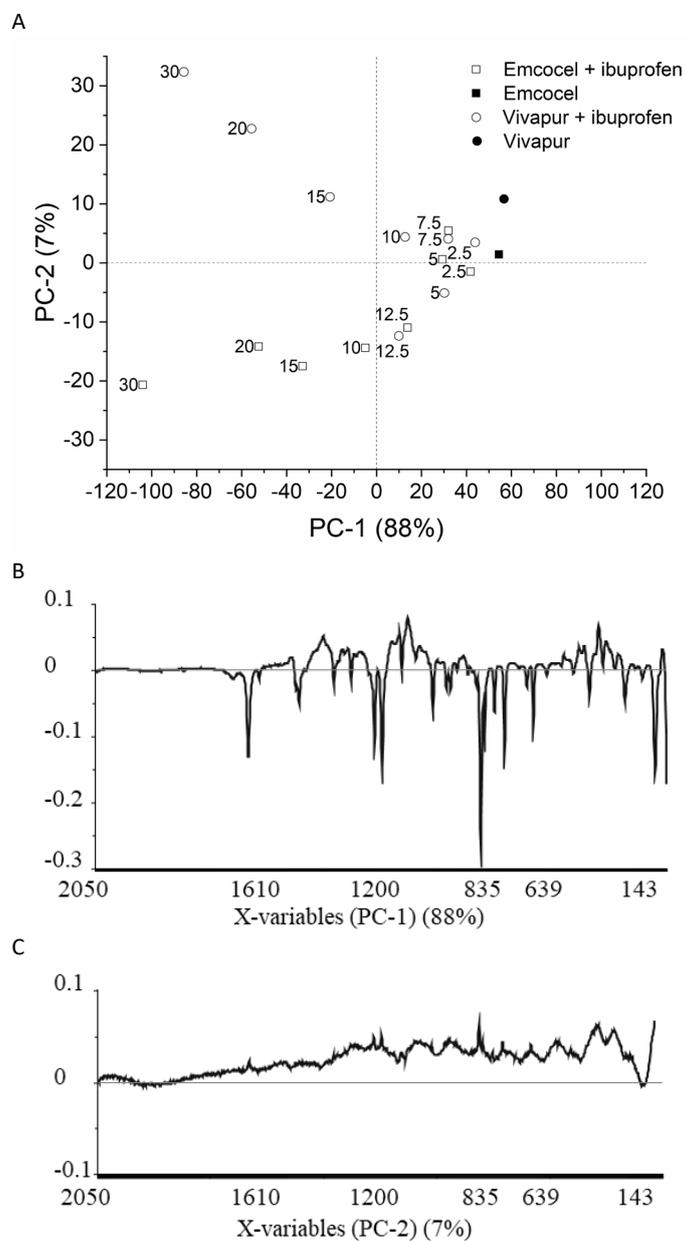
278
 279 PCA analysis of Raman spectra of the surface of tablets showed that the first principal
 280 component (PC-1) captured the effect of drug loading, while the second principal
 281 component (PC-2) captured variability within the samples due to both MCC grade and due
 282 to ibuprofen drug loading. The scores plot showed samples that are similar or different from
 283 each other, i.e. samples geometrically located at distance are dissimilar to each other while
 284 neighbouring samples are similar (Figure 2a). PC-1 and PC-2 explained 88% and 7% of the
 285 variance captured by the model, respectively. Raman shifts of 93, 142.8, 639, 748, 835,
 286 1184, 1209, and 1610 cm⁻¹ were the variables that mostly contributed to discriminate the
 287 samples along the first component of the model (PC-1) (Figure 2b). Those bands are

288 characteristic of ibuprofen (Sütő et al., 2016) and they were not observed in the spectra of
289 the tablets containing pure MCC (Figure 1). As stated previously, these peak heights
290 increased with the increase in drug loading. The PCA model showed that tablets containing
291 air stream dried MCC differed from tablets containing spray dried MCC along PC-2. Loadings
292 of PC-2 contained peaks assigned to both, ibuprofen and MCC. PC-2 also showed an
293 upwards shifting of the baseline reduction in Raman shift. Both chemical and physical
294 attributes of the tablets may explain this variability in Raman spectra. Differences in tablet
295 porosity was observed with increase in drug loading and between MCC grades, Table 2.
296 Raman spectra can show stronger intensities for samples for more compacted (less porous)
297 samples due to an increased number of scattering molecules that will produce a Raman
298 signal (<https://doi.org/10.1016/j.vibspec.2018.10.011>). The upwards shift may also be due to Raman
299 fluorescence, which is a material-dependant phenomenon; fluorescence is phenomena
300 intrinsic to MCC. Microcrystalline cellulose is known to be fluorescent mainly due to the
301 presence of lignin (Castellan et al., 2007). Variance related to ibuprofen peaks (e.g. at the
302 shifts 835 and 1610 cm^{-1}) and the main characteristic Raman bands assigned to cellulose
303 (e.g. at 1096 cm^{-1} and within the region 275-550 cm^{-1}) (Wiley and Atalla, 1987) are also
304 present in the loadings of PC-2. The region of 275-550 cm^{-1} is known to hold crystallinity
305 information of cellulosic materials (Agarwal et al., 2010). Thus, the separation of the

Commented [CA2]: <https://doi.org/10.1016/j.vibspec.2018.10.011>
Gomez et al

306 samples along PC-2 may be an indication that the differences between the spray dried and
307 the air stream dried MCC grades included crystallinity, lignin content, and compact density.

308



309 Figure 2 (A) Scores plot and (B and C) loading plots of the principal component analysis of
 310 Raman spectra acquired from spray dried MCC and air stream dried tablets and tablets
 311 containing a range of ibuprofen loadings (2.5 % to 30 % w/w ibuprofen). PC-1 and PC-2 are
 312 the first and the second principal components, respectively.

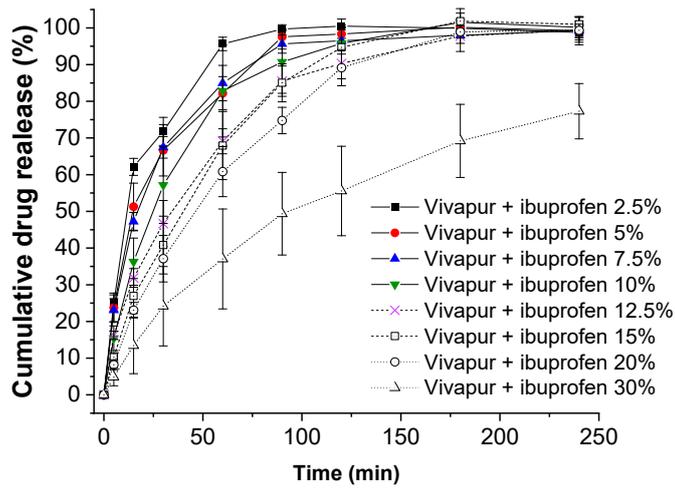
313 4.3 Disintegration and Dissolution

314 Tablet disintegration using Ph. Eur. 2.9.1 disintegration apparatus showed that all tablets
315 had completely disintegrated in less than 5 min. Differences between tablets containing
316 different drug loadings or MCC grades could not be accurately determined. The tablets
317 investigated contain a high percentage in mass of MCC, which is highly hydroscopic and a
318 noted disintegrant (Rowe et al., 2009). Thus, disintegration happened fast independently of
319 the MCC grade.

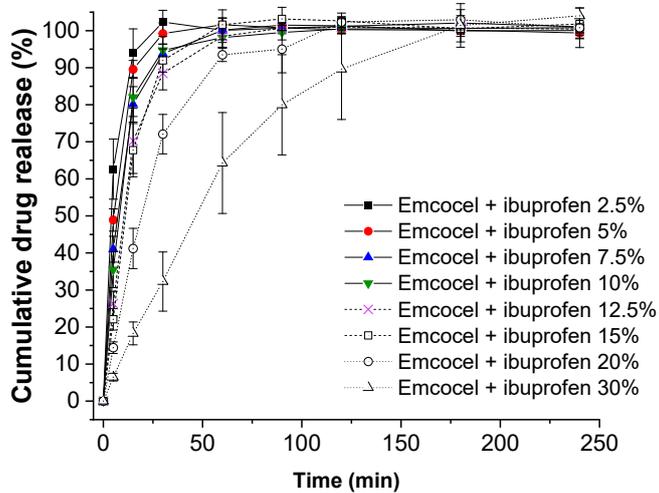
320 The results of the Ph. Eur. 2.9.3 dissolution study showed that increased ibuprofen
321 concentration had a negative impact on the dissolution behaviour of Ibuprofen/MCC tablets
322 (Figure 3). The effect of ibuprofen loading on drug dissolution was evident for ibuprofen
323 concentrations above the percolation threshold, 20 and 30 %w/w of ibuprofen; time to
324 achieve 100% ibuprofen release increased significantly (Figure 4). Tablets containing the air
325 stream dried MCC showed statistically significantly greater times to achieve 100% ibuprofen
326 release for all drug loadings in comparison to tablets containing the spray dried MCC.
327 Tablets containing 30% w/w IBU and air stream dried MCC did not reach %100 release in
328 240 min. Complete release was confirmed after 24h. It is also interesting to note the step
329 change in time to reach 100% cumulative drug release between 2.5 and 5% drug loading (air
330 stream dried MCC) and 2.5 and 7.5% drug loading (spray dried MCC respectively) (Figure 4).
331 This change in dissolution behaviour is not related to a percolation threshold of ibuprofen in
332 the MCC matrix but may be due to other factors such a differences in porosity influencing
333 disintegration behaviour (Desai et al., 2016) and hence dissolution.

