

Title	Draft genome sequence of <i>Bacillus thuringiensis</i> DPC6431, producer of the bacteriocin thuricin CD
Authors	Arbulu, Sara;Murphy, Kiera;O'Sullivan, Orla;Rea, Mary C.;Hill, Colin;Ross, R. Paul
Publication date	2019-08-15
Original Citation	Arbulu, S., Murphy, K., O'Sullivan, O., Rea, M. C., Hill, C. and Ross, R. P. (2019) 'Draft Genome Sequence of <i>Bacillus thuringiensis</i> DPC6431, Producer of the Bacteriocin Thuricin CD', <i>Microbiology Resource Announcements</i> , 8(33), e00398-19 (2pp.) DOI:10.1128/MRA.00398-19
Type of publication	Article (peer-reviewed)
Link to publisher's version	https://mra.asm.org/content/8/33/e00398-19 - 10.1128/MRA.00398-19
Rights	©2019 Arbulu et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license. - https://creativecommons.org/licenses/by/4.0/
Download date	2024-10-07 17:08:30
Item downloaded from	https://hdl.handle.net/10468/8715




UCC

University College Cork, Ireland
Coláiste na hOllscoile Corcaigh



Draft Genome Sequence of *Bacillus thuringiensis* DPC6431, Producer of the Bacteriocin Thuricin CD

 Sara Arbulu,^{a,b} Kiera Murphy,^{a,b} Orla O'Sullivan,^{a,b} Mary C. Rea,^{a,b} Colin Hill,^{b,c} R. Paul Ross^{a,b,c}

^aFood Biosciences Department, Teagasc Food Research Centre, Fermoy, County Cork, Ireland

^bAPC Microbiome Ireland, University College Cork, Cork, Ireland

^cSchool of Microbiology, University College Cork, Cork, Ireland

ABSTRACT We report the draft genome sequence of *Bacillus thuringiensis* DPC6431, a producer of the anticlostridial bacteriocin thuricin CD and isolated from a human fecal sample. The assembly comprises 96 contigs for a total of 5,581,839 bp, with 32.5% G+C content.

Bacillus spp. are ubiquitous Gram-positive and spore-forming bacteria known to produce ribosomally and nonribosomally synthesized antimicrobials, as well as enzymes and other metabolites with biotechnological potential (1, 2). *Bacillus thuringiensis* DPC6431 was isolated from a human fecal sample and produces the two-component sactibiotic thuricin CD, a narrow-spectrum bacteriocin which is particularly active against *Clostridium difficile* (3). In order to isolate the strain, stool samples were diluted in anaerobic diluent and spread on Wilkins-Chalgren anaerobic plates (Oxoid). Colonies were overlaid with *Clostridium* agar (Oxoid) inoculated with *C. difficile* ATCC 43593. Inhibition halos led to the detection of *B. thuringiensis* DPC6431 that was further purified and grown in brain heart infusion broth (BHI; Oxoid). According to biochemical tests with the API 50CHB and 20E kits (bioMérieux), the isolate was classified as a *Bacillus* sp. Further sequencing of the 16S rRNA and *gyrB* genes determined the isolate to belong to *B. thuringiensis*. Thuricin CD was purified from cell-free supernatants of BHI broth *B. thuringiensis* DPC6431 cultures, and molecular mass and structure were confirmed by matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry and nuclear magnetic resonance (NMR), respectively. Genetic sequencing of the thuricin CD operon was also performed (3). Thuricin CD has shown an effect on *C. difficile* similar to that of traditional antibiotics, and it is active against biofilm formation (4, 5). Moreover, it displayed minimal impact on a human gut microbiota research model (6), and preliminary trials for oral and rectal administration of thuricin CD in mice and pigs, respectively, have also been conducted (7).

The strain is designated *B. thuringiensis* DPC6431 (from the Dairy Products Centre Culture Collection) and APC20 (from the APC Microbiome Ireland culture collection). The strain is also deposited with the National Collection of Industrial and Marine Bacteria (accession number 41490).

B. thuringiensis DPC6431 was grown overnight on agar plates BHI at 37°C from the glycerol –80°C stocks and then transferred to BHI broth for DNA extraction. The genomic DNA of *B. thuringiensis* DPC6431 was isolated using a DNeasy UltraClean microbial kit (Qiagen) and assessed using a NanoDrop ND-1000 spectrophotometer and by electrophoresis. Genomic DNA libraries were prepared using the Nextera XT library prep kit (Illumina, San Diego, CA, USA), following the manufacturer's protocol but with the following modifications: 2 ng of DNA instead of 1 ng was used as input, and the PCR elongation time was increased to 1 min from 30 s. DNA quantification and library preparation were carried out on a Hamilton Microlab Star automated liquid-handling system. Pooled libraries were

Citation Arbulu S, Murphy K, O'Sullivan O, Rea MC, Hill C, Ross RP. 2019. Draft genome sequence of *Bacillus thuringiensis* DPC6431, producer of the bacteriocin thuricin CD. *Microbiol Resour Announc* 8:e00398-19. <https://doi.org/10.1128/MRA.00398-19>.

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2019 Arbulu et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to R. Paul Ross, p.ross@ucc.ie.