334

A

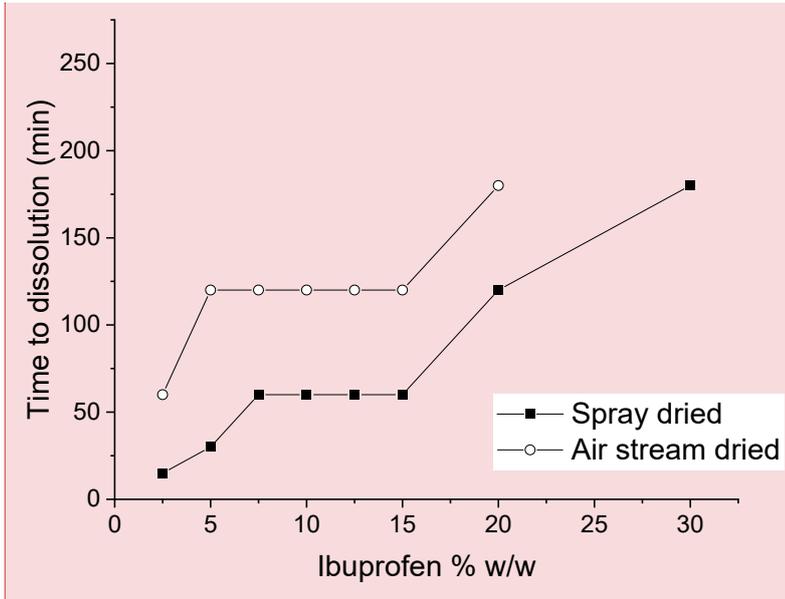


B



336 Figure 4. Dissolution profiles of tablets containing (A) Spray dried MCC® and (B) Air stream
 337 dried® and different ibuprofen w/w/ loadings (2.5 to 30% w/w). Dissolution was performed
 338 in phosphate buffer pH 7.2 at 37°C. Average values shown with γ -error bars indicating
 339 standard deviation, n= 5.

340



Commented [CA3]: Can you rescale Y axis to 200 min

341

342 Figure 4. Time to reach 100% ibuprofen release during dissolution of tablets containing
343 spray dried and air stream dried microcrystalline cellulose grades and different ibuprofen
344 w/w/ loadings (2.5 to 30% w/w). Tablets containing air stream dried MCC and 30% w/w of
345 ibuprofen did not reach %100 release in 240 min. However, complete release was confirmed
346 after 24h. Dissolution was performed in phosphate buffer pH 7.2 at 37°C. Average values
347 shown with y-error bars indicating standard deviation, n= 5.

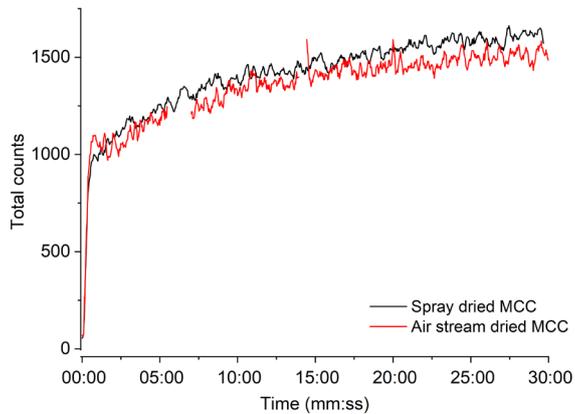
348

349 4.4 FBRM monitoring

350 FBRM was used as a PAT tool to determine if tablet disintegration played a role in the
351 differences observed between dissolution of tablets containing air stream dried and spray
352 dried MCC grades with increasing drug loading. Two aspects were investigated: the
353 differences among tablets below and above the percolation threshold and the differences
354 between both MCC grades. This analysis was complementary to the pharmacopoeial
355 disintegration test which was not able to capture differences regarding these two aspects.
356 As mentioned previously, FBRM gives particle count, dimension information in real-time

357 (Barrett and Glennon, 1999). It was hoped that the ibuprofen clusters observed in the
358 tablets by Raman imaging could be observed in the disintegration medium and potentially
359 explain the differences in dissolution observed between tablets containing spray dried and
360 air stream dried MCC grades at different drug loadings.

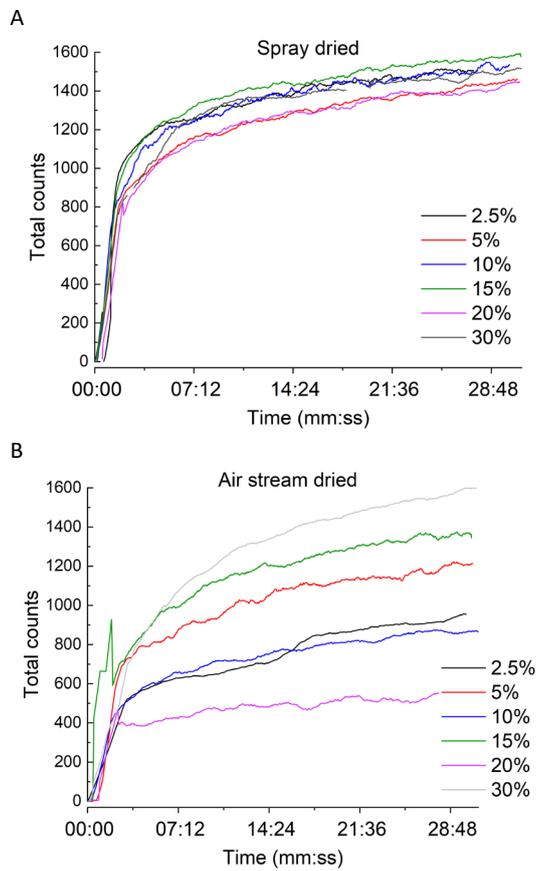
361 Initially, disintegration monitored by FBRM was performed using spray dried and air stream
362 dried MCC tablets without ibuprofen to determine differences in disintegration due to MCC
363 grade. Both tablets displayed very similar behaviour with a sharp increase in particle counts
364 upon addition of the tablet to the phosphate buffer pH 7.2. Figure 5 shows that the counts
365 vs time profile for the two MCC grades were very similar, indicating that the tablets
366 disintegrated at the same rate.



367 Figure 5. Focused Beam Reflectance Measurement (FBRM) counts 1-1000 μm versus time
368 for tablets containing air stream dried and spray dried MCC in phosphate buffer pH 7.2, and
369 temperature of 37 °C.

371
372 Figure 6 shows the FBRM total counts vs time for ibuprofen tablets added to the
373 disintegration medium for the first 30 minutes. For all tablets there is a rapid increase in
374 counts for the first 7 minutes approximately, indicating that, as with the tablets of air

375 stream dried and spray dried MCC without ibuprofen, tablet disintegration began
376 immediately upon addition of the tablet to the medium for all ibuprofen loadings. A similar
377 profile is seen for each tablet of spray dried MCC regardless of ibuprofen loading. The
378 counts increased sharply in the first 30 seconds after addition to the medium and then
379 continued to increase at a slower rate for the following minutes.



380
381 Figure 6. Focused Beam Reflectance Measurement (FBRM) total particle counts (counts 1-
382 1000 μm) versus time for tablet containing (A) the spray dried MCC and (C) the air stream
383 dried MCC and a range of ibuprofen loading (% w/w) in phosphate buffer pH 7.2 and
384 temperature of 37 °C.