Received 25 April 2019

Accepted 21 July 2019

Published 15 August 2019

quantified using the Kapa Biosystems library quantification kit for Illumina on a Roche LightCycler 96 quantitative PCR (qPCR) machine. Libraries were sequenced on an Illumina HiSeq platform using a 250-bp paired-end protocol. Reads were adapter trimmed using Trimmomatic 0.30 with a sliding window quality cutoff of Q15 (8). *De novo* assembly was performed on samples using SPAdes version 3.7 (9), and contigs were annotated using the RAST server (10–12). Both platforms were used on default mode.

The whole-genome sequencing was performed at 30× coverage, yielding a total of 1,824,237 reads after trimming and quality filtering. The draft genome of *B. thuringiensis* DPC6431 consists of 96 contigs with a contig N_{50} length of 304,462 bp, a total of 5,581,839 bp, and a G+C content of 32.5%. The largest contig was 756,911 bp, and the smallest contig was 255 bp. The median insert size was 855 bases. The total number of coding sequences was 5,894, and the number of RNAs was 107.

In silico analysis using the BAGEL4 Web server (13) confirmed the presence of a thuricin CD biosynthetic cluster and revealed the presence of cerein B structural, transporter, regulation, and immunity genes, as well as other areas of interest related to linear azol(in)e-containing peptides (LAPs). Two potential clustered regularly interspaced short palindromic repeat (CRISPR) arrays involved in antiviral defense mechanisms were identified using CRISPRFinder (14). The public availability of this draft genome will strengthen the value of the strain as a bacteriocin producer and allow for deeper analysis for future biotechnological applications.

Data availability. This draft genome sequence has been deposited at DDBJ/ENA/GenBank under the accession number [SCLP00000000](https://doi.org/10.1093/bioinformatics/btu170). The version described in this paper is version SCLP01000000. Raw sequencing data have been deposited in the SRA database under accession number [PRJNA515439](https://doi.org/10.1093/bioinformatics/btu170).

ACKNOWLEDGMENTS

This work was supported by Science Foundation Ireland (SFI) under grant number SFI/12/RC/2273.

Genome sequencing was provided by MicrobesNG (<https://microbesng.uk/>), which is supported by the BBSRC (grant number BB/L024209/1).

REFERENCES

- Elshaghabee FMF, Rokana N, Gulhane RD, Sharma C, Panwar H. 2017. *Bacillus* as potential probiotics: status, concerns, and future perspectives. *Front Microbiol* 8:1490. <https://doi.org/10.3389/fmicb.2017.01490>.
- Melo AL, Soccol VT, Soccol CR. 2016. *Bacillus thuringiensis*: mechanism of action, resistance, and new applications: a review. *Crit Rev Biotechnol* 36:317–326. <https://doi.org/10.3109/07388551.2014.960793>.
- Rea MC, Sit CS, Clayton E, O'Connor PM, Whittall RM, Zheng J, Vederas JC, Ross RP, Hill C. 2010. Thuricin CD, a posttranslationally modified bacteriocin with a narrow spectrum of activity against *Clostridium difficile*. *Proc Natl Acad Sci U S A* 107:9352–9357. <https://doi.org/10.1073/pnas.0913554107>.
- Mathur H, O'Connor PM, Hill C, Cotter PD, Ross RP. 2013. Analysis of anti-*Clostridium difficile* activity of thuricin CD, vancomycin, metronidazole, ramoplanin, and actagardine, both singly and in paired combinations. *Antimicrob Agents Chemother* 57:2882–2886. <https://doi.org/10.1128/AAC.00261-13>.
- Mathur H, Rea MC, Cotter PD, Hill C, Ross RP. 2016. The efficacy of thuricin CD, tigecycline, vancomycin, teicoplanin, rifampicin and nitazoxanide, independently and in paired combinations against *Clostridium difficile* biofilms and planktonic cells. *Gut Pathog* 8:20. <https://doi.org/10.1186/s13099-016-0102-8>.
- Rea MC, Dobson A, O'Sullivan O, Crispie F, Fouhy F, Cotter PD, Shanahan F, Kiely B, Hill C, Ross RP. 2011. Effect of broad- and narrow-spectrum antimicrobials on *Clostridium difficile* and microbial diversity in a model of the distal colon. *Proc Natl Acad Sci U S A* 108:4639–4644. <https://doi.org/10.1073/pnas.1001224107>.
- Rea MC, Alemayehu D, Casey PG, O'Connor PM, Lawlor PG, Walsh M, Shanahan F, Kiely B, Ross RP, Hill C. 2014. Bioavailability of the anti-clostridial bacteriocin thuricin CD in gastrointestinal tract. *Microbiology* 160:439–445. <https://doi.org/10.1099/mic.0.068767-0>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- van Heel AJ, de Jong A, Song C, Viel JH, Kok J, Kuipers OP. 2018. BAGEL4: a user-friendly Web server to thoroughly mine RiPPs and bacteriocins. *Nucleic Acids Res* 46:W278–W281. <https://doi.org/10.1093/nar/gky383>.
- Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <https://doi.org/10.1093/nar/gkm360>.