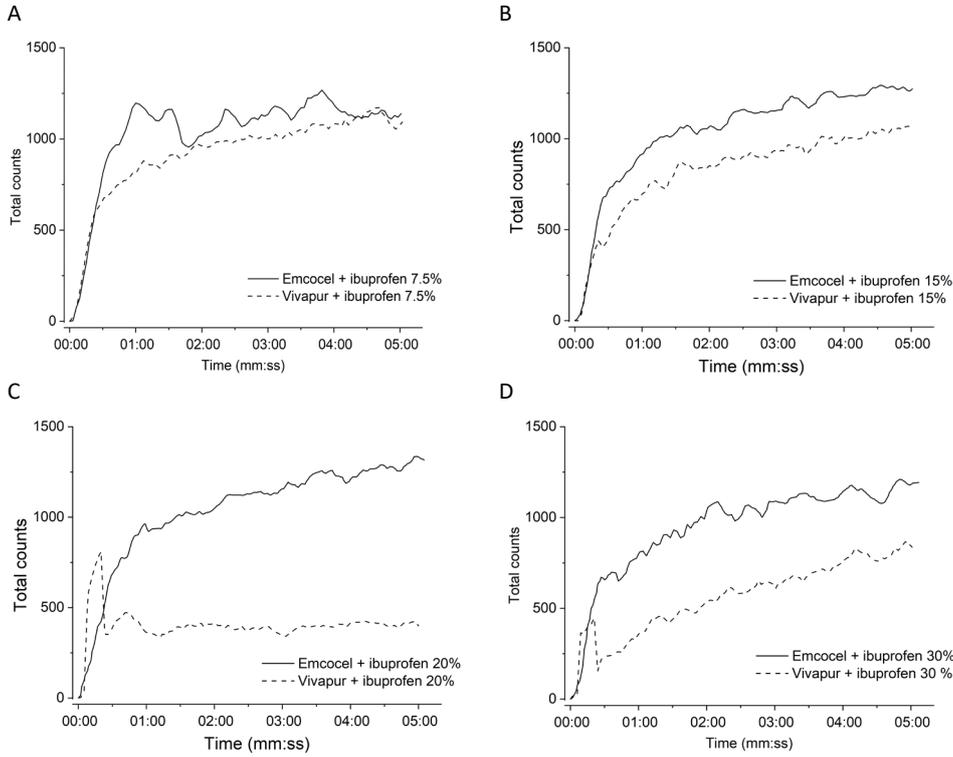
385 Overall, air stream dried MCC tablets had a much less consistent total counts profile over
386 time for different drug loading, in comparison to spray dried MCC tablets. When the
387 different ibuprofen loadings were compared, the loadings of 20 % and 30 % w/w ibuprofen
388 showed different profiles compared to lower drug loadings. For 20 % and 30 % w/w
389 ibuprofen, a sharp increase in total counts was observed upon addition of tablet and
390 followed by a rapid decrease. For tablets of air stream dried MCC at greater ibuprofen
391 loadings the total count was lower after 5 minutes compared to lower tablet loadings.

392 The total counts profiles for tablets containing air stream dried MCC was also lower in
393 comparison to tablets containing spray dried MCC, for all ibuprofen loadings (Figure 7).

394 These differences were more pronounced above the percolation threshold > 15% ibuprofen.

395

396



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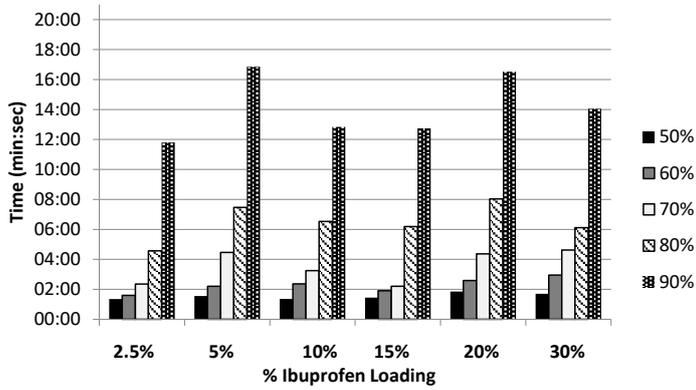
398 Figure 3. Focused Beam Reflectance Measurement (FBRM) total counts over time for
399 Vivapur® and Emcocel® tablets containing ibuprofen (A) 7.5 %, (B) 15 %, (C) 20 %, and (D)
400 30% w/w in phosphate buffer pH 7.2 and temperature of 37 °C.

401

402 Due to the variability in the number of counts for each tablet a relative increase in FBRM
403 counts was measured for each tablet to enable comparison between tablets. Figure 8 shows
404 the time necessary to reach 50, 60, 70, 80, 90% total counts. All tablets were tracked for 30
405 minutes. Thus, the total counts of 100% were considered the total counts at 30 min. The
406 overall trend showed a longer time for tablets containing air stream dried MCC compared to
407 spray dried MCC, indicating a slower disintegration rate.

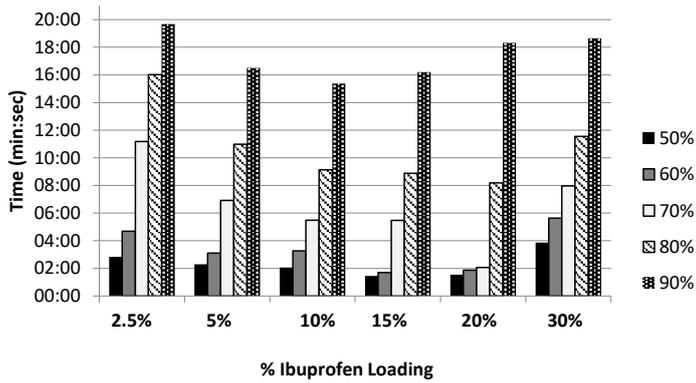
408

A



409

B



410

411 Figure 8. Time to reach percentage of total particle counts measured by FBRM for tablets
412 after containing (A) spray dried and (B) air stream dried microcrystalline cellulose grades
413 and different ibuprofen w/w/ loadings (2.5 to 30% w/w) disintegrating in phosphate buffer
414 pH 7.2 at 37°C. Percentages expressed relative to total particle counts at 30 mins considered
415 100%.

416

417 An indication of particle size distribution following disintegration was obtained from the

418 FBRM chord length distributions and square weighted chord length distributions (SQWT).

419 Representative tablets with 12.5, 15 % w/w, and 30 % w/w ibuprofen loadings after 5

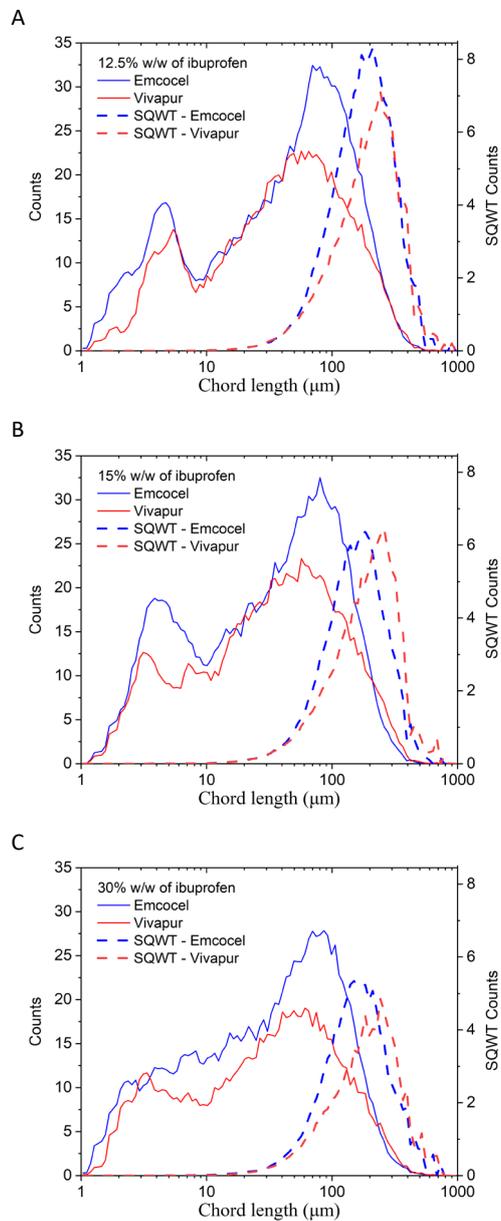
420 minutes of dissolution are shown in Figure 9 for the spray dried and air stream dried MCC
421 grades. CLD and SQWCLD for all drug loadings investigated are available in the
422 supplementary material. These distributions are automatically generated for the user by the
423 iC FBRM software. The CLD is comparable to a particle size distribution. This is the number
424 of chord lengths recorded in the measurement scan time in vs the chord length. The
425 SQWCLD is useful for visual comparison of systems by emphasising differences in the course
426 counts (100 – 1000 μm). This is achieved by applying a channel (size intervals or bins)
427 specific weight w_i to counts n_i . The weighted channels y_i are obtained via:

$$y_i = w_i \cdot n_i \quad (\text{Equation 2})$$

428 The weights w_i are obtained from the channel midpoints M_i via:

$$w_i = \frac{M_i^\gamma}{\sum_{j=1}^N M_j^\gamma} \cdot N \quad (\text{Equation 3})$$

429 Where γ is 2 for the square weight, N is the number of channels, which was 90 in this study,
430 $i = 1, 2, \dots, N$ and $j = 1, 2, \dots, N$.

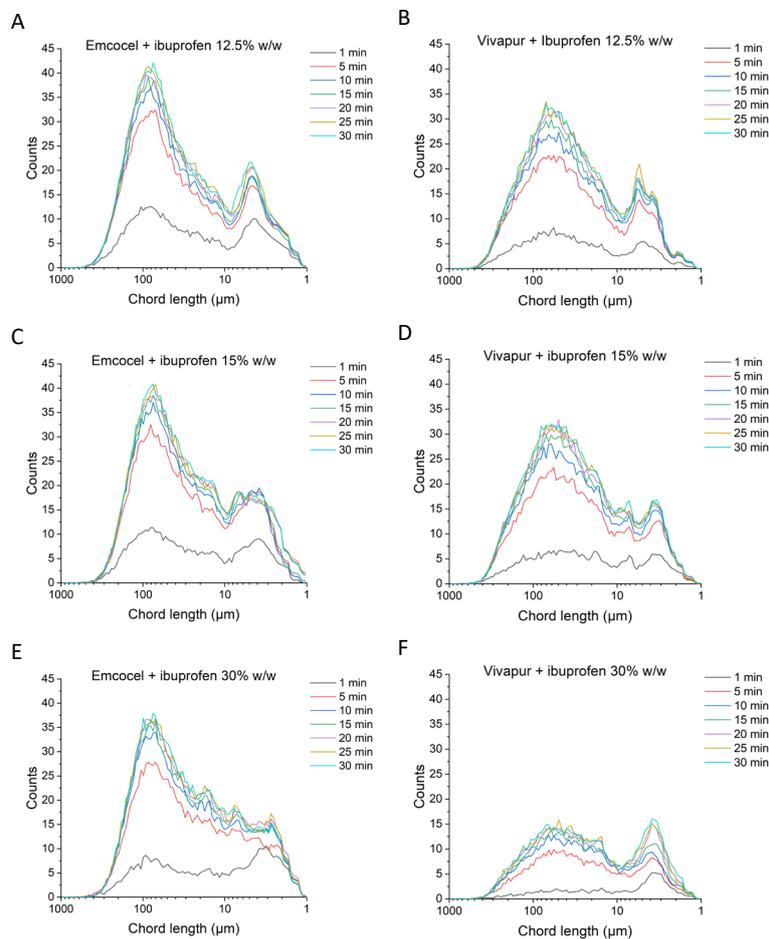


431 Figure 9. Focused Beam Reflectance Measurement (FBRM) chord length distributions and
 432 square weighted chord length distributions for Spray dried MCC® (spray dried MCC) and Air
 433 stream dried® (air stream dried MCC) tablets with ibuprofen loading (A) 12.5 %, (B) 15%,
 434 and (C) 30 % w/w, 5 minutes after addition to the disintegration medium, phosphate buffer
 435 pH 7.2 and temperature of 37 °C.

436 Coarse counts account for a much larger proportion of the mass of material compared with
437 fine counts (1-10 μm). While the CLDs for both systems have a similar shape profile the
438 increased number of total counts and shorter chord length counts was evident for the spray
439 dried compared to the air stream dried MCC. Fine counts may be related to disaggregation
440 of MCC particles which is composed of cellulose fibrils agglomerated into larger particles
441 (Queiroz et al., 2019.) When the square weighted CLDs are compared there is a distinct shift
442 to the right for air stream dried, highlighting the increased particle size present 5 minutes
443 after the tablet addition to the buffer. An increase in fine counts (1-10 μm) present in the
444 spray dried MCC system suggested that tablets of the spray dried MCC disintegrated more
445 effectively than tablets of the air stream dried MCC at a 30 % w/w ibuprofen loading. For
446 loading below the percolation threshold, a similar trend was seen although the shift to the
447 right for air stream dried in the square weighted CLD is less pronounced. This is exemplified
448 by 12.5 % w/w ibuprofen tablets in Figure 9.

449 The plots of chord length distributions at different time points during disintegration for each
450 individual tablet were generated (supplementary data) and selected tablets shown in Figure
451 10. The increase in counts over time happens similarly across all chord lengths for a same
452 tablet, i.e. the distribution did not show a shape change at different time points.
453 Interestingly, the increase in count is clearly more significant up to 5 min. After that, the
454 change in counts is comparatively very small.

455



456 Figure 10. Focused Beam Reflectance Measurement (FBRM) chord length distributions for
 457 tablets with ibuprofen loadings of 12.5 %, 15%, and 30 % w/w and (A, C, and D) Spray dried
 458 MCC® (spray dried MCC) and (B, D, and F) Air stream dried® (air stream dried MCC),
 459 respectively, at different times after addition to the disintegration medium, phosphate
 460 buffer pH 7.2 and temperature of 37 °C. This analysis is available for tablets at all drug
 461 loadings in the supplementary material.

462

463 Based on the results shown in Figure 11, the cumulative drug release at 5 min was plotted
 464 against IBU concentration in order to investigate the differences in drug release below and
 465 above the threshold following tablet disintegration (Figure 5). Interestingly, after 5 min time

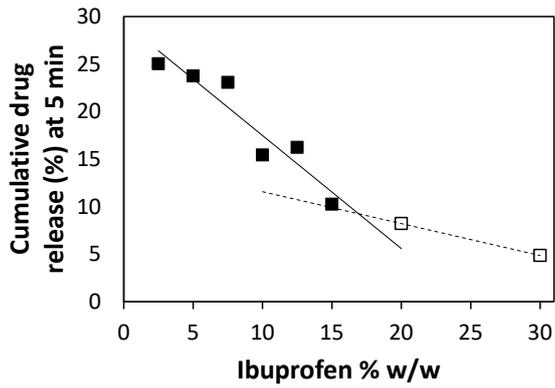
466 point the pharmacopeial disintegration tested showed complete disintegration. Similar
467 graphical approaches have been previously used to determine the percolation threshold
468 from disintegration and dissolution (Kimura et al., 2007a; Wenzel et al., 2017).

469 Cumulative release decreased sharply from 2.5 to 15% w/w of ibuprofen loading. However,
470 at drug loadings above the percolation threshold (20% w/w and 30% w/w IBU) the reduction
471 in drug release with increase in drug loading was decreased. Tablets containing the air
472 stream dried MCC showed significantly lower drug release for all drug loadings in
473 comparison to tablets containing the spray dried MCC. These findings showed the change in
474 dissolution behaviour at drug concentrations above and below the predicted percolation
475 threshold following tablet disintegration.

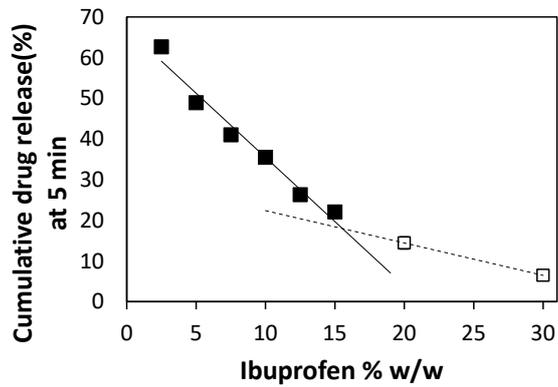
476

477

A



B



478

479 Figure 11. Estimation of percolation threshold based on the dissolution cumulative release
480 of ibuprofen from the tablets containing (A) air stream dried and (B) spray dried
481 microcrystalline cellulose, at 5 min of dissolution. The time of 5 minutes was chosen as all
482 tablets had disintegrated at 5 min based on FBRM results.

483

484

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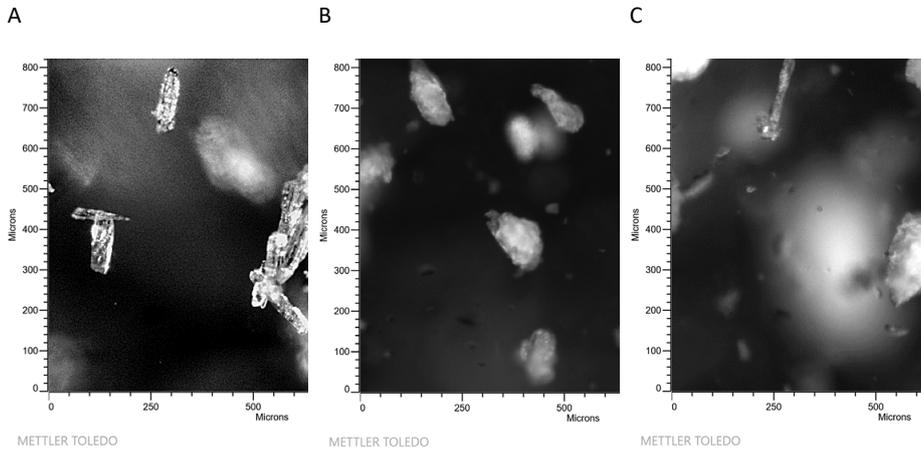
486 4.5 PVM analysis

487 PVM analysis was performed to provide real-time images of particles in dissolution medium
488 during tablet disintegration. Besides the images, PVM relative backscatter index was used as
489 a quantitative measure of disintegration. Initially, images were collected using the PVM
490 probe for ibuprofen powder and both MCC only tablets in buffer. The ibuprofen particles
491 had a distinct rod-shaped habit (Figure 12a). Dispersed ibuprofen powder was present as
492 both discrete particles and aggregates. The ibuprofen particles appeared to be between 100
493 and 300 μm in length and 30 and 50 μm in width. When compared with the particle size for
494 the dry powder from laser diffraction (Table 1), where the D50 was 55 μm , it appears that
495 larger ibuprofen particles in the PVM images may be aggregates.

496 PVM images of the disintegrated spray dried MCC tablet (Figure 12b) indicated that fine
497 material was present along with uniform distinct particles having a rough surface. Images of
498 disintegrated air stream dried MCC tablet (Figure 12c) are similar to spray dried MCC. These
499 images would support the presence of fine particles observed during FBRM analysis, Figures
500 9 and 10. These appeared to be rod shaped particles present which are similar in
501 appearance to the ibuprofen particles.

502

503



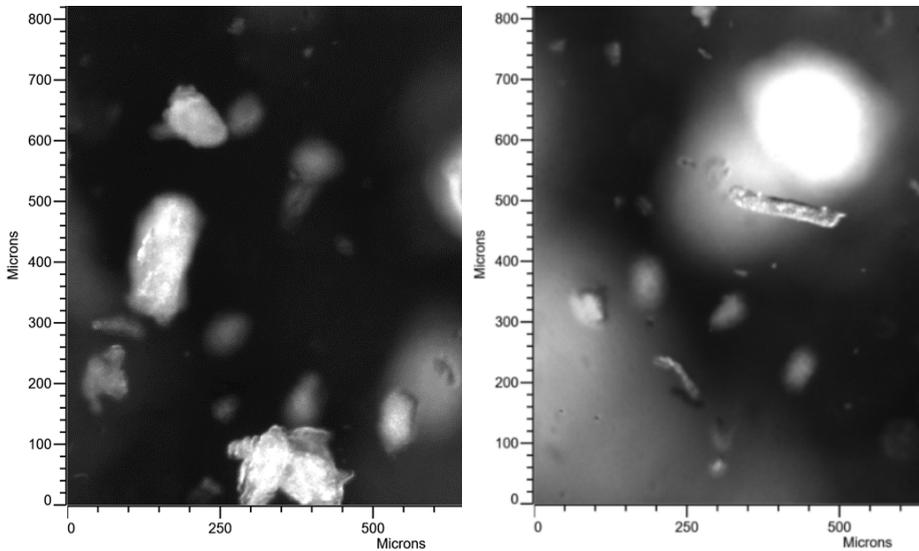
504 Figure 12. PVM images of (A) ibuprofen powder, and (B) Spray dried MCC® and (C) Air
505 stream dried® particles following tablet disintegration in phosphate buffer pH7.2 and
506 temperature of 37 °C.

507

508 Side by side comparison of PVM images of disintegrated tablets containing spray dried and
509 air stream dried MCC with equivalent ibuprofen loading showed that it is difficult to
510 distinguish definite differences between the two systems. One difference noted was that air
511 stream dried tablets showed more elongated particles (Figure 13). There are rod shaped
512 particles present in the air stream dried suspension (Figure 12), hence it was not possible to
513 distinguish whether this rod-shaped material following the 15% w/w ibuprofen tablet
514 disintegration is ibuprofen or MCC.

A

B



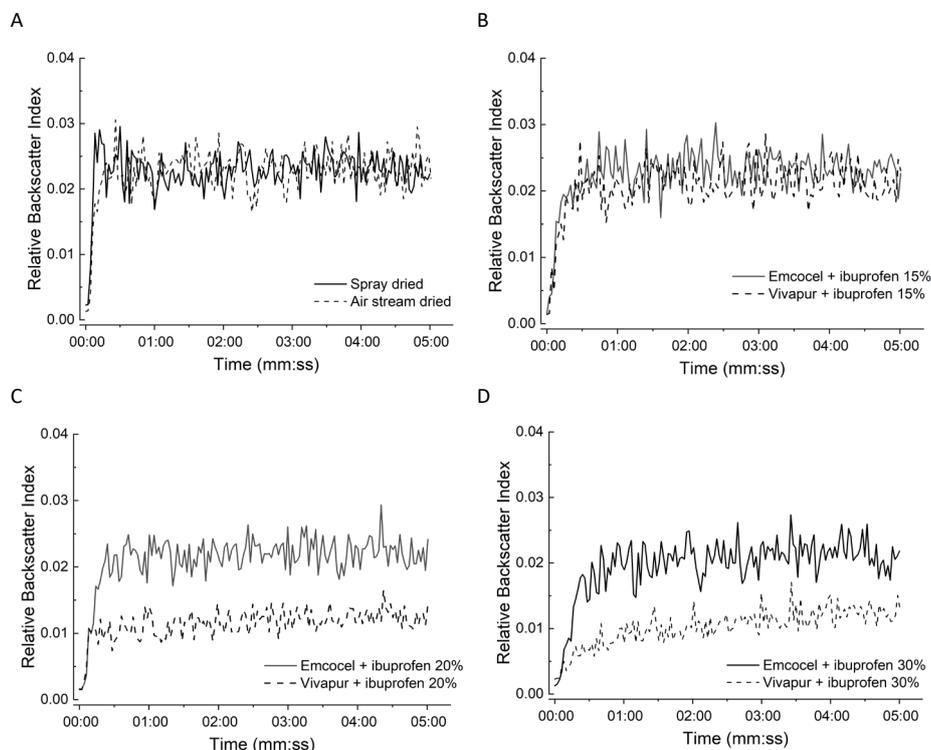
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515 Figure 13. Representative PVM images of particles following disintegration of (A) spray dried
 516 MCC and (B) air stream dried MCC tablets with a 15 % w/w ibuprofen loading in phosphate
 517 buffer pH7.2 and temperature of 37 °C.
 518

519 Relative Backscatter Index (RBI) measured by PVM is the relationship between the incident
 520 and the detected light. As disintegration progresses the number of particles in the media
 521 increases due to fragmentation of larger particles to smaller particles and RBI increases. The
 522 change in the PVM RBI versus time during disintegration does indicate differences for
 523 tablets containing 20% and 30% ibuprofen loading tablets compared to tablets containing
 524 lower ibuprofen loadings (Figure 14). For ibuprofen loadings below the percolation
 525 threshold the RBI for both spray dried and air stream dried MCC tablets was similar. RBI
 526 increased to a greater extent for the spray dried MCC tablets with increased drug loadings
 527 compared to air stream dried MCC (Figure 14d). The air stream dried MCC tablets with a 20
 528 and 30% w/w ibuprofen has a significantly lower final RBI during disintegration.

529



530 Figure 14. Relative Backscatter Index (RBI) vs time following disintegration of Spray dried
 531 MCC® and Air stream dried® tablets (A) 0% w/w, (B) 15% w/w, (C) 20% w/w and (D) 30%
 532 w/w ibuprofen in phosphate buffer pH7.2 and temperature of 37 °C.
 533

534 **5. DISCUSSION**

535 The research presented was conducted to better understand whether a percolation
 536 threshold determined from compaction data can translate to experimental tablet
 537 disintegration and dissolution data and the effect of the presence of a critical drug load
 538 (percolation threshold) on tablet disintegration and drug dissolution.

539 Previous studies determined a percolation threshold value from disintegration and
 540 dissolution experimental data (Kimura et al., 2007a; Stillhart et al., 2017; Wenzel et al.,
 541 2017). The earlier study was focused on predicting mathematically and confirming

542 experimentally the existence of a percolation threshold from a blend properties and
543 compaction behaviour perspective. In the present study, disintegration and dissolution
544 experiments were carried out to experimentally to confirm the percolation threshold, which
545 had been predicted using an statistical hybrid model (Queiroz et al., 2019).

546 Dissolution testing confirmed the presence of the percolation threshold in the region
547 previously reported. The change in behaviour above the percolation threshold was observed
548 during dissolution; % drug released at 5 min following tablet disintegration (Figure 11) and
549 time to achieve complete dissolution (Figure 4). Blends above the percolation threshold
550 showed slower dissolution profiles. Kimura, Betz and Leuenberger, 2007 also revealed a
551 decreased disintegration performance above the critical loading of a poorly water-soluble
552 drug. In the case of tablets containing MCC and ibuprofen, it was hypothesised that the
553 connected MCC particles would form the water-conducting clusters promoting
554 disintegration. Above the threshold predicted a continuous cluster of ibuprofen particles is
555 formed. Relative to MCC, ibuprofen is poorly water soluble, and the formation of
556 continuous ibuprofen clusters would decrease disintegration. Thus, the explanation for the
557 reduction in dissolution above the percolation threshold can be attributed to the combined
558 effect of decreased drug surface area to mass due to the presence of continuous clusters
559 evidenced by Raman imaging and a change in the disintegration process.

560 In this study, Raman imaging and image domain analysis were combined to confirm
561 percolation threshold in pharmaceutical tablets. The methodology developed confirmed the
562 percolation threshold previously predicted for the binary blend investigated (Queiroz et al.,
563 2019) by an increasing number of drug cluster up to the percolation threshold and reduction
564 above due to the formation of continuous clusters. For the drug loading above 15%, the
565 number of ibuprofen domains dramatically decreased, and their equivalent circle diameter

566 increased which confirms the cluster formation and would contribute to a slower rate of
567 ibuprofen dissolution due to a reduced surface area to mass ratio.

568 The influence of the presence of continuous ibuprofen clusters on tablet disintegration was
569 difficult to establish by pharmacopoeial disintegration testing. However, the use of FBRM
570 and PVM to determine the tablet disintegration behaviour with respect to drug loading
571 demonstrated a change in behaviour above the percolation threshold, particularly for
572 tablets containing air stream dried MCC (Figures 7 and 14). PVM images of disintegration
573 showed an omnidirectional enlargement of particles prior fragmentation of the tablet. This
574 is a typical behaviour of immediate-release tablets and characterizes the swelling
575 disintegration mechanism (Caramella et al., 1988). The dissolution of the disintegrated
576 particles could not be observed by FBRM nor PVM due to similarity in morphology of the
577 disaggregated MCC particles and ibuprofen particles (Figure 13) and the insoluble nature of
578 MCC in the disintegration medium. However, it may be possible to monitor drug dissolution
579 for formulations with high drug loadings and soluble excipients, such as lactose, using PVM
580 and FBRM techniques.

581 The secondary objective of this study was to investigate the influence of MCC grade on
582 disintegration and dissolution behaviour relative to the percolation threshold. Despite the
583 MCC grades having similar bulk properties, the interaction of each grade with the model
584 drug ibuprofen resulted in differing dissolution behaviour. In all cases, the tablets containing
585 the air stream dried grade showed slower disintegration and dissolution rates. Air stream
586 dried MCC tablets showed a reduction in disintegration rate above the percolation threshold
587 value while spray dried MCC did not (Figure 6 and 14). These tablets resulted in the slower
588 dissolution rates (Figure 4) across all drug loading. Tablets produced from the air stream
589 dried MCC grade were slightly less porous (Table 2). Tablet porosity is a function of drug

590 loading and for poorly compressible drugs reduced above the percolation threshold value,
591 but also a dependent on the grade of MCC employed (Queiroz et al., 2019). Differences in
592 MCC crystallinity has also been related to lower swelling of MCC; polymers with less dense
593 crystalline regions were observed to swell more as they are more accessible for the water
594 molecules and the cohesive forces between the chain segments are weaker in comparison
595 to the crystalline domains (Desai et al., 2016; Schott, 1992). Further studies are required to
596 determine the exact mechanisms causing in the reduced dissolution rates for tablets
597 containing air stream dried MCC compared to the spray dried grade.

598 It was challenging to discriminate between the effects of tablet porosity and percolation
599 threshold in relation to tablet disintegration. The compaction parameters and bonding
600 mechanism of particles during compaction directly impact tablet porosity, the ingress of the
601 disintegration medium, MCC swelling and hence tablet disintegration (Yassin et al., 2015).
602 Despite confounding differences observed in tablet porosity in this study, a clear step
603 change in dissolution behaviour was observed for tablets with drug loadings above the
604 percolation threshold.

605

606 **6. CONCLUSIONS**

607 Dissolution data showed that a percolation threshold value previously determined for
608 ibuprofen/MCC binary blends, from compaction data, translated to tablet dissolution data.
609 Slower ibuprofen dissolution behaviour was observed for tablets above the predetermined
610 percolation threshold level and confirmed the presence of the percolation threshold
611 relevant to dissolution. In addition, slower dissolution was observed for all tablets
612 containing an air stream dried MCC grade compared to a spray dried MCC grade. FBRM and

613 PVM showed less efficient disintegration above the percolation threshold for tablets
614 containing air stream dried MCC. The results experimentally demonstrate that both larger
615 drug domains, quantified by Raman imaging, and a less efficient tablet disintegration (in the
616 case of air stream dried MCC) measured by FBRM and PVM contributed to slower ibuprofen
617 dissolution profiles above the percolation threshold.

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623 Pharmacy University College Cork.

624

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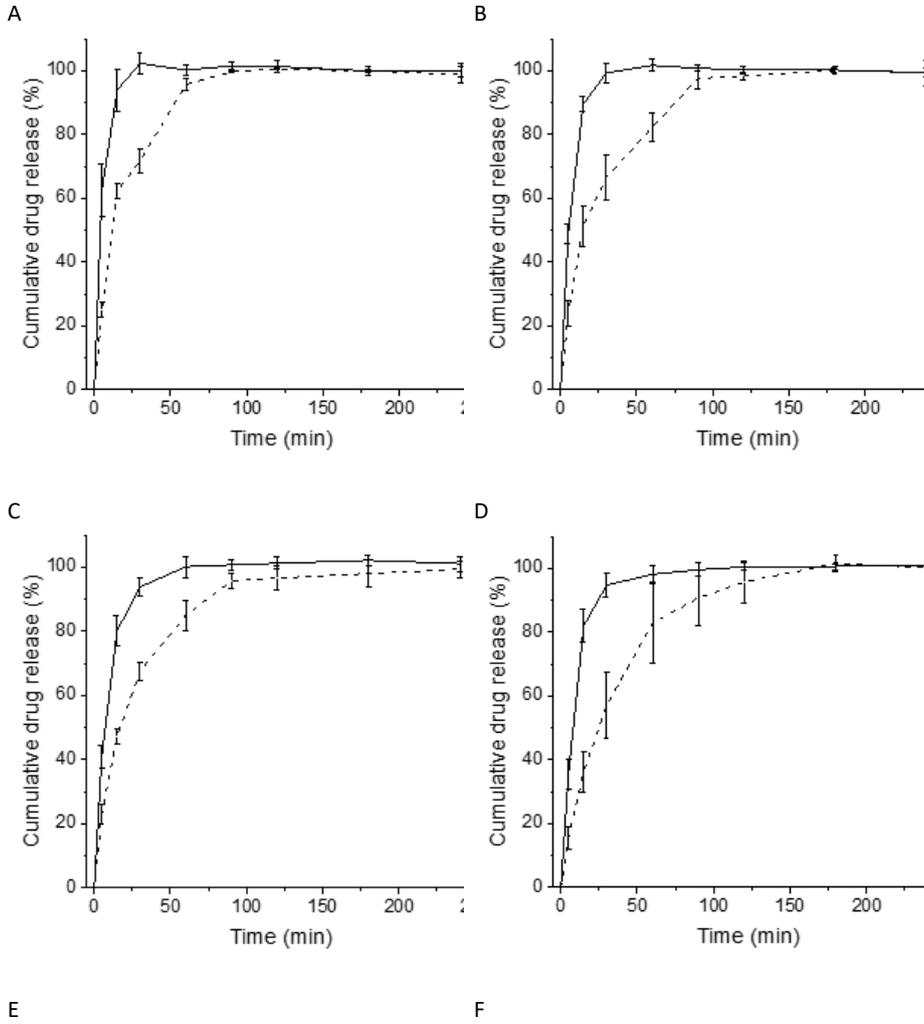
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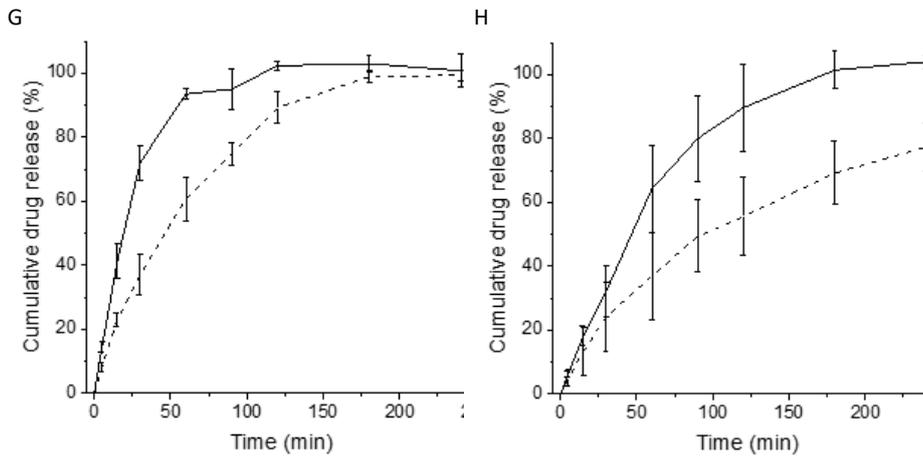
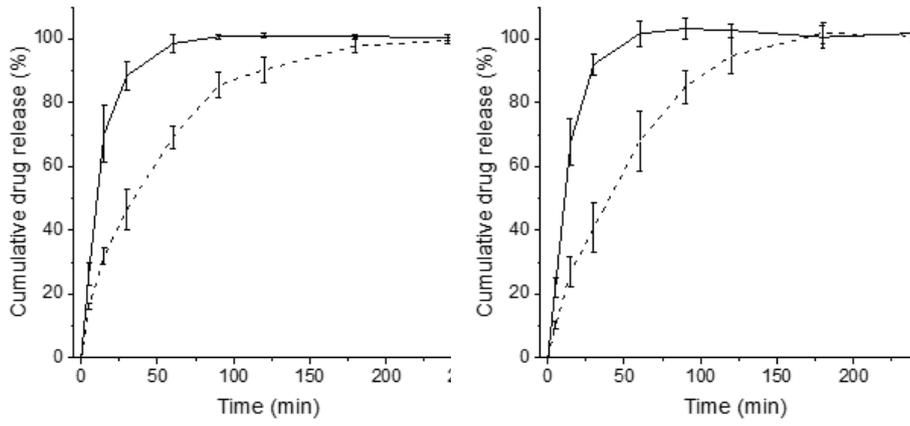
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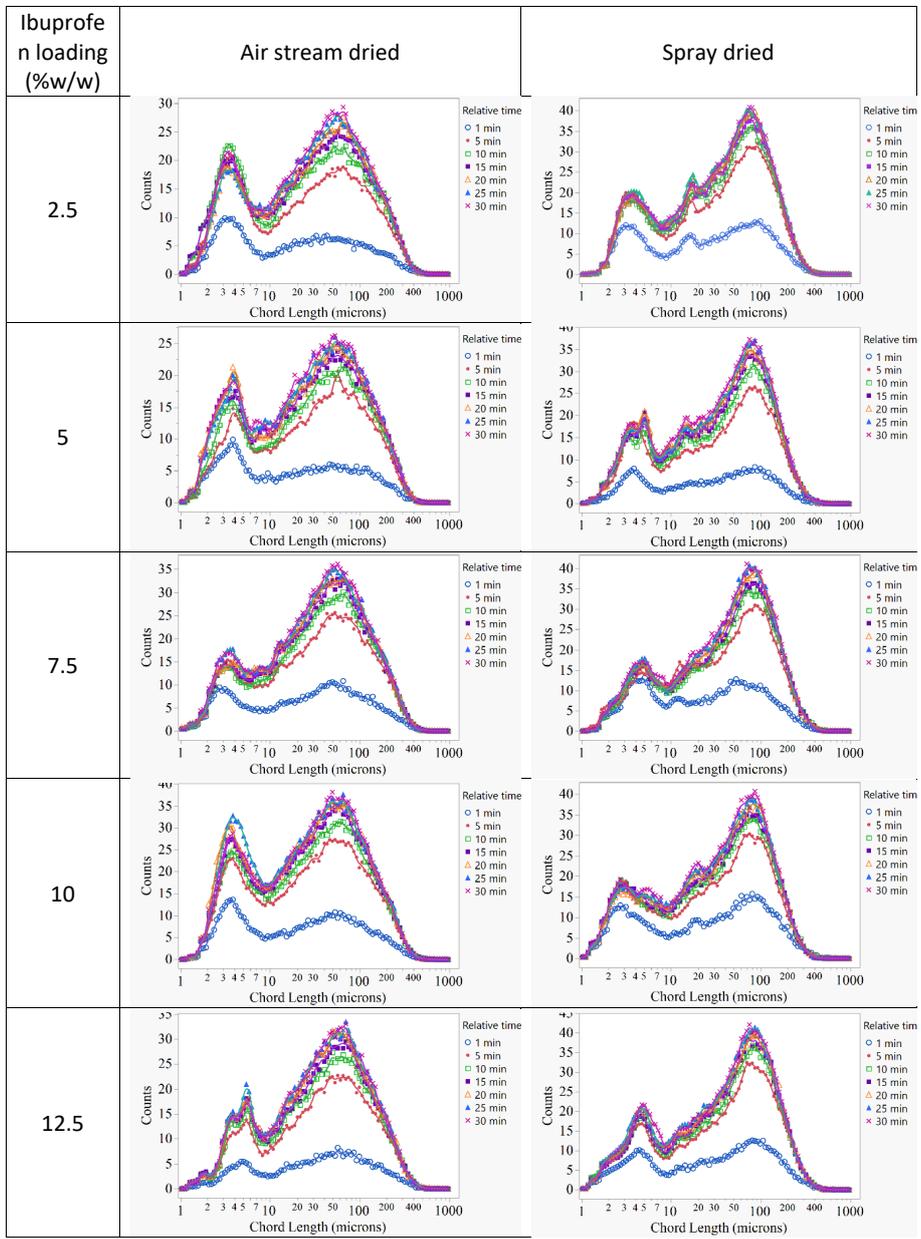
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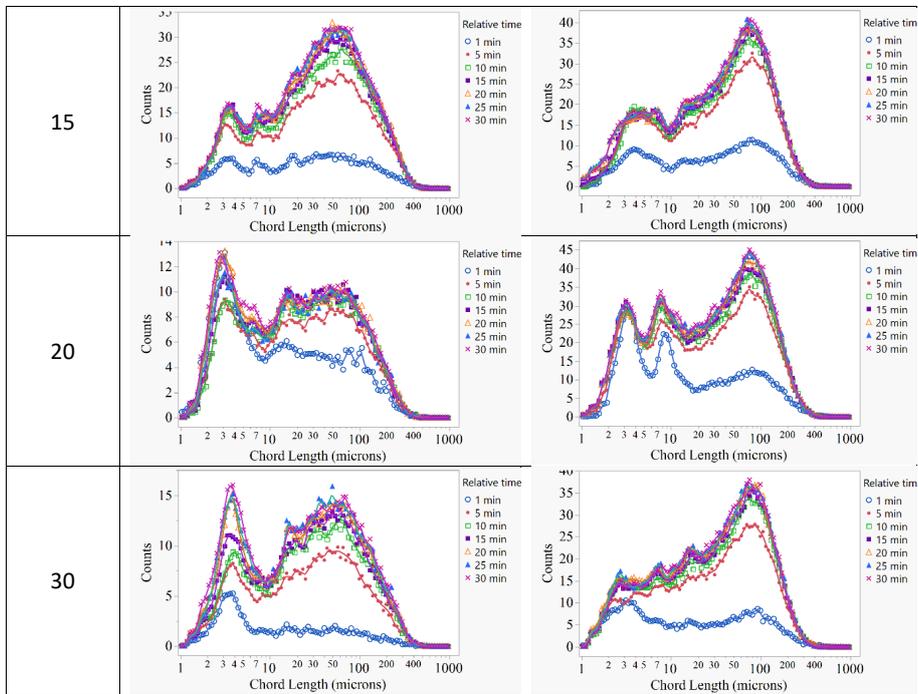




789 Figure SM-1. Dissolution profiles of tablets containing spray dried MCC (Emcocel®) (solid
 790 line) and air stream dried MCC (Vivapur®) (dashed line) containing ibuprofen (a) 2.5 %, (b) 5
 791 %, (c) 7.5 %, (d) 10 %, (e) 12.5 %, (f) 15 %, (g) 20 %, and (h) 30 % w/w. Dashed and solid lines
 792 correspond to air stream and spray dried MCCs, respectively. Dissolution was performed in
 793 phosphate buffer pH 7.2 at 37°C. Average values shown with y-error bars indicating
 794 standard deviation, n= 5.

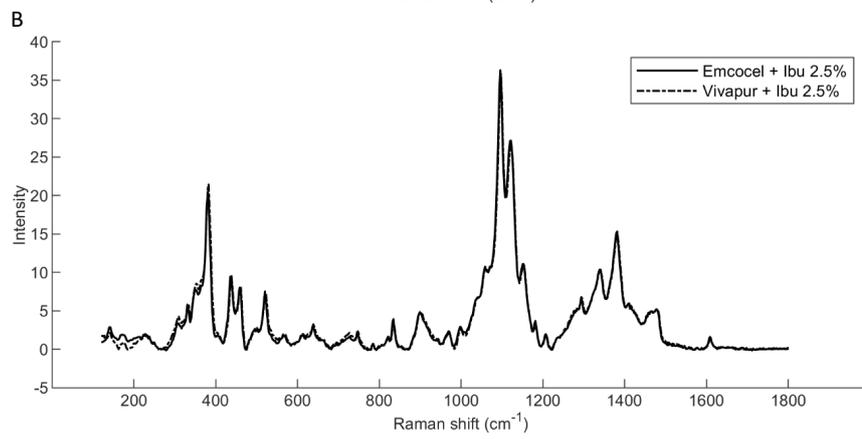
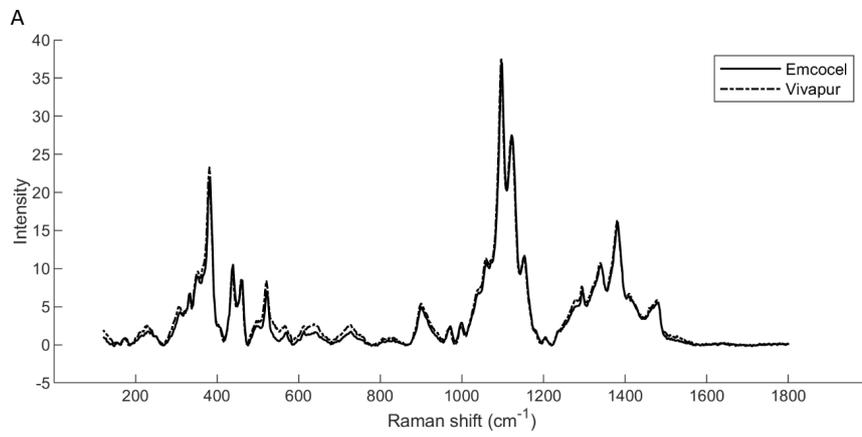
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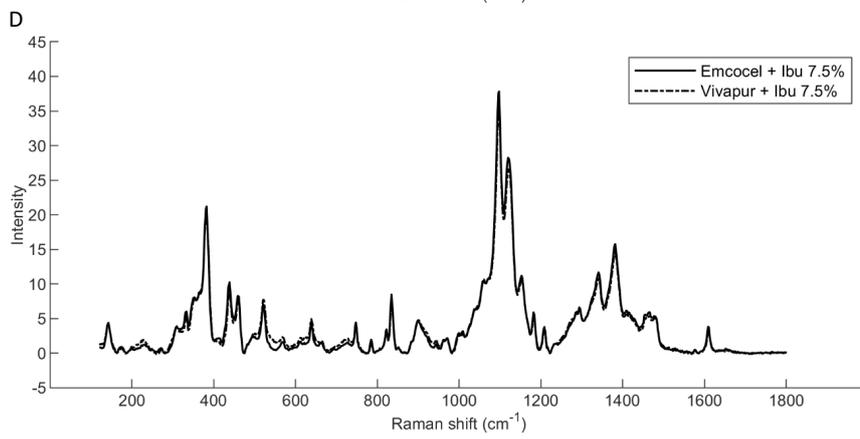
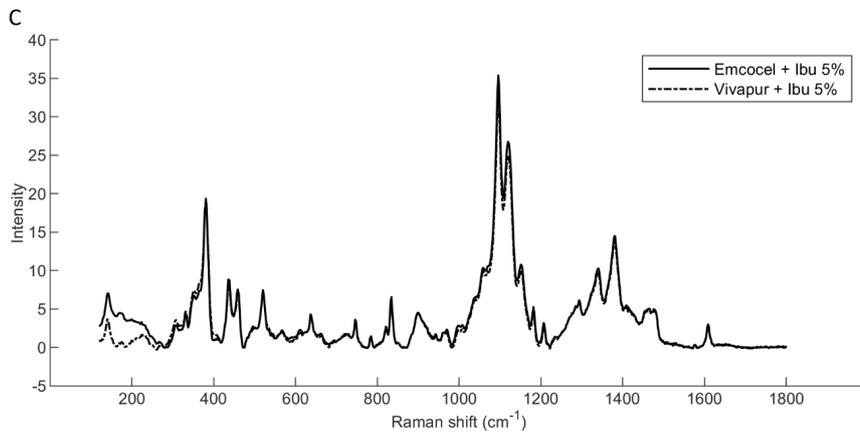


796 Figure SM-2. Focused Beam Reflectance Measurement (FBRM) chord length distributions for
 797 tablets with ibuprofen loadings of 2.5 % - 30 % w/w spray dried MCC (Emcocel®) and air
 798 stream dried MCC (Vivapur®), respectively, at different time points after addition to the
 799 disintegration medium, phosphate buffer pH 7.2 and temperature of 37 °C.

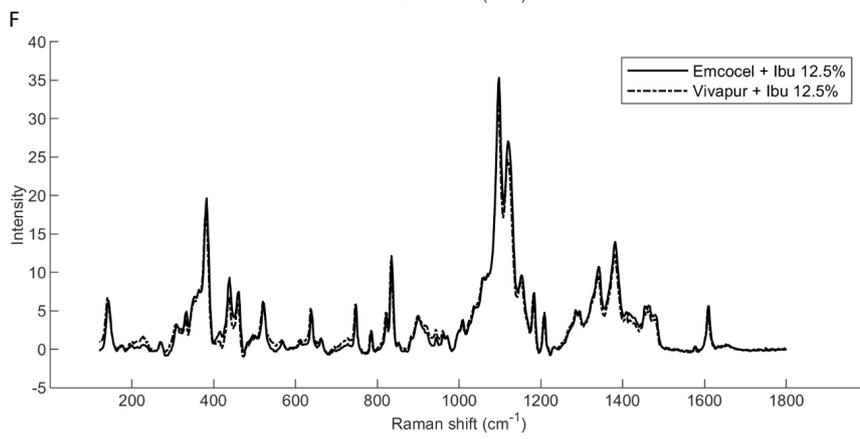
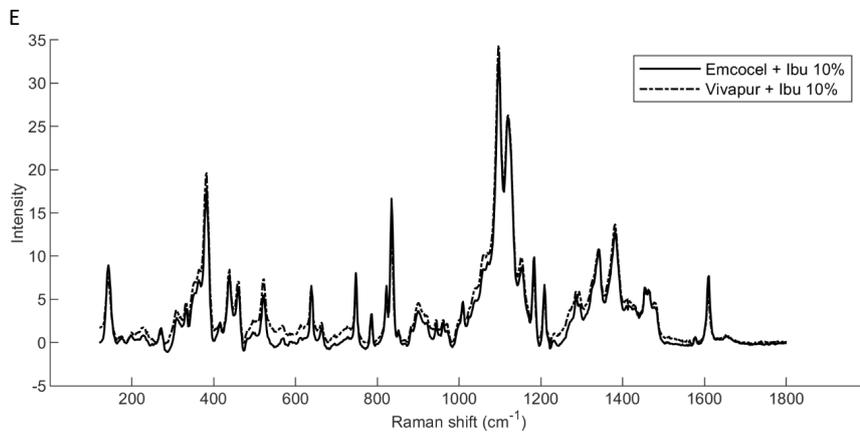
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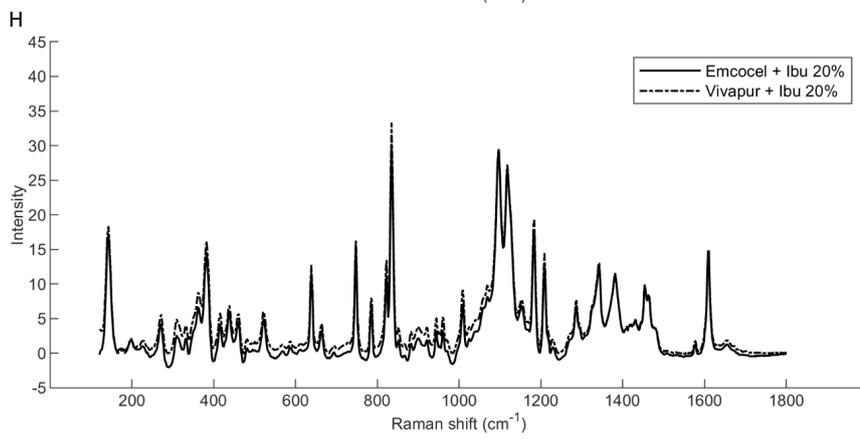
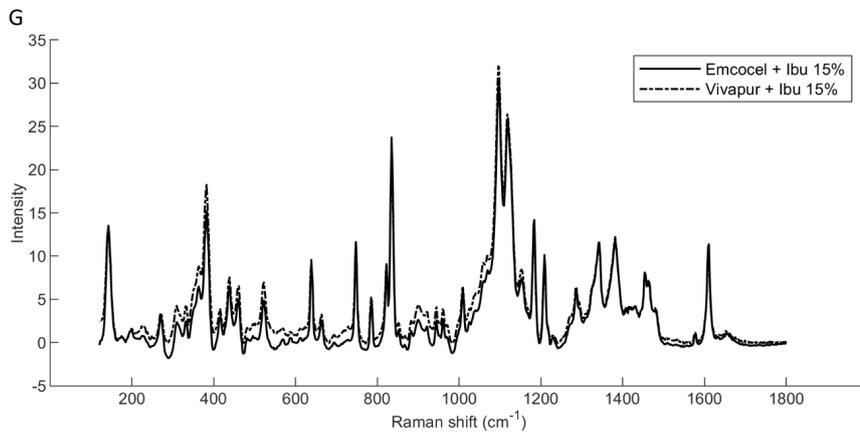
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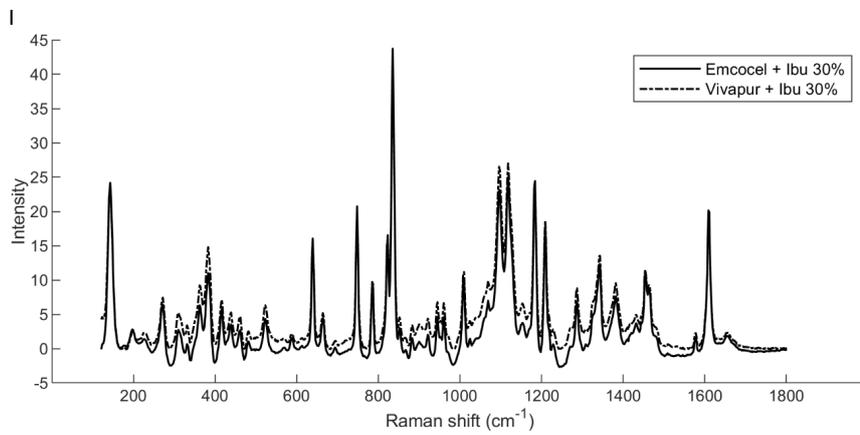
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805 Figure SM-3. Raman spectra of tablet of (A) Spray dried MCC (Emcocel®) and Air stream
806 dried MCC (Vivapur®) and of the blends with ibuprofen at (B) 2.5%, (C) 5%, (D) 7.5%, (E)
807 10%, (F) 12.5%, (G) 15%, (H) 20%, (I) 30% w/w ibuprofen/MCC.

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